



Bioactivity and Delivery Strategies of Phytochemical Compounds in Bone Tissue Regeneration

Anna Valentino ^{1,2,†}, Francesca Di Cristo ^{3,†}, Michela Bosetti ¹, Amal Amaghnouje ⁴, Dalila Bousta ⁴, Raffaele Conte ^{5,*} and Anna Calarco ^{2,*}

- ¹ Dipartimento di Scienze del Farmaco, Università del Piemonte Orientale "A. Avogadro", Largo Donegani, 2, 28100 Novara, Italy; anna.valentino@uniupo.it (A.V.); michela.bosetti@uniupo.it (M.B.)
- ² Research Institute on Terrestrial Ecosystems (IRET)-CNR, Via Pietro Castellino 111, 80131 Naples, Italy
- ³ Elleva Pharma S.R.L. via Pietro Castellino 111, 80131 Naples, Italy; francesca.dicristo@ellevapharma.com
 ⁴ Laboratory of Biotechnology Environment Agrifood and Health University of Sidi Mohamed Ben Abdellal
 - Laboratory of Biotechnology, Environment, Agrifood, and Health, University of Sidi Mohamed Ben Abdellah,
- 30000 Fez, Morocco; Amal.amaghnouje@usmba.ac.ma (A.A.); Boustadalila@usmba.ac.ma (D.B.)
- ⁵ AMES Group Polydiagnostic Center, via Padre Carmine Fico, 24, 80013 Casalnuovo di Napoli, Italy
- Correspondence: raffaele.conte86@tiscali.it (R.C.); anna.calarco@cnr.it (A.C.)
- + These authors contributed equally to this work.

Abstract: Plant-derived secondary metabolites represent a reservoir of phytochemicals for regenerative medicine application because of their varied assortment of biological properties including anti-oxidant, anti-inflammatory, antibacterial, and tissue remodeling properties. In addition, bioactive phytochemicals can be easily available, are often more cost-effective in large-scale industrialization, and can be better tolerated compared to conventional treatments mitigating the long-lasting side effects of synthetic compounds. Unfortunately, their poor bioavailability and lack of long-term stability limit their clinical impact. Nanotechnology-based delivery systems can overcome these limitations increasing bioactive molecules' local effectiveness with reduction of the possible side effects on healthy bone. This review explores new and promising strategies in the area of delivery systems with particular emphasis on solutions that enhance bioavailability and/or health effects of plant-derived phytochemicals such as resveratrol, quercetin, epigallocatechin-3-gallate, and curcumin in bone tissue regeneration.

Keywords: bone regeneration; phyto-bioactive compounds; molecular signaling pathways; bone-devices

1. Introduction

Bone defect, due to traumatic injury, congenital disease, or tumor resection, represents a severe ailment affecting millions of people. However, bone regeneration is a complex and dynamic process that involves several actors including osteoprogenitor cells that respond to intracellular and extracellular molecular signaling pathways to ensure bone functional recovery [1,2]. Although the bone tissue is capable of self-repair and renew, regenerative medicine approaches are essential to promote and speed up the healing of bone defects recovering the normal and healthy function of the skeletal system [3]. The conventionally therapeutic approaches have demonstrated a limited efficacy due, for example, to graft rejection, pathogen transmission, and invasive surgical procedures [4]. Owing to the drawbacks and limitations of many bone grafts, bioactive materials that integrate various delivery vehicles, bioactive molecules, stem cells, or demineralized bone matrix may help bone repair creating microenvironments that favor and guide bone regeneration [5,6]. An engineering bone substitute should operate as a proper template for new bone ingrowth showing osteogenic properties (osteoinductive and/or osteoconductive) and being biocompatible with the host tissue. In the last decades, the nanotechnological approach has made great strides in the design of materials able to mimic the natural characteristics of the bone. To improve bone formation/regeneration, phytochemicals such as curcumin,



Citation: Valentino, A.; Di Cristo, F.; Bosetti, M.; Amaghnouje, A.; Bousta, D.; Conte, R.; Calarco, A. Bioactivity and Delivery Strategies of Phytochemical Compounds in Bone Tissue Regeneration. *Appl. Sci.* 2021, *11*, 5122. https://doi.org/10.3390/ app11115122

Academic Editor: Won Ho Park

Received: 16 April 2021 Accepted: 27 May 2021 Published: 31 May 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). resveratrol, oleuropein, quercetin, etc., have often been incorporated into biomaterials as a natural and non-toxic therapeutic alternative to traditional treatments [7,8]. Phytobioactive substances, defined as non-nutritive plant secondary metabolites, could interact with enzymes, proteins, and membrane receptors modulating cell-signal transduction cascade and specific molecular pathways leading to bone anabolic effects and decreasing bone resorption [9,10]. In addition, epidemiological studies have reported a correlation between a diet rich in biologically active factors such as fruits, vegetables, and olive oil and the reduced risk of bone loss and bone-related trauma [11,12]. However, these compounds present in vivo limited biological activity, lack long-term stability, and are subject to oxidation overtime under exposure to oxygen, light, moisture, and heat [13,14]. Therefore, a nanotechnological approach that involves controlled drug delivery systems for natural bioactive molecules could be a solution to avoid invasive procedures and minimize off-target cell behaviors. In addition, these alternative strategies could provide phytochemicals better performance, enhance their low water solubility or very short circulating half-life, improving the functionality and clinical utility [3,15]. This review attempts to summarize the recent works involving delivery systems (i.e., synthetic ceramic, scaffolds, nanoparticles) and phytochemicals to guarantee the protection of natural biomolecules from environmental degradation, to modulate compounds release, and to prolong delivery at localized injury sites. A short chapter on the structure of bone tissue, its functional activities, and the regulatory mechanisms of bone remodeling/regeneration will help in understanding the results discussed.

2. Bone Structure and Function

Bone is a metabolically active tissue with self-healing capability, in constant renewal, adapting its structure to mechanical stimuli, stress, hormonal changes, and repairing structural injuries through a process of remodeling [16]. Bone homeostasis is preserved by the coordinated action between osteoblasts (bone-generating cells derived from mesenchymal stem cells, MSCs) responsible for bone growth and osteoclasts (multinucleated bone-resorbing cells differentiated from the hematopoietic stem) involved in bone resorption [17]. During bone matrix synthesis, first osteoblasts secrete the organic matrix: collagen proteins, mainly collagen type I and non-collagen proteins, such as osteonectin (ON), osteocalcin (OCN), bone sialoprotein (BSPI/II), and osteopontin (OPN), and proteoglycan [18–20]. Secondly, deposition and mineralization of the bone matrix take place through the production of a protein mixing called osteoid that promotes calcium and phosphate adhesion, resulting in the organization and mineralization of new bone [21,22]. Mature osteocytes (derived from MSCs through osteoblastic differentiation) are completely trapped inside the mineralized bone matrix. Due to their strategic location, osteocytes maintain the connections to other osteocytes and osteoblasts and react to several biochemical signaling paths and mechanical stimuli contributing to the regulation of calcium and phosphate homeostasis [23]. When bone resorption should not take place, lining cells avoid direct interface among osteoclasts and bone matrix, playing a significant role in calcium homeostasis and osteoclast differentiation [24–27]. Osteoclasts resorb bone through the release of enzymes and acids capable of dissolving and digesting minerals in the bone, but they also secrete cytokines that affect the activity of surrounding cells inducing mesenchymal stem cells and osteoblasts to initiate osteogenesis in resorption lacuna (remodeling) or another non resorbed site (modeling) [28-30]. The RANKL/RANK/OPG system is a crucial mediator of osteoclastogenesis. In particular, the interaction of RANK with RANKL is required for osteoclast formation, differentiation, activation, and survival. On the contrary, OPG can block RANK/RANKL interaction, thus preventing osteoclast differentiation and activation [31,32]. Irregularities in osteoclastic activity lead to disorders such as osteoporosis and osteopetrosis [33].

