

1     **The adenylate cyclase activator forskolin potentiates the positive inotropic effect of the**  
2     **phosphodiesterase inhibitor milrinone but not of the calcium sensitizer levosimendan**  
3     **nor of its hemodynamically active metabolites: an apparent conundrum.**

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5     Jouko Levijoki, MSc,<sup>\*</sup> Piero Pollesello, PhD,<sup>\*</sup> Elena Grossini, PhD,<sup>†</sup> Zoltán Papp, MD, PhD<sup>‡§</sup>

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7     <sup>\*</sup>Critical Care, Orion Pharma, Espoo, Finland; <sup>†</sup>Laboratory of Physiology, Department of  
8     Translational Medicine, University East Piedmont, Novara, Italy; <sup>‡</sup>Division of Clinical  
9     Physiology, Department of Cardiology, Faculty of Medicine, Hungary; <sup>§</sup>HAS-UD Vascular  
10    Biology and Myocardial Pathophysiology Research Group, Hungarian Academy of Sciences,  
11    Budapest, Hungary

12  
13    CORRESPONDING AUTHOR: Piero Pollesello, PhD, Adj. Prof., FESC, FHFA, Orion  
14    Pharma, P.O. Box 65, 02101 Espoo, Finland; tel. +358509664191; e-mail:  
15    piero.pollesello@orionpharma.com

## 16 ABSTRACT

17 OR-1855 and OR-1896 are two hemodynamically active metabolites of the inodilator  
18 levosimendan, with calcium sensitizing activity, but their mechanism of action is still not fully  
19 understood.

20 It has been previously reported that the positive inotropic effect of levosimendan is  
21 not potentiated by the adenylate cyclase activator forskolin whereas forskolin does potentiate the  
22 effects of the phosphodiesterase (PDE) inhibitor milrinone.

23 To ascertain whether the active metabolites follow the same pattern of levosimendan,  
24 the positive inotropic effects of OR- 1855 and OR-1896, were studied in guinea-pig isolated  
25 papillary muscle in the presence and absence of forskolin. OR-1855 and OR-1896 were also  
26 tested as inhibitors of PDE-III and PDE-IV.

27 Our result show that 0.1  $\mu$ M forskolin did not potentiate the positive inotropic effect  
28 of either OR-1855 or OR-1896, as in the case of the parent compound levosimendan. As in  
29 previous studies, the positive inotropic effect of milrinone was markedly potentiated in the  
30 presence of forskolin.

31 From these data we propose an explanation for the divergent behaviour of the calcium  
32 sensitizing drugs and PDE inhibitors.

33

34 **KEYWORDS:** levosimendan, inotropy, phosphodiesterase inhibitor, mechanism of action,  
35 cAMP

## 36 1. INTRODUCTION

37 Levosimendan is an i.v. inodilator used in acutely decompensated heart failure,<sup>1</sup> in  
38 perioperative settings,<sup>2</sup> and in intensive care.<sup>3</sup> Levosimendan has a triple mechanism of  
39 action<sup>4</sup> with clinical effects including an increase in output and cardiac index and in an  
40 improvement of both systemic and pulmonary venous congestion.<sup>5</sup> The inotropic effect of  
41 levosimendan is driven by calcium sensitization of the contractile apparatus via a selective  
42 binding of levosimendan on the N-terminal of the cardiac isoform of troponin C.<sup>6</sup>  
43 Levosimendan, however, is also a potent phosphodiesterase (PDE) inhibitor with a uniquely  
44 high selectivity for PDE-III relative to PDE-IV.<sup>7</sup> There has been lengthy debate about whether  
45 the PDE inhibitory property of levosimendan plays a role in the inotropic effects and overall  
46 in the clinical effects of the drug and two lines of thought have been developed: (1) the  
47 presence of both PDE-III and PDE-IV in cardiomyocytes implies the existence of parallel  
48 cyclic adenosine monophosphate (cAMP) decyclization pathways. This redundancy would  
49 make a highly selective PDE-III inhibitor such as levosimendan unable to increase cAMP  
50 since the PDE-IV path would still be operational;<sup>8</sup> or (2) PDE inhibition by levosimendan  
51 does increase cAMP levels sufficiently to create a synergy with the calcium sensitizing effect  
52 but not enough to affect the contractile apparatus per se.<sup>9</sup>