3. Bone Regeneration Process

The bone healing process is a complex mechanism of bone regeneration, which involves inflammation, bone production, and bone remodeling phases [34]. Inflammation is usually observed immediately following the fracture at the injury site since tissues swell, bone cells die, and blood vessels break, with consequent formation of hematoma, a source of hematopoietic cells capable of secreting growth factors. The injury to bone leads to the secretion of pro-inflammatory factors like tumor necrosis factor-alpha (TNF- α), bone morphogenetic proteins (BMPs), and interleukins (IL-1, IL-6, IL-11, IL-23). These molecules attract, at the fracture site, macrophages, monocytes, and lymphocytes able to take out damaged necrotic tissue and secrete factors (i.e., vascular endothelial growth factor, VEGF) to stimulate angiogenesis and healing [35]. MSCs migrate to the fracture site and, under BMPs control, start to differentiate into fibroblasts, osteoblasts, and chondroblasts. As a result, chondrogenesis begins to occur, and a fibrocartilaginous callus (also called "soft callus") forms within two weeks. The soft callus undergoes endochondral ossification, which converts fibrocartilaginous callus to bony callus (also identified as "hard callus"). The expression of RANKL promotes further differentiation of chondroblasts, osteoblasts, and osteoclasts [36–38]. Bone regeneration is thinly modulated by several signaling pathways and transcriptional factors. During the early stages of bone healing, the Wnt pathway suppresses the differentiation of mesenchymal stem cells into osteoblasts, while in the later stages it controls the commitment of the undifferentiated cells to the osteoblasts [39]. Notch signaling is another potential pathway with osteoinductive properties. Notch receptors, through their ligand (Jag-1), increase the expression of genes related to osteoblasts as ALP and BSP, inducing osteoclastogenesis [40]. An interesting study demonstrates that the functionalization of titanium implant surface with Jag-1 contributes to the enhancement of osteoblast differentiation, improving osteogenic properties [41]. Indeed, the activation of the Notch pathway leads to an increase of osteogenic differentiation, as highlighted by an upregulation of osteogenic markers, in particular BSP and OCN; bone differentiation proteins as BMP2 and BMP6; and growth/differentiation factor 15 (GDF15) [42]. Several other factors are implicated in bone regeneration processes, such as mitogen-activated protein kinase (MAPK) associated pathways, phosphatidylinositol-4,5-bisphosphate 3-kinase (PI3K)/Akt, growth factors as fibroblast growth factors (FGFs), insulin-like growth factors (IGFs), and VEGF [43]. MAPKs pathway takes part in bone formation and bone healing post-fracture through the transduction of signals induced by numerous growth factors or adhesion molecules. The (PI3K)/Akt signaling pathway promotes the expression of OPG, Runx2, *p*-Akt, and BMP-2 proteins, as well as the proliferation, differentiation, and osteogenesis of osteoblasts [44]. Devi and Dixit demonstrated that the release of rh-VEGF, rh-IGF-I from a polylactide-polyglycolide acid membrane and β-tricalcium phosphate bone graft led to better clinical results such as bone pocket reduction and bone filling simultaneously with respect to growth factors used alone [45]. Furthermore, FGF signaling plays a pivotal role in the intramembranous and endochondral signaling pathway regulating process in osteoprogenitor cells [46]. It has been reported that treatment with hydrogel-bFGF after a fracture has a higher rate of mineralization, as well as an upregulation of Runx2 and osteocalcin in mice [47,48]. Normal bone development requires the downregulation of Runx2 to form mature bone [49].

4. Bioactive Phytochemicals and Bone Signaling Pathways

Phyto-bioactive compounds (i.e., polyphenolic compounds, carotenoids, tocopherols, and phytosterols) are defined as natural secondary metabolites available in fruits, vegetables, grains, and other plant-based foods which provide health benefits and reduce the risk of major chronic diseases [50–53]. In addition to health benefits, bioactive molecules of natural origin are being used as prominent alternatives to chemical preservatives and additives [54–56] as well as in the green synthesis of nanomaterials (i.e., graphene nanosheets, gold nanoparticles, etc.) [57–61]. In bone regenerative medicine, their antioxidant and anti-inflammatory beneficial properties can regulate bone regeneration signaling pathways,

offering an innovative potential therapeutic strategy [62]. Notably, phytochemicals target several critical molecular pathways involved in bone metabolism, such as estrogen signaling pathway, MAPK cascade, Wnt/ β -catenin, sirtuin 1 (Sirt1), TGF- β /BMP, PI3K/Akt, and adenosine monophosphate protein kinase (AMPK) [63]. These pathways can be split into three main classes based on their activity: anti-inflammatory, antioxidants, and bone cell differentiation activity (Figure 1).

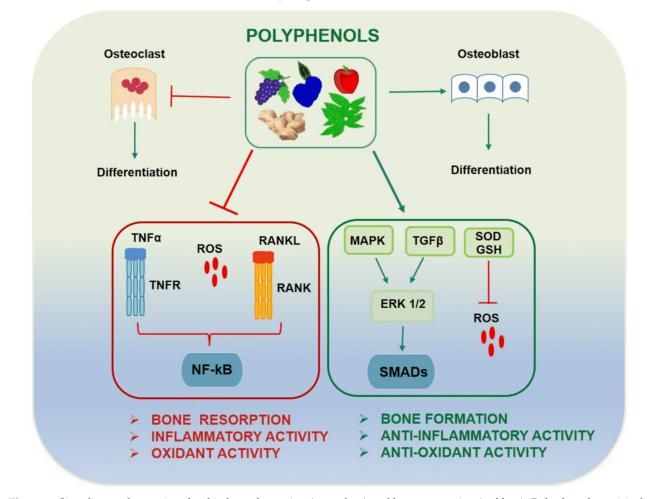


Figure 1. Signaling pathways involved in bone formation (green box) and bone resorption (red box). Polyphenols positively regulate MAPKs/TGF β /ERK1-2 pathway that activates and translocate Smads complex into the nucleus. Activated Smads regulate the expression of transcriptional factors and coactivators important in osteoblasts differentiation and bone formation process including Dlx5, Runx2, and Osx. Polyphenols also up-regulate genes involved in antioxidant activity such as superoxide dismutase (SOD) and glutathione synthetase (GSH). At the same time, polyphenols down-regulate RANKL and TNF α , two master gene regulators of osteoclasts differentiation and inflammatory pathways, respectively.

4.1. Anti-Inflammatory Activity

The anti-inflammatory activity of phyto-molecules is related to the inhibition of genes expression such as TNF- α , [64] monocyte chemotactic protein (MCP)-1, [65] and matrix metalloproteinases (MMPs) [66], and the decrease of pro-inflammatory molecules, such as IL-6, IL-10, and IL-1 β [67–69]. For example, curcumin, an extract from *Curcuma longa*, has been extensively studied due to its ability to inhibit NF-kB and the activation of the activator protein 1 (AP-1) after an inflammatory stimulus [70]. This bioactive molecule suppresses the transcription of pro-inflammatory genes, such as TNF α , IL-6, [64] cyclooxygenase 2 (COX2), and inducible nitric oxide synthase iNOS [71], and contributes to inhibition of MMPs synthesis [72]. Similarly, dried plum polyphenols and tannins indirectly suppress osteoclast differentiation and activity via lowering TNF- α and NO production [73,74] and down-regulating RANKL expression. Epigallocatechin gallate (EGCG),

the most abundant catechin in green tea, exerts an anti-inflammatory effect through MAPKs pathway. Moreover, EGCG reduces the phosphorylation levels of MEK1/2and Raf-1 upstream of ERK1/2 MAPK cascade, [75] promoting bone anabolism, enhancing osteoblasts proliferation, differentiation, and mineralization, and decreasing inflammatory mediators [76,77]. Flavonoids, polyphenols present at relatively low concentrations in most fruit and vegetables, are known as food-based anti-inflammatory agents. Important flavonoids such as quercetin, quercitrin, icaritin, and phloridzin, through downregulation of COX-2 and hypoxia-inducible factor 1-alpha (HIF-1 α) pathways, help to reduce the production of prostaglandin E2 (PGE2), [78] exerting anti-inflammatory and antioxidant actions simultaneously [79].

4.2. Antioxidants Activity

Phytochemicals that operate as direct antioxidant substances are able to activate and regulate antioxidant enzymes, with simultaneous action on the inhibition of oxidases, cyclooxygenases, and other enzymes such as iNOS, involved in radical generation [80]. In general, polyphenols, thanks to their B-ring hydroxyl configuration, display a significant antioxidant action that enhances with the increasing of the total number of OH groups and the attendance of the 3,4-catechol structure [81]. By decreasing the oxidative state, bioactive molecules provide for osteoblasts proliferation, activity, and differentiation through the involvement of crucial molecular signaling pathways.

The ROS-scavenging activity is particularly visible in icaritin (a flavonoid isolated from *Epimedium pubescens*), which can reduce superoxide generation in osteoclasts by indirect action on NFATc1 [82] and in naringin, a flavanone with effects on lipid peroxidation, glutathione (GSH) oxidation, and DNA cleavage [83]. Through the same mechanism of action, curcumin and resveratrol (a polyphenolic compound found in grapes and wine) upregulate in the osteoclast the antioxidant enzymes like glutathione peroxidase (Gpx)-1 and superoxide dismutase (SOD), thus modulating ROS levels [84,85]. Curcumin acts on osteoclastogenesis contributing to mitigate bone loss during osteoclast formation and function, preventing ROS and cytokine production. Myricitrin, a glycoside from myricetin, is able to inhibit bone-resorbing cytokines production under oxidative conditions, displaying protective effects against osteoblast cytotoxicity [86]. On the other hand, polyphenols can exert their antioxidant activity through a mechanism of chelation interacting with metals, in particular, Fe and Zn [87,88]. The EGCG shows cytotoxic properties on osteoclasts thanks to its reductive action on Fe (III) catalyzed by the Fenton reaction, leading to hydroxyl radical's generation [89,90].

4.3. Bone Cells Differentiation Activity

Phytomolecules are not only responsible for bone resorption inhibition, but also promote bone formation by aiming at osteoblasts differentiation. For example, EGCG positively acts on osteoblast differentiation and MSC proliferation by upregulating BMP2 and Runx2 expression [91]. Likewise, myricetin can promote osteoblast differentiation and activity, by targeting SMAD-1/5/8, downstream of BMP signaling [92]. It has been demonstrated that myricetin and Baicalin affect osteoblast and osteoclast differentiation and function also through Wnt/ β -catenin pathway [93,94]. Additionally, due to structural similarity to mammalian estrogens, some polyphenols can bind estrogen receptors (Ers) that are called phytoestrogens [95]. Among them, vanillic acid upregulates the expression of osteoblastic differentiation markers, i.e., Runx2, OCN, and OPG, by activating ERs pathway [96]. Rutin, instead, downregulates the RUNX suppressor genes [97] and exerts its osteogenic effect through an ER-mediated mechanism [98]. In addition, phyto-derived neurotransmitters such as dopamine, commonly found in fruit and vegetables (in particular bananas), promote VEGF and bFGF expression, leading to enhanced angiogenesis and osteogenesis [99]. Furthermore, several studies have highlighted the crucial role of polyphenols in regulating gene activation or silencing through epigenetic modifications such as DNA methylation and histone modification [100]. Resveratrol is one of the main

activators of Sirt, a known NAD-dependent deacetylase, which induces a conformational change in proteins, translating into an increase in enzymatic activity [101]. Resveratrol induces the MSC differentiation into osteoblasts via a very complex mechanism, which could be direct or indirect. Indirectly, resveratrol, through the interaction of Sirt1 with nuclear receptor co-repressor (NcoR), inhibits peroxisome proliferator-activated receptor gamma (PPAR γ) [102], while directly activating RUNX2 transcription factor by forming a Sirt1-Runx2 complex [103]. Resveratrol-mediated activation of Sirt1 enhances phosphorylation of downstream kinases involved in osteoblastic differentiation, such as PKB/Akt, SMAD1/5/8, AMPK, and MAPKs [104,105]. Furthermore, quercetin stimulates osteoblast differentiation through the stimulation of the expression of TGF- β 1, BMP-2, and Runx2, via activation of ERK1/2, p38, and JNK MAPKs [106]. Finally, curcumin regulates the expression of genes implicated in RANKL-induced osteoclast differentiation through the suppression of NF-kB [107]. Table 1 recaps the phytochemical-related bone regeneration signaling pathways.

Table 1. Summary of phytochemical signaling pathways for bone regeneration.

Phytochemical	Signaling Pathway	Reference	
Curcumin	NF-kB pathways,	[75,89,90]	
	Redox-sensitive signaling pathways		
EGCG	MAPKs signaling pathways	[82]	
Quercetin	COX2/HIF1αsignaling	[83,84,108]	
	ERK1-2/MAPKs signaling		
Resveratrol	Redox-sensitive signaling pathways		
	Sirt/RUNX2 signaling	[89,90,109]	
	MAPKs pathways		
Incaritin	NFATc signaling	[87]	
Naringin	Glutathione Pathway (GSH)	[88]	
Myricetin	SMAD/BMP signaling	[97]	
	Wnt/ β -catenin signaling		
Vanillic acid	ERs pathways	[101]	
Rutin	ERs pathways	[102,103]	

The cellular responses resulting from the activation of different biological pathways underline the importance of natural bioactive molecules and their ability to modulate inflammatory processes, oxidative stress, and cellular differentiation.

5. Phytochemical-Delivery Vehicles in Bone Tissue Regeneration

To date, the most effective treatment for bone-defect restoration is represented by living tissue transplantation (autologous bone) and/or devitalized donor bone (allograft) because of their notable osteoconductive and osteoinductive properties [108,109]. However, the potential for disease transmission, the limited amount of donor tissue, and postoperative pain at the donor site represent some of drawbacks related to these approaches. To overcome these challenges, new and promising strategies involving delivery systems and phytochemicals have been developed in bone tissue regeneration [110–113]. A successful delivery system should be able to protect phytochemicals from degradation, enhancing their poor bioavailability and minimizing off-target tissue effects [114,115]. These constructs, determining a localized delivery of the natural bioactive molecules, should promote the normal process of bone regeneration and minimize tissue toxicity caused by systemic drug administration (Figure 2) [116].

5.1. Ceramics

Synthetic ceramic materials are inorganic material favorably used in dentistry that proved excellent mechanical properties and osteo-conduciveness owing to their good bio-compatibility, reproducibility, non-immunogenicity [117,118]. Ceramic nanocomposites in the form of particles or nanofibers could mimic the hierarchical arrangement of native bone mineral phase, providing a functional scaffold for cell adhesion [119–122], but are

not able to prevent the cause of bone resorption, to control specific anti-osteoclastogenic actions, or to counteract the damage related to oxidative stress. To overcome these disadvantages, polyphenols are widely used to enhance periodontal regeneration or to remineralize bone tissue, due to their antioxidant, free-radical scavenging, and antimicrobial properties [123–125]. The preparation of these composites is often optimized to form nanopores (pore diameter $< 0.1 \,\mu$ m) to increase the specific surface area of the scaffolds, allowing for better drug loading and higher bioactivity [125]. For example, Iviglia G et al. patented a polyphenol-based collagen gel with granular ceramic fillers to fill the peri-implant bone defects. Such material, characterized by strong mechanical scaffolding properties, combines the pro-osteogenic action of collagen with the anti-inflammatory, antioxidant, and anti-osteoclastogenic activity of a polyphenolic mixture extracted from the pomace of the Croatina grape variety [126]. In vitro and in vivo results demonstrated that both the control of inflammation and oxidative stress and the enhancement of early bone matrix deposition are necessary in the case of oral disease. Cazzola M et al. produced a silica-based bioactive glass coupled with gallic acid and polyphenols extracted from red grape skins and green tea leaves. These modified bioactive glasses showed enhanced free radical scavenging activity [127]. Zhou et al. proposed a simple protocol to functionalize porous calcium phosphate ceramics (PCPC) using dietetic tea polyphenols (TP). TP molecules modulated the nucleation and crystallization of calcium phosphate nanorods and promoted bone mesenchymal stem cell (BMSC) proliferation and differentiation, increasing BMP2, ErK/MAPK, and JNK/MAPK levels and cell mineralization capacity [128].