53 In clinical settings, it has been shown that the hemodynamic effects of  
54 levosimendan are prolonged due to the formation of two hemodynamically active plasma  
55 metabolites, OR-1855 and OR-1896 (see their chemical structures in the supplementary  
56 material).<sup>10</sup> The pharmacokinetics and pharmacodynamics characteristics of those metabolites  
57 has been described in details<sup>11</sup> and their role in the clinical effects of levosimendan treatment  
58 has been discussed abundantly.<sup>12</sup>

59 OR-1896 exerts a positive inotropy effect in *ex vivo* models<sup>13</sup> and inhibits PDE-III  
60 selectively in purified enzyme preparations.<sup>14</sup> In our present research, we sought to shed

61 further light on the mechanism of action of both metabolites and understand which effect(s)  
62 underpin their inotropic properties.

63 In a previous report<sup>15</sup> the positive inotropic effect of the parent compound  
64 levosimendan, seen as increase of contraction force, was not potentiated by forskolin, a  
65 labdane diterpene derived from geranylgeranyl pyrophosphate commonly used to increase the  
66 levels of cAMP by stimulation of adenylate cyclase,<sup>16,17</sup> whereas forskolin did potentiate the  
67 effects of the classic PDE inhibitor, milrinone, which has inhibitory effects on both PDE-III  
68 and PDE-IV.<sup>18</sup>

69 The aim of this study was to investigate whether the positive inotropic effects of  
70 OR-1855 and OR-1896 follow the behaviour of the parent compound levosimendan and are  
71 not potentiated by forskolin, or behave as the PDE inhibitor, milrinone.

72

## 73 2. MATERIALS AND METHODS

### 74 2.1 Chemicals

75 The compound used were levosimendan, batch LS, Orion Pharma; OR-1855, batch LS, Orion  
76 Pharma; OR-1896, batch L7, Orion Pharma; Milrinone, batch LS, Orion Pharma; Forskolin,  
77 Lot B25975, Calbiochem-Novabiochem Corp, La Jolla, CA, USA. All the test compounds  
78 were dissolved in dimethyl sulfoxide (DMSO). Stock solutions were diluted so that the final  
79 DMSO concentration was 0.4% throughout the experiment.

80

### 81 2.2. Phosphodiesterase inhibition

82 Highly purified PDE-III and PDE-IV isozymes were isolated from human platelets and a  
83 promonocytic cell line of patients with myeloid leukaemia (U-937), respectively, according to  
84 published methods.<sup>19</sup> In brief, the supernatant fraction of the tissue homogenate was added to  
85 a diethylaminoethanol-sepharose column and then eluted with a linear sodium acetate gradient

86 buffer. Collected fractions with peak PDE activities were analyzed for cAMP PDE activity.  
87 Purified PDE isozymes were incubated at 30°C for 30 min in a reaction mixture containing  
88 [3H]-cAMP (0.1 μM) and cAMP (0.1 μM) in the presence or absence of the test compounds.  
89 The amount of [3H]-5'-AMP regarded as a degradation product, was determined by using  
90 liquid scintillation detection as described previously.<sup>20</sup> Inhibitory assays were performed in  
91 duplicates.

92

### 93 2.3. Animals

94 The present study was performed in accordance with the guidelines of the Council of Europe  
95 and the US National Research Council. Approval was granted by the Animal Ethics  
96 Committees of Orion Pharma, Finland. Adult guinea-pigs of either sex (Dunkin Hartley,  
97 purchased from Mollegaard Breeding Center LTD., Denmark, or Crl:(Charles River,  
98 Germany), weighing 300- 400 g were used. Guinea-pigs were housed at 20 ± 1 °C with  
99 relative humidity of 50 ± 10%. Light-dark cycle was adjusted with lights on from 06.00 to  
100 20.00h. The guinea-pigs were kept on a standard guinea-pig diet (Altromin 3120) and tap  
101 water ad libitum.

102

### 103 2.4. Papillary muscle preparations

104 Guinea-pigs were killed by a blow on the skull and the heart was excised. Right ventricular  
105 papillary muscle was dissected and rinsed in ice-cold Tyrode solution. Thereafter the papillary  
106 muscle was mounted for the measurement of isometric force in organ baths containing  
107 modified Tyrode solution (37°C) bubbled with carbogen (95% O<sub>2</sub>, 5% CO<sub>2</sub>). The composition  
108 of the Tyrode solution was 135 mM NaCl, 1 mM MgCl<sub>2</sub>×6H<sub>2</sub>O, 5 mM KCl, 2 mM  
109 CaCl<sub>2</sub>×2H<sub>2</sub>O, 15 mM NaHCO<sub>3</sub>, 1 mM Na<sub>2</sub>HPO<sub>4</sub>×2H<sub>2</sub>O, 10 mM glucose, at pH 7.35±0.05.  
110 The volume of the open horizontal chamber was 1 ml and the flow rate of the bathing solution

111 flowing through the chamber was 5 ml/min. The papillary muscle (<1 mm in diameter) was  
112 stretched horizontally between a force-displacement transducer (FT 0.3 C) and a needle fixed  
113 to the bottom of the chamber. The papillary muscle was electrically stimulated (Stimulator  
114 model S 48 F, Grass Instruments) at 1 Hz with rectangular pulses. The pulse duration was 4  
115 ms. The stimulation strength was twice the threshold voltage.

116

## 117 2.5. Experimental procedure

118 After a stabilisation period of 60 min, 0.1  $\mu$ M forskolin was added to the bathing solution (no  
119 addition in control experiments). After a further period of 30 min, a test compound  
120 (levosimendan, OR-1855, OR-1896, or milrinone) was added to the bathing solution at a  
121 starting concentration of 0.03  $\mu$ M. (see an example of trace in **Figure 1**). Thereafter, the  
122 concentration of the test compounds was increased to 0.1, 0.3, 1, 3, 10, and 30  $\mu$ M at 10 min  
123 intervals. The highest two concentrations were not tested for levosimendan, for solubility  
124 reasons (see the dosing schedules in **Figure 2**). All the experiments were carried out at 37 °C.  
125 The baselines values were measured at time “0” (as in Figure 2), i.e., immediately before the  
126 first addition of the test compounds. The baseline in experiments with forskolin were thus  
127 measured 30 minutes after the addition of 0.1  $\mu$ M forskolin, and immediately before the first  
128 addition of the test compounds. We selected the concentration of forskolin based on a  
129 previous study on the positive inotropic action of the drug by Metzger H and Lindner E.,<sup>21</sup>  
130 assuming that the induced cAMP activation would be maintained from the beginning to the  
131 end of the papillary muscle contraction experiments as described previously.<sup>22</sup> The increase  
132 of contraction force from the baseline during the up-titration of every test compound was  
133 measured and analysed.

134

## 135 2.6. Statistics

136 Results obtained from five to nine experiments were combined and expressed as mean  $\pm$   
137 SEM. Differences between and within test groups were analysed by using Two Way Repeated  
138 Measures ANOVA followed by the Šídák test (Prism 9.1.0, GraphPad, CA, USA). A p-value  
139  $<0.05$  was considered statistically significant.

140

### 141 3. RESULTS

142 The baseline contraction force values of the guinea-pig papillary muscle prepares are shown  
143 in **Table 1**. No significant differences are seen between the experiments with forskolin and  
144 without forskolin.