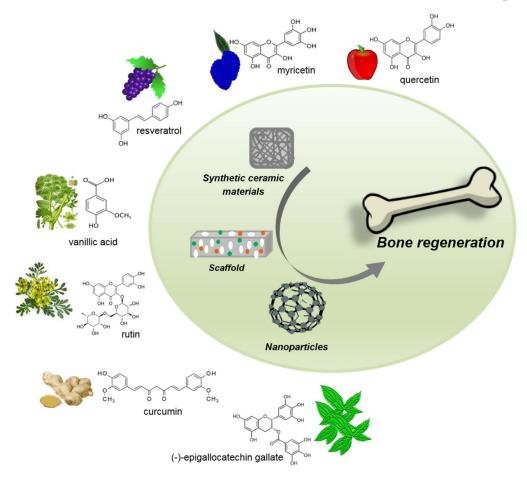


Figure 2. Schematic representation of the main phyto-bioactive nano delivery systems involved in bone regeneration.

5.2. Scaffolds

Since the scaffold surface functionalization with polyphenols increases the bone regeneration capacity, the use of enriched scaffolds is considered a promising tool for bone bioengineering [129–131]. Ideally, a scaffold should mimic the extracellular matrix characteristics of the organ of interest to form cell/tissue-specific combination, pattern/topology, and mechanical properties able to support tissue formation [131]. The combination of scaffolds and phytochemicals is used to improve adhesion, growth, or differentiation of cells through the action of the bioactive compound for which the release can be controlled through three methods. When drugs are incorporated into the matrix of the scaffold, the release kinetics are regulated by the degradation of the polymer. Similarly, when scaffold surfaces are coated with a polymer/drug layer, the release is controlled by diffusion and degradation of the coating polymer. Conversely, the integration of micro-nano spheres into scaffolds offers a third mechanism of release based on the degradation of the scaffold and the consequent diffusion of the particle [132]. For example, Santin M et al. used biodegradable antioxidant scaffolds based on soybean (SB), as bone filler. This material showed an invitro block of osteoclast activation succeeding incubation with SB, with a parallel inhibitory effect on monocyte/macrophage activity and an improved ability to induce mineralization in osteoblasts [133]. In another study, the SB granules implanted in rabbits led to bone repair with distinctive morphology from non-treated defects [134].

Wang W et al. produced scaffolds loaded with resveratrol by grafting the polyphenol to polyacrylic acid (PAA) and integrating this molecular drug into atelocollagen (Coll) hydrogels (Coll/PAA-RSV) [135]. These scaffolds supported the growth of chondrocytes and BMSCs protecting cells against reactive oxygen species. Moreover, the in vivo implantation on rabbits led to the disappearance of osteochondral defects and the integration of the newly formed tissue with surrounding tissue and subchondral bone. Li et al. functionalized a poly-ε-caprolactone (PCL) surface with resveratrol, obtaining an osteogenic porous material. The presence of polyphenol on a scaffold surface increases the mineralization in stromal cells with an improvement in bone regenerating capacity [136]. Kamath MS et al. formulated a porous composite scaffold integrating PLC with resveratrol-loaded albumin nanoparticles. This 3D material produced a significant increment in cell proliferation, ALP activity, and mineralization imparting osteogenic properties to PCL scaffold [137]. Riccitiello et al. synthesized a PLA electrospinning membrane able to release resveratrol in a tunable manner for the preservation of the alveolar socket after tooth extraction. The controlled release of resveratrol influenced in vitro osteoblast and osteoclast differentiation [138]. In another work, the resveratrol released form PLA membrane presents a significant antibacterial and antibiofilm activity versus Pseudomonas aeruginosa and Streptococcus mutans, becoming a promising solution for the prevention of implant-associated infections [51]. For cranio-facial tissue-engineering applications, Wang et al. combine a collagen scaffold loaded with resveratrol with human adipose stem cells (hASCs). This composite promotes epidermal wound healing and bone mineralization [139]. Wang et al. introduced dopamine (D) onto strontium-doped calcium polyphosphate (SCPP) scaffolds with silk fibroin (SF). SCPP/D/SF stimulated angiogenic factor secretion, osteogenesis, and had great biocompatibility (Figure 3). Then, SCPP/D/SF could fulfill a role as a potential scaffold for bone tissue engineering with the ability to speed up bone regeneration and vascularization. (Figure 4) [140]. Dhand et al. reported the synthesis by electrospinning of bone-like composite structures containing catecholamines and Ca(2+). Human fetal osteoblasts seeded on these collagen scaffolds exhibited enhanced cell adhesion, penetration, proliferation, and differentiation as well as increased osteogenic expression of osteocalcin, osteopontin, and bone matrix elements [141]. Lee et al. reported an easy, multifunctional surface modification using catechin to enhance the polymeric scaffolds functionality for bone regeneration by stem cells. These catechin-functionalized polymer nanofiber scaffolds, in a critical-sized calvarial bone defect, markedly supported bone formation by hADSC transplantation [142].

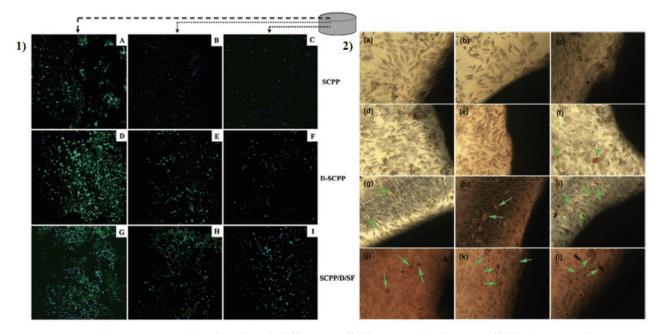


Figure 3. Image (1) represents cells cultured with different scaffolds. Image (2) shows scaffolds alizarin red staining on day 4 (**a**–**c**), day 7 (**d**–**f**), day 10 (**g**–**i**), day 14 (**j**–**l**); SCPP (**a**,**d**,**g**,**j**), D-SCPP (**b**,**e**,**h**,**k**), SCPP/D/SF (**c**,**f**,**i**,**l**), respectively. Green arrows: calcium nodules. Courtesy of: Wang X, Gu Z, Jiang B, Li L, Yu X. Surface modification of strontium-doped porous bioactive ceramic scaffolds via poly(DOPA) coating and immobilizing silk fibroin for excellent angiogenic and osteogenic properties. Biomater Sci. April 2016;4(4):678–88. doi: 10.1039/c5bm00482a. Epub 12 February 2016. PMID: 26870855.

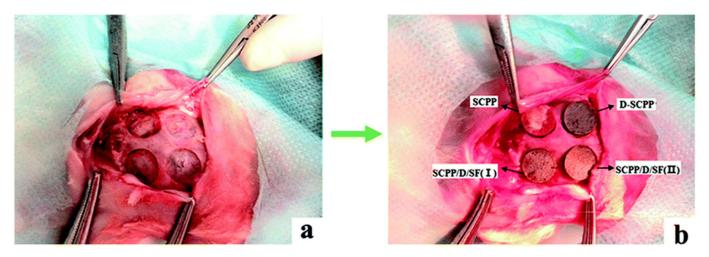


Figure 4. Photographs of animal modeling in vivo. Defects on the cranial bone (**a**,**b**) materials implanted. Courtesy of: Wang X, Gu Z, Jiang B, Li L, Yu X. Surface modification of strontium-doped porous bioactive ceramic scaffolds via poly(DOPA) coating and immobilizing silk fibroin for excellent angiogenic and osteogenic properties. Biomater Sci. April 2016;4(4):678–88. doi: 10.1039/c5bm00482a. Epub 12 February 2016. PMID: 26870855.