145 OR-1855 and OR-1896 increased the inotropy of guinea-pig papillary muscle from  
146 baseline by maximum values of  $312 \pm 118$  mg (n=5) and  $341 \pm 82$  mg (n=8), respectively  
147 (**Figure 3**). The presence of forskolin  $0.1 \mu\text{M}$ , did not potentiate significantly the positive  
148 inotropic effect of either compound: the maximum increases in contraction force in presence  
149 of the adenylate cyclase stimulant were  $265 \pm 62$  mg (n=5) for OR-1855 and  $334 \pm 31$  mg  
150 (n=6) for OR-1896.

151 For levosimendan, the maximum increases in contraction were  $331 \pm 58$  mg (n=5)  
152 and  $393 \pm 69$  mg (n=6) in the absence and presence of forskolin, respectively (n.s.). The  
153 maximal force increase from baseline with milrinone in the absence of forskolin was  $219 \pm 42$   
154 mg (n=5). That effect was significantly potentiated by forskolin ( $393 \pm 69$  mg; n=6)  
155 ( $p < 0.005$ ).

156 The  $\text{IC}_{50}$  for PDE-III and PDE-IV were calculated from the relevant dose-  
157 dependent inhibition curves of the four compounds (see **Table 2**). The PDE-III to PDE-IV  
158  $\text{IC}_{50}$  ratio was also calculated; those values, reflecting selectivity of inhibition of PDE-III are  
159 7619 for levosimendan, 3043 for OR-1986, 100 for OR-1855, and 39 for milrinone.

160

## 161 4. DISCUSSION

162 The adenylyate cyclase activator forskolin increases intracellular cAMP level and thereby  
163 stimulates cAMP-dependent protein kinase A, which in turn increases calcium current<sup>23</sup> and  
164 enhances contraction force. On the other hand, the positive inotropic effect of some PDE  
165 inhibitor is potentiated by forskolin as previously demonstrated for instance with milrinone.<sup>15</sup>  
166 The two major plasma metabolites of levosimendan, OR-1855 and OR-1896 are thought to  
167 exert a positive inotropic activity by calcium sensitization of troponin C in the cardiomyocyte  
168 contractile apparatus. Nevertheless, these metabolites also inhibit the PDE-III isozyme in a  
169 highly selective manner in purified enzyme preparations.

170 One can hypothesize that the combination of the three aforementioned mechanisms  
171 (activation of adenylyate cyclase, inhibition of phosphodiesterase, and calcium sensitization)  
172 would lead to an increased positive inotropy. Moreover, if these pathways are independent  
173 than their effects will be strictly additive. If there were to be any overlap (and hence non-  
174 additive effect on inotropy) between these mechanisms it might most likely arise between the  
175 two (activation of adenylyate cyclase and inhibition of phosphodiesterase) sharing a common  
176 factor, *i.e.*, cAMP.

177 Our findings that the positive inotropic effects of OR-1855 and OR-1896 were not  
178 additive to the effects of forskolin (as was also the similarly as in the case or of their parent  
179 compound levosimendan) while whereas the effect of milrinone was, would appear to diverge  
180 from the above hypothesis.

181 One possible reason for the lack of contribution of adenylyate cyclase stimulus to  
182 the inotropic effects of levosimendan and its metabolites is their high selectivity in PDE-III  
183 inhibition over the PDE-IV isoenzyme. It has been suggested that both PDE-III and PDE-IV  
184 should be inhibited to high levels in order to increase the amplitude of the intracellular  
185 calcium transient,<sup>24</sup> because an uninhibited PDE isozyme (*i.e.*, PDE-IV in this case) can



186 potentially offset any effect from the inhibition of the other isoform (i.e., PDE-III). In keeping  
187 with this proposition, milrinone, which inhibits both isoenzymes, was potentiated by forskolin  
188 in our experiments. Accordingly, the effect of milrinone on intracellular cAMP and calcium  
189 concentrations is more prevalent than that for levosimendan.<sup>15</sup>

190 It is also to note that in previous studies, that levosimendan induced NO  
191 production but that co-stimulation with cilostazol (another PDE-III inhibitor) failed to  
192 potentiate the effects of levosimendan on NO release in coronary endothelial cells.<sup>25</sup> This also  
193 speaks to a selective inhibition of PDE-III by levosimendan.

194

## 195 5. CONCLUSION

196 Like their parent compound levosimendan, the metabolites OR-1855 and OR-1896 have a  
197 positive inotropy effect which is not potentiated by forskolin. Conversely, the inotropic effect  
198 of the PDE-III/PDE-IV inhibitor milrinone is potentiated by adenylate cyclase activation. This  
199 different behaviour could be explained by the fact that positive inotropic effects evoked by  
200 milrinone or by levosimendan and its active metabolites are exerted via different mechanisms  
201 of action with different roles for cAMP.

202 The oral formulation of levosimendan is currently under scrutiny as treatment of  
203 pulmonary hypertension associated with heart failure<sup>26</sup> and the role of the active metabolites  
204 is paramount in this new pharmacokinetic/pharmacodynamics situation. Hence, the full  
205 characterization of OR-1855 and OR-1896 mode of action and pharmacology is of the  
206 utmost importance.

207

## 208 6. LIMITATIONS

209 In the study we used papillary muscles with diameter  $\leq 1$  mm. Diameters of individual  
210 preparates varied, however, as did the contraction force of individual samples. This is why we

211 used the increase of force from baseline for our analysis. These preparations have intrinsic  
212 problems, such as a radius-dependent performance and the possibility that the core of the  
213 muscle bundle is hypoxic or even anoxic, However, radius-dependent decline of performance  
214 in isolated cardiac muscle does not reflect inadequacy of diffusive oxygen supply.<sup>27</sup>

215

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219

#### 220 CONTRIBUTION

221 JL and PP designed the research study. JL performed the research. PP contributed essential  
222 reagents or tools. JL, PP, ZP and EG analyzed the data. PP and ZP wrote the paper.

223

#### 224 DISCLOSURES

225 PP and JL are full time employee of Orion Pharma where the inotrope levosimendan was  
226 discovered and developed. Both EG and ZP have received, in the latest 5 years, either  
227 research grants and/or speaker honoraria and/or support for conference attendance from Orion  
228 Pharma.

229

230 **Table 1:** Baseline contraction force values of guinea-pig papillary muscle prepartate

Study compound	without Forskolin			with Forskolin			difference
	Mean, mg	SEM	n	Mean, mg	SEM	n	
Levosimendan	260	47	5	238	32	6	ns
OR-1896	315	51	8	297	34	6	ns
OR-1855	327	38	5	354	58	5	ns
Milrinone	200	12	5	299	30	6	ns

231

232

233 **Table 2:** PDE-III/PDE -IV IC<sub>50</sub> for OR-1896, OR1855, and levosimendan and for milrinone

<b>Compound</b>	<b>PDE-III, μM</b>	<b>PDE-IV, μM</b>	<b>PDE-IV vs PDE-III IC<sub>50</sub> ratio</b>
Levosimendan	0.0021	16	7619
OR-1896	0.094	286	3043
OR-1855	5	500	100
Milrinone	0.45	17.5	39

234

235

236 **LEGENDS TO THE FIGURES**

237

238 **Figure 1**

239 Example of tracing of papillary muscle contraction. The papillary muscle was electrically  
240 stimulated at 1 Hz with rectangular pulses of 4 ms. The stimulation strength was twice the  
241 threshold voltage. After a stabilisation period of 60 min, 0.1  $\mu$ M forskolin was added to the  
242 bathing solution. After a further period of 30 min, milrinone was added to the bathing solution  
243 at increasing concentrations.

244

245 **Figure 2**

246 Dosing schedule. The same color coding is used also in Figure 3.

247

248 **Figure 3**

249 Positive inotropic effect of levosimendan (upper left panel, yellow hexagons), OR-1896  
250 (upper right panel, green dots), OR-1855 (lower left panel, blue triangles) and milrinone  
251 (lower right panel, red squares) in the presence and the absence of forskolin (0.1  $\mu$ M) in  
252 guinea-pig isolated papillary muscle. Shown are mean changes of twitch tension  $\pm$  SEM from  
253 the baseline levels. An asterisk (\*) indicates a statistical significant difference ( $p < 0.05$ ) from  
254 the baseline level. A dagger (†) indicates a statistical significant difference ( $p < 0.05$ ) between  
255 the groups with and without forskolin. Data were analysed for statistical differences using  
256 two-way ANOVA followed by the Šídák test (Prism 9.1.0, GraphPad, CA, USA).