5.3. Nanoparticles

The therapeutic efficacy of natural bioactive molecules can be improved by nanotechnological approaches. The design of drug delivery systems with pre-determined physico-chemical properties permits an increase of phytochemical bioavailability and reduces their toxic side effects [53]. For example, positive surface charges could facilitate the transport of nanoparticles through small intestinal epithelial cells improving the oral bioavailability, while the introduction of polyethylene glycol chains limits opsonization prolonging circulation times [143–145]. Moreover, the modulation of structure-property relationships (i.e., size, geometry or shape, material composition, etc.) of nanoparticles can influence the transport mechanism, facilitating their internalization into the target cells.

He L et al. produced tea polyphenol-modified calcium phosphate nanoparticles (TP-CaP) able to enhance remineralization of preformed enamel lesions on bovine incisors. Moreover, the released tea polyphenols inhibited bacterial growth and enzyme activities [146]. Wang produced gold nanoparticles (Au-NPs) formed using Anogeissus latifolia (A. latifolia) phytochemicals. Such nano-vehicles showed great osteoinductive potential and analgesic properties and were characterized by exceptional blood compatibility and cytocompatibility [147]. Felice et al. synthesized polyphenol-based mucoadhesive polymeric nanoparticles (GSE-NP) able to protect endothelial progenitor cells (EPCs) from oxidative stress. These vehicles demonstrated strong antioxidant capacity thanks to their high content in total polyphenols [148]. Del Prado Audelo et al. synthesized nanoparticles of PCL and Pluronic[®] F-68, loaded with curcumin. These nanoparticles were able to reduce cell proliferation without affecting cell migration and adhesion, and decrease the oxidative stress induced by hydrogen peroxide exhibiting a cytoprotective effect [149]. Finally, Malathy S and Priya R Iyer used chitosan to prepare Naringin-loaded chitosan nanoparticles (NCN). The NCN had strong anti-inflammatory, anti-coagulant, antioxidant, and anti-cancerous effects. Furthermore, these nanoparticles promoted osteoblast differentiation, so they could be considered an efficient model for bone tissue regeneration [150]. All described delivery systems are summarized in Table 2.

Table 2. Summary of bioactive compounds-based devices system and their effects on bone regeneration.

Bioactive Compound	Device	Effect	Ref
- Gallic acid - Tea polyphenols	Synthetic ceramic materials	- pro-osteogenic - anti-inflammatory - antioxidant	[128,134]
- Resveratrol - Phlorotannins - Catechins	Scaffolds -hydrogel - PLGA - PCL and PLA	- antioxidant - pro-differentiating - pro-osteogenic - wound healing	[135–137,142]
- EGCG - Catechin - Pro-anthocyanidins - Curcumin - Naringin	Nanoparticles - TP-CaP - Au-NPs - GSE-NP - Cur–PCL	 osteopromotive osteoblast differentiation osteoblast proliferation cytoprotective remineralization antioxidant anti inflammatory 	[147–150]

6. Conclusions

In this review paper, recent developments in delivery systems for phytochemicals release for bone tissue regeneration were discussed. Despite the updates reviewed in this paper, more work is required to develop materials that can present controlled release kinetics and degradation, and that directly influence the rate of new bone formation. Soon, researchers, in accordance with clinicians, might be able to design and develop new delivery systems with improved characteristics that mimic bone microenvironment at the site of implantation, promoting the inflammation, angiogenesis, and osteogenesis phases of new bone formation. The interaction between a nanotechnological approach and natural-derived compounds with osteogenic, anti-oxidant, antimicrobial, and anti-inflammatory activities will open up a new era of advanced treatment and solutions to prevent and/or treat bone-related complications.

Author Contributions: Conceptualization, A.C. and R.C.; writing—original draft preparation, A.V., F.D.C., and R.C.; writing—review and editing, A.V., F.D.C., R.C., and A.C.; visualization, A.A. and D.B.; supervision, M.B. and A.C.; funding acquisition, A.C. and M.B. All authors have read and agreed to the published version of the manuscript.

Funding: This work was financially supported by the Italian Ministry of University and Research, PON 03PE_00110_1/ptd1_000410 Titolo: Sviluppo di nanotecnologie Orientate alla Rigenerazione e Ricostruzione tissutale, Implantologia e Sensoristica in Odontoiatria/oculistica (SORRISO) (approval date 02/01/2019); POR Campania FESR 2014_2020 "Tecnologie abilitanti per la sintesi ecosostenibile di nuovi materiali per la Restaurativa dentale"—ABILTECH (approval date 29 October 2018).This work was supported also by grant from Fondazione Cariplo, "High added-value bioactive polyphenols recovered from waste of olive oil production" (agreement 2018-1001), "Economia Circolare-Ricerca per un Futuro Sostenibile" program.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Acknowledgments: We would like to extend our sincere thanks to the project H2020- MSCA-RISE-Marie Sklodowska-Curie Actions (MSCA) Research and Innovation Staff Exchange (RISE) for funding this work, Project Acronym: VAHVISTUS-Project Number: 734759.

Conflicts of Interest: The authors have not declared any conflict of interest.

References

- 1. Li, Y.; Liu, C. Nanomaterial-based bone regeneration. *Nanoscale* 2017, 9, 4862–4874. [CrossRef] [PubMed]
- Bosetti, M.; Leigheb, M.; Brooks, R.A.; Boccafoschi, F.; Cannas, M.F. Regulation of osteoblast and osteoclast functions by FGF-6. J. Cell. Physiol. 2010, 225, 466–471. [CrossRef] [PubMed]
- 3. Yi, H.; Ur Rehman, F.; Zhao, C.; Liu, B.; He, N. Recent advances in nano scaffolds for bone repair. *Bone Res.* **2016**, *4*, 16050. [CrossRef]
- 4. Webber, M.J.; Khan, O.F.; Sydlik, S.A.; Tang, B.C.; Langer, R. A perspective on the clinical translation of scaffolds for tissue engineering. *Ann. Biomed. Eng.* **2015**, *43*, 641–656. [CrossRef]
- 5. Bosetti, M.; Lloyd, A.W.; Santin, M.; Denyer, S.P.; Cannas, M. Effects of phosphatidylserine coatings on titanium on inflammatory cells and cell-induced mineralisation in vitro. *Biomaterials* **2005**, *26*, 7572–7578. [CrossRef]
- Bosetti, M.; Boccafoschi, F.; Calarco, A.; Leigheb, M.; Gatti, S.; Piffanelli, V.; Peluso, G.; Cannas, M. Behaviour of human mesenchymal stem cells on a polyelectrolyte-modified HEMA hydrogel for silk-based ligament tissue engineering. *J. Biomater. Sci. Polym. Ed.* 2008, 19, 1111–1123. [CrossRef] [PubMed]
- Byberg, L.; Bellavia, A.; Larsson, S.C.; Orsini, N.; Wolk, A.; Michaëlsson, K. Mediterranean Diet and Hip Fracture in Swedish Men and Women. J. Bone Miner. Res. Off. J. Am. Soc. Bone Miner. Res. 2016, 31, 2098–2105. [CrossRef] [PubMed]
- Lambert, M.N.T.; Thybo, C.B.; Lykkeboe, S.; Rasmussen, L.M.; Frette, X.; Christensen, L.P.; Jeppesen, P.B. Combined bioavailable isoflavones and probiotics improve bone status and estrogen metabolism in postmenopausal osteopenic women: A randomized controlled trial. *Am. J. Clin. Nutr.* 2017, *106*, 909–920. [CrossRef]
- 9. Nijveldt, R.J.; van Nood, E.; van Hoorn, D.E.; Boelens, P.G.; van Norren, K.; van Leeuwen, P.A. Flavonoids: A review of probable mechanisms of action and potential applications. *Am. J. Clin. Nutr.* **2001**, *74*, 418–425. [CrossRef]
- 10. Fraga, C.G.; Galleano, M.; Verstraeten, S.V.; Oteiza, P.I. Basic biochemical mechanisms behind the health benefits of polyphenols. *Mol. Asp. Med.* **2010**, *31*, 435–445. [CrossRef]
- 11. Di Meo, F.; Valentino, A.; Petillo, O.; Peluso, G.; Filosa, S.; Crispi, S. Bioactive Polyphenols and Neuromodulation: Molecular Mechanisms in Neurodegeneration. *Int. J. Mol. Sci.* **2020**, *21*, 2564. [CrossRef]
- 12. Pandey, K.B.; Rizvi, S.I. Plant polyphenols as dietary antioxidants in human health and disease. *Oxid. Med. Cell. Longev.* **2009**, *2*, 270–278. [CrossRef] [PubMed]
- 13. Scalbert, A.; Williamson, G. Dietary intake and bioavailability of polyphenols. *J. Nutr.* **2000**, *130* (Suppl. 8S), 2073s–2085s. [CrossRef]
- 14. D'Archivio, M.; Filesi, C.; Varì, R.; Scazzocchio, B.; Masella, R. Bioavailability of the polyphenols: Status and controversies. *Int. J. Mol. Sci.* **2010**, *11*, 1321–1342. [CrossRef] [PubMed]
- 15. Zhu, L.; Luo, D.; Liu, Y. Effect of the nano/microscale structure of biomaterial scaffolds on bone regeneration. *Int. J. Oral Sci.* **2020**, *12*, *6*. [CrossRef] [PubMed]
- 16. Ansari, M. Bone tissue regeneration: Biology, strategies and interface studies. Prog. Biomater. 2019, 8, 223–237. [CrossRef]
- Rosenberg, N.; Rosenberg, O.; Soudry, M. Osteoblasts in bone physiology-mini review. *Rambam Maimonides Med. J.* 2012, 3, e0013. [CrossRef]
- Ducy, P.; Zhang, R.; Geoffroy, V.; Ridall, A.L.; Karsenty, G. Osf2/Cbfa1: A Transcriptional Activator of Osteoblast Differentiation. *Cell* 1997, 89, 747–754. [CrossRef]

- 19. Fakhry, M.; Hamade, E.; Badran, B.; Buchet, R.; Magne, D. Molecular mechanisms of mesenchymal stem cell differentiation towards osteoblasts. *World J. Stem Cells* **2013**, *5*, 136–148. [CrossRef]
- Nakashima, K.; Zhou, X.; Kunkel, G.; Zhang, Z.; Deng, J.M.; Behringer, R.R.; de Crombrugghe, B. The novel zinc finger-containing transcription factor osterix is required for osteoblast differentiation and bone formation. *Cell* 2002, *108*, 17–29. [CrossRef]
- 21. James, R.; Deng, M.; Laurencin, C.T.; Kumbar, S.G. Nanocomposites and bone regeneration. *Front. Mater. Sci.* 2011, *5*, 342–357. [CrossRef]
- 22. Sikavitsas, V.I.; Temenoff, J.S.; Mikos, A.G. Biomaterials and bone mechanotransduction. *Biomaterials* **2001**, *22*, 2581–2593. [CrossRef]
- 23. Brannigan, K.; Griffin, M. An Update into the Application of Nanotechnology in Bone Healing. *Open Orthop. J.* **2016**, *10*, 808–823. [CrossRef] [PubMed]
- 24. Miller, S.C.; de Saint-Georges, L.; Bowman, B.M.; Jee, W.S. Bone lining cells: Structure and function. *Scanning Microsc.* **1989**, *3*, 953–960, discussion 60-1. [PubMed]
- 25. Aarden, E.M.; Nijweide, P.J.; Burger, E.H. Function of osteocytes in bone. J. Cell. Biochem. 1994, 55, 287–299. [CrossRef]
- Andersen, T.L.; Sondergaard, T.E.; Skorzynska, K.E.; Dagnaes-Hansen, F.; Plesner, T.L.; Hauge, E.M.; Plesner, T.; Delaisse, J.M. A physical mechanism for coupling bone resorption and formation in adult human bone. *Am. J. Pathol.* 2009, 174, 239–247. [CrossRef]
- 27. Everts, V.; Delaissé, J.M.; Korper, W.; Jansen, D.C.; Tigchelaar-Gutter, W.; Saftig, P.; Beertsen, W. The Bone Lining Cell: Its Role in Cleaning Howship's Lacunae and Initiating Bone Formation. *J. Bone Miner. Res.* **2002**, *17*, 77–90. [CrossRef]
- Boyce, B.F.; Hughes, D.E.; Wright, K.R.; Xing, L.; Dai, A. Recent advances in bone biology provide insight into the pathogenesis of bone diseases. *Lab. Investig. J. Tech. Methods Pathol.* 1999, 79, 83–94.
- Crockett, J.C.; Mellis, D.J.; Scott, D.I.; Helfrich, M.H. New knowledge on critical osteoclast formation and activation pathways from study of rare genetic diseases of osteoclasts: Focus on the RANK/RANKL axis. Osteoporos. Int. J. Establ. Result Coop. Eur. Found. Osteoporos. Natl. Osteoporos. Found. USA 2011, 22, 1–20. [CrossRef] [PubMed]
- Boyce, B.F.; Xing, L. Functions of RANKL/RANK/OPG in bone modeling and remodeling. Arch. Biochem. Biophys. 2008, 473, 139–146. [CrossRef]
- Yavropoulou, M.P.; Yovos, J.G. Osteoclastogenesis—Current knowledge and future perspectives. J. Musculoskelet. Neuronal Interact. 2008, 8, 204–216.
- 32. Phan, T.C.; Xu, J.; Zheng, M.H. Interaction between osteoblast and osteoclast: Impact in bone disease. *Histol. Histopathol.* 2004, 19, 1325–1344. [CrossRef]
- 33. Kular, J.; Tickner, J.; Chim, S.M.; Xu, J. An overview of the regulation of bone remodeling at the cellular level. *Clin. Biochem.* **2012**, 45, 863–873. [CrossRef]
- Schindeler, A.; McDonald, M.M.; Bokko, P.; Little, D.G. Bone remodeling during fracture repair: The cellular picture. Semin. Cell Dev. Biol. 2008, 19, 459–466. [CrossRef] [PubMed]
- 35. Sheen, J.R.; Garla, V.V. Fracture Healing Overview; StatPearls Publishing: Treasure Island, FL, USA, 2019.
- Ghiasi, M.S.; Chen, J.; Vaziri, A.; Rodriguez, E.K.; Nazarian, A. Bone fracture healing in mechanobiological modeling: A review of principles and methods. *Bone Rep.* 2017, 6, 87–100. [CrossRef] [PubMed]
- 37. Marsell, R.; Einhorn, T.A. The biology of fracture healing. *Injury* 2011, 42, 551–555. [CrossRef] [PubMed]
- Kostenuik, P.; Mirza, F.M. Fracture healing physiology and the quest for therapies for delayed healing and nonunion. J. Orthop. Res. Off. Publ. Orthop. Res. Soc. 2017, 35, 213–223. [CrossRef]
- Krishnan, V.; Bryant, H.U.; Macdougald, O.A. Regulation of bone mass by Wnt signaling. J. Clin. Investig. 2006, 116, 1202–1209. [CrossRef] [PubMed]
- Zhu, F.; Sweetwyne, M.T.; Hankenson, K.D. PKCδ is required for Jagged-1 induction of human mesenchymal stem cell osteogenic differentiation. *Stem Cells* 2013, *31*, 1181–1192. [CrossRef]
- Dishowitz, M.I.; Zhu, F.; Sundararaghavan, H.G.; Ifkovits, J.L.; Burdick, J.A.; Hankenson, K.D. Jagged1 immobilization to an osteoconductive polymer activates the Notch signaling pathway and induces osteogenesis. *J. Biomed. Mater. Res. Part A* 2014, 102, 1558–1567. [CrossRef]
- Chakravorty, N.; Hamlet, S.; Jaiprakash, A.; Crawford, R.; Oloyede, A.; Alfarsi, M.; Xiao, Y.; Ivanovski, S. Pro-osteogenic topographical cues promote early activation of osteoprogenitor differentiation via enhanced TGFβ, Wnt, and Notch signaling. *Clin. Oral Implants Res.* 2014, 25, 475–486. [CrossRef] [PubMed]
- Herford, A.S.; Stoffella, E.; Tandon, R. Reconstruction of mandibular defects using bone morphogenic protein: Can growth factors replace the need for autologous bone grafts? A systematic review of the literature. *Plast. Surg. Int.* 2011, 2011, 165824. [CrossRef] [PubMed]
- 44. Zheng, A.Q.; Xiao, J.; Xie, J.; Lu, P.P.; Ding, X. bFGF enhances activation of osteoblast differentiation and osteogenesis on titanium surfaces via PI3K/Akt signaling pathway. *Int. J. Clin. Exp. Pathol.* **2016**, *9*, 4680–4692.
- 45. Devi, R.; Dixit, J. Clinical Evaluation of Insulin like Growth Factor-I and Vascular Endothelial Growth Factor with Alloplastic Bone Graft Material in the Management of Human Two Wall Intra-Osseous Defects. J. Clin. Diagn. Res. JCDR 2016, 10, ZC41–ZC46. [CrossRef]
- 46. Ornitz, D.M.; Marie, P.J. FGF signaling pathways in endochondral and intramembranous bone development and human genetic disease. *Genes Dev.* 2002, *16*, 1446–1465. [CrossRef]