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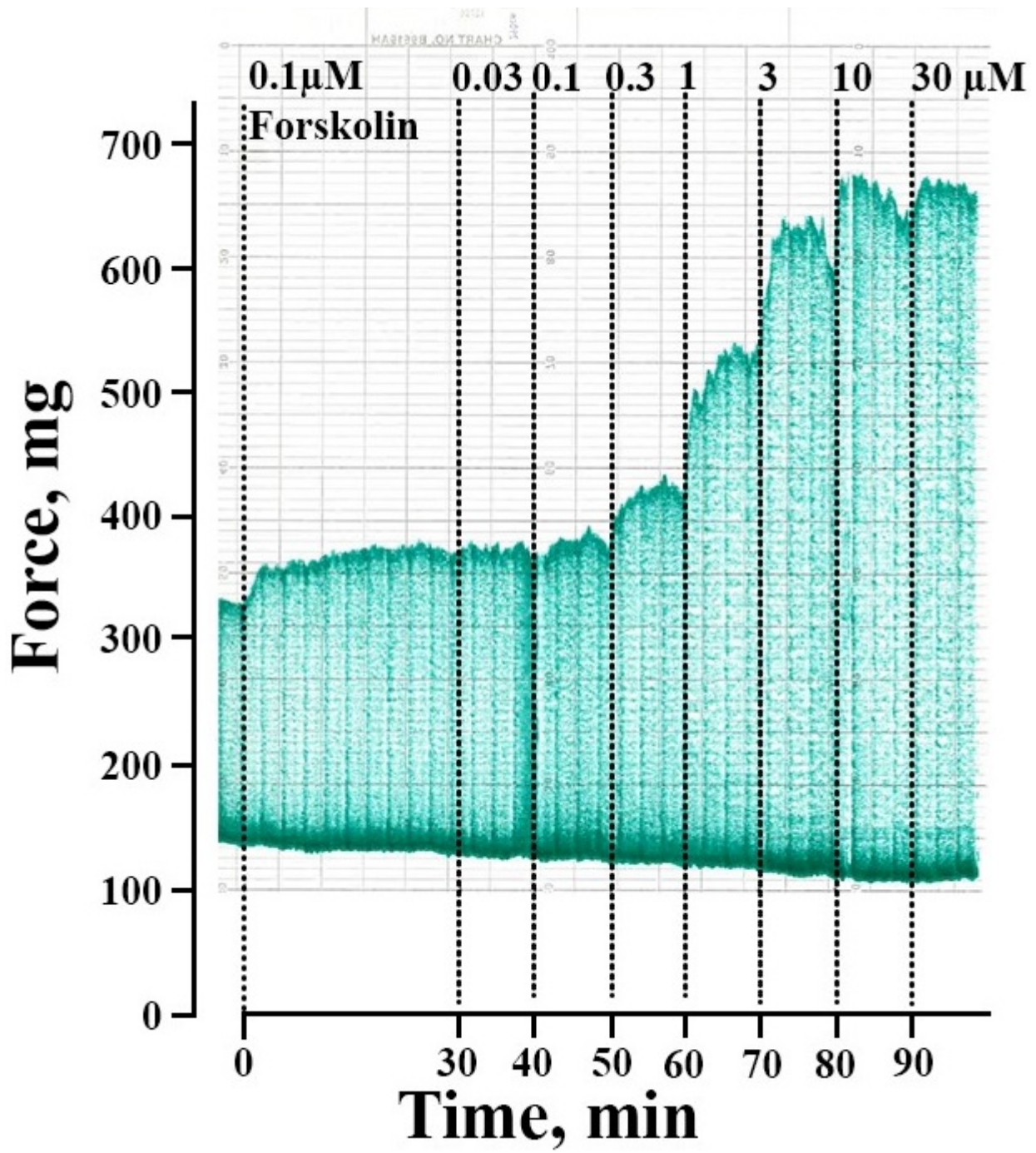