- 47. Furuya, H.; Tabata, Y.; Kaneko, K. Bone regeneration for murine femur fracture by gelatin hydrogels incorporating basic fibroblast growth factor with different release profiles. *Tissue Eng. Part A* 2014, 20, 1531–1541. [CrossRef]
- Liu, T.M.; Lee, E.H. Transcriptional regulatory cascades in Runx2-dependent bone development. *Tissue Eng. Part B Rev.* 2013, 19, 254–263. [CrossRef]
- 49. Zhang, X.; Yang, M.; Lin, L.; Chen, P.; Ma, K.T.; Zhou, C.Y.; Ao, Y.F. Runx2 overexpression enhances osteoblastic differentiation and mineralization in adipose–derived stem cells in vitro and in vivo. *Calcif. Tissue Int.* **2006**, *79*, 169–178. [CrossRef]
- 50. Conte, R.; Luca, I.D.; Luise, A.D.; Petillo, O.; Calarco, A.; Peluso, G. New Therapeutic Potentials of Nanosized Phytomedicine. *J. Nanosci. Nanotechnol.* **2016**, *16*, 8176–8187. [CrossRef]
- 51. Bonadies, I.; Di Cristo, F.; Valentino, A.; Peluso, G.; Calarco, A.; Di Salle, A. pH-Responsive Resveratrol-Loaded Electrospun Membranes for the Prevention of Implant-Associated Infections. *Nanomaterials* **2020**, *10*, 1175. [CrossRef]
- Amrati, F.E.; Bourhia, M.; Slighoua, M.; Ibnemoussa, S.; Bari, A.; Ullah, R.; Amaghnouje, A.; Di Cristo, F.; El Mzibri, M.; Calarco, A.; et al. Phytochemical Study on Antioxidant and Antiproliferative Activities of Moroccan Caralluma europaea Extract and Its Bioactive Compound Classes. *Evid. Based Complement. Altern. Med.* 2020, 2020, 8409718. [CrossRef] [PubMed]
- 53. Conte, R.; Marturano, V.; Peluso, G.; Calarco, A.; Cerruti, P. Recent Advances in Nanoparticle-Mediated Delivery of Anti-Inflammatory Phytocompounds. *Int. J. Mol. Sci.* 2017, *18*, 709. [CrossRef] [PubMed]
- Moccia, F.; Agustin-Salazar, S.; Berg, A.L.; Setaro, B.; Micillo, R.; Pizzo, E.; Weber, F.; Gamez-Meza, N.; Schieber, A.; Cerruti, P. Pecan (Carya illinoinensis (Wagenh.) K. Koch) Nut Shell as an Accessible Polyphenol Source for Active Packaging and Food Colorant Stabilization. ACS Sustain. Chem. Eng. 2020, 8, 6700–6712. [CrossRef] [PubMed]
- Marturano, V.; Bizzarro, V.; De Luise, A.; Calarco, A.; Ambrogi, V.; Giamberini, M.; Tylkowski, B.; Cerruti, P. Essential oils as solvents and core materials for the preparation of photo-responsive polymer nanocapsules. *Nano Res.* 2018, 11, 2783–2795. [CrossRef]
- Agustin-Salazar, S.; Gamez-Meza, N.; Medina-Juárez, L.A.; Malinconico, M.; Cerruti, P. Stabilization of Polylactic Acid and Polyethylene with Nutshell Extract: Efficiency Assessment and Economic Evaluation. ACS Sustain. Chem. Eng. 2017, 5. [CrossRef]
- Agustin-Salazar, S.; Gamez-Meza, N.; Medina-Juàrez, L.À.; Soto-Valdez, H.; Cerruti, P. From Nutraceutics to Materials: Effect of Resveratrol on the Stability of Polylactide. ACS Sustain. Chem. Eng. 2014, 2, 1534–1542. [CrossRef]
- 58. Srivastava, S.K.; Ogino, C.; Kondo, A. Green synthesis of thiolated graphene nanosheets by alliin (garlic) and its effect on the deposition of gold nanoparticles. *RSC Adv.* **2013**, *4*, 5986. [CrossRef]
- 59. Elif, Ö.; Belma, Ö.; İlkay, Ş. Production of biologically safe and mechanically improved reduced graphene oxide/hydroxyapatite composites. *Mater. Res. Express* **2017**, *4*, 015601. [CrossRef]
- 60. Alegria, E.C.B.A.; Ribeiro, A.P.C.; Mendes, M.; Ferraria, A.M.; do Rego, A.M.B.; Pombeiro, A.J.L. Effect of Phenolic Compounds on the Synthesis of Gold Nanoparticles and its Catalytic Activity in the Reduction of Nitro Compounds. *Nanomaterials* **2018**, *8*, 320. [CrossRef]
- 61. Swilam, N.; Nematallah, K.A. Polyphenols profile of pomegranate leaves and their role in green synthesis of silver nanoparticles. *Sci. Rep.* **2020**, *10*, 14851. [CrossRef]
- 62. Conte, R.; Calarco, A.; Napoletano, A.; Valentino, A.; Margarucci, S.; Di Cristo, F.; Di Salle, A.; Peluso, G. Polyphenols Nanoencapsulation for Therapeutic Applications. *Biomol. Res. Ther.* **2016**, *5*:2. [CrossRef]
- 63. Torre, E. Molecular signaling mechanisms behind polyphenol-induced bone anabolism. *Phytochem. Rev.* **2017**, *16*, 1183–1226. [CrossRef]
- 64. Zhou, T.; Chen, D.; Li, Q.; Sun, X.; Song, Y.; Wang, C. Curcumin inhibits inflammatory response and bone loss during experimental periodontitis in rats. *Acta Odontol. Scand.* **2013**, *71*, 349–356. [CrossRef]
- 65. Bandyopadhyay, S.; Lion, J.M.; Mentaverri, R.; Ricupero, D.A.; Kamel, S.; Romero, J.R.; Chattopadhyay, N. Attenuation of osteoclastogenesis and osteoclast function by apigenin. *Biochem. Pharmacol.* 2006, 72, 184–197. [CrossRef] [PubMed]
- 66. La, V.D.; Howell, A.B.; Grenier, D. Cranberry proanthocyanidins inhibit MMP production and activity. *J. Dent. Res.* 2009, *88*, 627–632. [CrossRef] [PubMed]
- 67. Pang, J.L.; Ricupero, D.A.; Huang, S.; Fatma, N.; Singh, D.P.; Romero, J.R.; Chattopadhyay, N. Differential activity of kaempferol and quercetin in attenuating tumor necrosis factor receptor family signaling in bone cells. *Biochem. Pharmacol.* 2006, 71, 818–826. [CrossRef] [PubMed]
- La, V.D.; Tanabe, S.; Grenier, D. Naringenin inhibits human osteoclastogenesis and osteoclastic bone resorption. J. Periodontal Res. 2009, 44, 193–198. [CrossRef]
- Comalada, M.; Ballester, I.; Bailón, E.; Sierra, S.; Xaus, J.; Gálvez, J.; Sánchez de Medina, F.; Zarzuelo, A. Inhibition of proinflammatory markers in primary bone marrow-derived mouse macrophages by naturally occurring flavonoids: Analysis of the structure–activity relationship. *Biochem. Pharmacol.* 2006, 72, 1010–1021. [CrossRef]
- 70. Bharti, A.C.; Takada, Y.; Aggarwal, B.B. Curcumin (diferuloylmethane) inhibits receptor activator of NF-kappa B ligand-induced NF-kappa B activation in osteoclast precursors and suppresses osteoclastogenesis. J. Immunol. 2004, 172, 5940–5947. [CrossRef]
- Chowdhury, T.T.; Salter, D.M.; Bader, D.L.; Lee, D.A. Signal transduction pathways involving p38 MAPK, JNK, NFkappaB and AP-1 influences the response of chondrocytes cultured in agarose constructs to IL-1beta and dynamic compression. *Inflamm. Res.* Off. J. Eur. Histamine Res. Soc. 2008, 57, 306–313. [CrossRef]
- 72. Kumar, D.; Kumar, M.; Saravanan, C.; Singh, S.K. Curcumin: A potential candidate for matrix metalloproteinase inhibitors. *Expert Opin. Ther. Targets* **2012**, *16*, 959–972. [CrossRef]