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372 **FIGURE 1**

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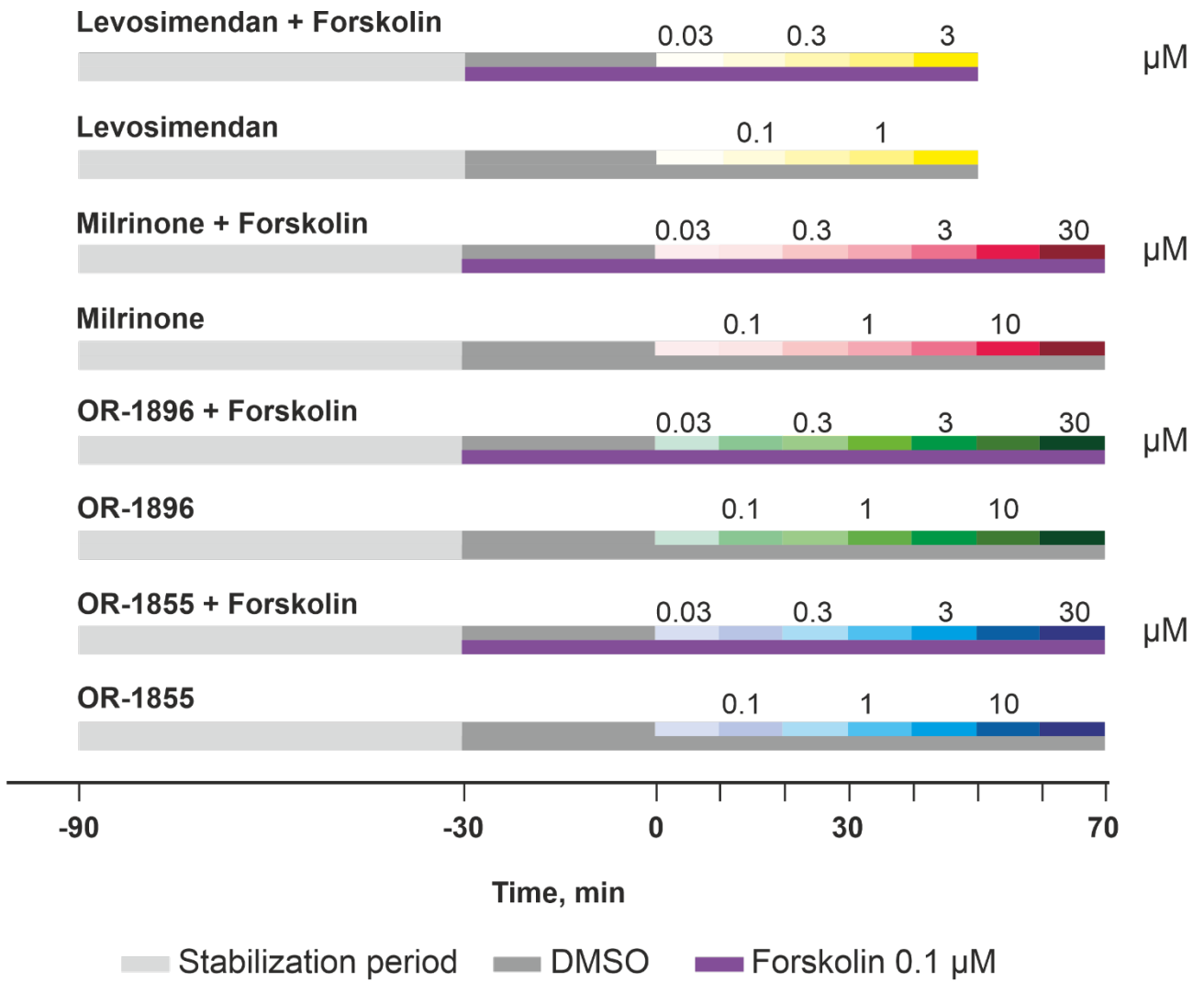


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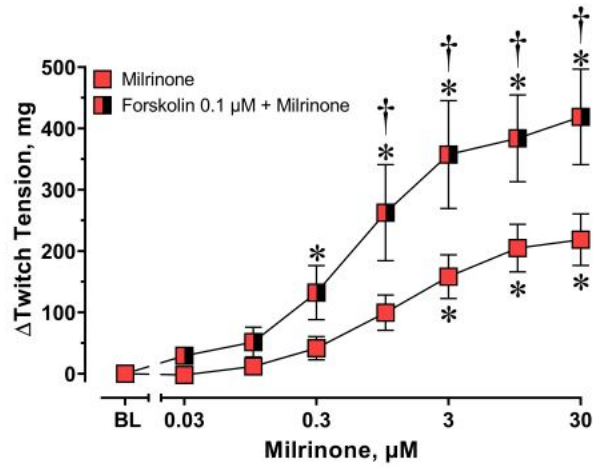
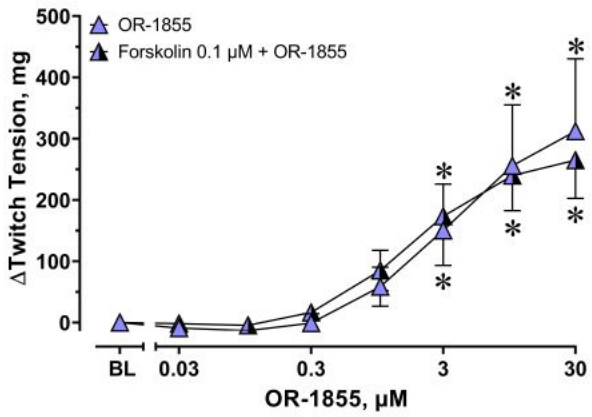
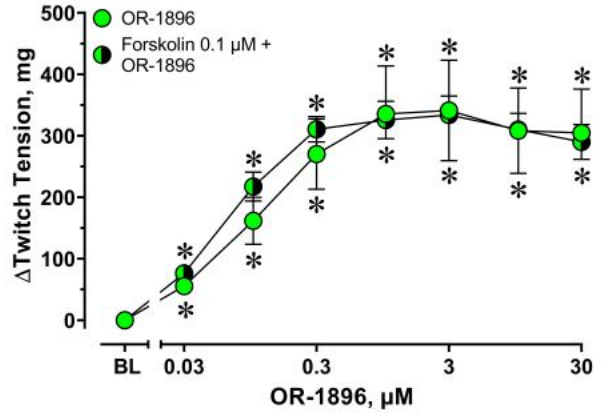
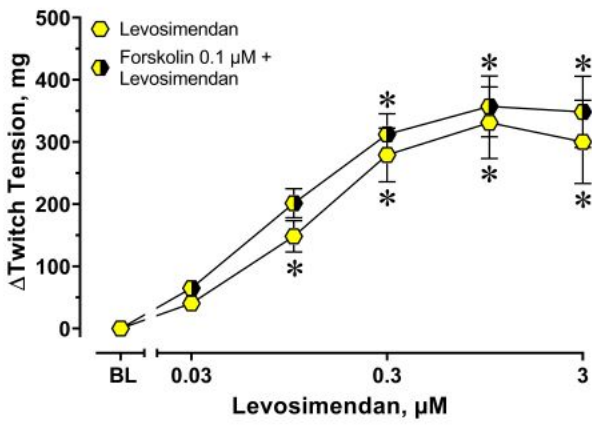
376 **FIGURE 2**

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379 **FIGURE 3**

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## 383 SUPPLEMENTARY MATERIAL

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## 385 CHEMICAL STRUCTURES

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