- 73. Bu, S.Y.; Hunt, T.S.; Smith, B.J. Dried plum polyphenols attenuate the detrimental effects of TNF-alpha on osteoblast function coincident with up-regulation of Runx2, Osterix and IGF-I. *J. Nutr. Biochem.* **2009**, *20*, 35–44. [CrossRef] [PubMed]
- 74. Bhatia, I.S.; Bajaj, K.L. Tannins in black-plum (Syzygium cumini L.) seeds. Biochem. J. 1972, 128, 56. [CrossRef]
- 75. Tokuda, H.; Takai, S.; Matsushima-Nishiwaki, R.; Akamatsu, S.; Hanai, Y.; Hosoi, T.; Harada, A.; Ohta, T.; Kozawa, O. (–)-epigallocatechin gallate enhances prostaglandin F2α-induced VEGF synthesis via upregulating SAPK/JNK activation in osteoblasts. J. Cell. Biochem. 2007, 100, 1146–1153. [CrossRef] [PubMed]
- 76. Shen, C.L.; Yeh, J.K.; Cao, J.J.; Tatum, O.L.; Dagda, R.Y.; Wang, J.S. Green tea polyphenols mitigate bone loss of female rats in a chronic inflammation-induced bone loss model. *J. Nutr. Biochem.* **2010**, *21*, 968–974. [CrossRef] [PubMed]
- 77. Shen, C.L.; Cao, J.J.; Dagda, R.Y.; Chanjaplammootil, S.; Lu, C.; Chyu, M.C.; Gao, W.; Wang, J.S.; Yeh, J.K. Green tea polyphenols benefits body composition and improves bone quality in long-term high-fat diet-induced obese rats. *Nutr. Res.* 2012, *32*, 448–457. [CrossRef]
- Li, Y.; Yao, J.; Han, C.; Yang, J.; Chaudhry, M.T.; Wang, S.; Liu, H.; Yin, Y. Quercetin, Inflammation and Immunity. *Nutrients* 2016, 8, 167. [CrossRef]
- 79. Hämäläinen, M.; Nieminen, R.; Asmawi, M.Z.; Vuorela, P.; Vapaatalo, H.; Moilanen, E. Effects of flavonoids on prostaglandin E2 production and on COX-2 and mPGES-1 expressions in activated macrophages. *Planta Med.* 2011, 77, 1504–1511. [CrossRef]
- Procházková, D.; Boušová, I.; Wilhelmová, N. Antioxidant and prooxidant properties of flavonoids. *Fitoterapia* 2011, 82, 513–523. [CrossRef]
- Sekher Pannala, A.; Chan, T.S.; O'Brien, P.J.; Rice-Evans, C.A. Flavonoid B-Ring Chemistry and Antioxidant Activity: Fast Reaction Kinetics. *Biochem. Biophys. Res. Commun.* 2001, 282, 1161–1168. [CrossRef]
- 82. Huang, J.; Yuan, L.; Wang, X.; Zhang, T.-L.; Wang, K. Icaritin and its glycosides enhance osteoblastic, but suppress osteoclastic, differentiation and activity in vitro. *Life Sci.* 2007, *81*, 832–840. [CrossRef]
- Cavia-Saiz, M.; Busto, M.D.; Pilar-Izquierdo, M.C.; Ortega, N.; Perez-Mateos, M.; Muñiz, P. Antioxidant properties, radical scavenging activity and biomolecule protection capacity of flavonoid naringenin and its glycoside naringin: A comparative study. J. Sci. Food Agric. 2010, 90, 1238–1244. [CrossRef] [PubMed]
- 84. Kim, W.K.; Ke, K.; Sul, O.J.; Kim, H.J.; Kim, S.H.; Lee, M.H.; Kim, H.J.; Kim, S.Y.; Chung, H.T.; Choi, H.S. Curcumin protects against ovariectomy-induced bone loss and decreases osteoclastogenesis. *J. Cell. Biochem.* **2011**, *112*, 3159–3166. [CrossRef]
- 85. Zhao, L.; Wang, Y.; Wang, Z.; Xu, Z.; Zhang, Q.; Yin, M. Effects of dietary resveratrol on excess-iron-induced bone loss via antioxidative character. J. Nutr. Biochem. 2015, 26, 1174–1182. [CrossRef]
- Huang, Q.; Gao, B.; Wang, L.; Hu, Y.Q.; Lu, W.G.; Yang, L.; Luo, Z.J.; Liu, J. Protective effects of myricitrin against osteoporosis via reducing reactive oxygen species and bone-resorbing cytokines. *Toxicol. Appl. Pharmacol.* 2014, 280, 550–560. [CrossRef]
- Cherrak, S.A.; Mokhtari-Soulimane, N.; Berroukeche, F.; Bensenane, B.; Cherbonnel, A.; Merzouk, H.; Elhabiri, M. In Vitro Antioxidant versus Metal Ion Chelating Properties of Flavonoids: A Structure-Activity Investigation. *PLoS ONE* 2016, 11, e0165575. [CrossRef] [PubMed]
- Hider, R.C.; Liu, Z.D.; Khodr, H.H. Metal Chelation of Polyphenols. Methods in Enzymology; Academic Press: Cambridge, MA, USA, 2001; pp. 190–203.
- Islam, S.; Islam, N.; Kermode, T.; Johnstone, B.; Mukhtar, H.; Moskowitz, R.W.; Goldberg, V.M.; Malemud, C.J.; Haqqi, T.M. Involvement of Caspase-3 in Epigallocatechin-3-gallate-Mediated Apoptosis of Human Chondrosarcoma Cells. *Biochem. Biophys. Res. Commun.* 2000, 270, 793–797. [CrossRef] [PubMed]
- Nakagawa, H.; Wachi, M.; Woo, J.-T.; Kato, M.; Kasai, S.; Takahashi, F.; Lee, I.S.; Nagai, K. Fenton Reaction Is Primarily Involved in a Mechanism of (–)-Epigallocatechin-3-gallate to Induce Osteoclastic Cell Death. *Biochem. Biophys. Res. Commun.* 2002, 292, 94–101. [CrossRef]
- Lin, S.-Y.; Kang, L.; Wang, C.-Z.; Huang, H.H.; Cheng, T.-L.; Huang, H.-T.; Lee, M.J.; Lin, Y.S.; Ho, M.L.; Wang, G.J.; et al. (-)-Epigallocatechin-3-Gallate (EGCG) Enhances Osteogenic Differentiation of Human Bone Marrow Mesenchymal Stem Cells. *Molecules* 2018, 23, 3221. [CrossRef]
- 92. Hsu, Y.-L.; Chang, J.-K.; Tsai, C.-H.; Chien, T.-T.C.; Kuo, P.-L. Myricetin induces human osteoblast differentiation through bone morphogenetic protein-2/p38 mitogen-activated protein kinase pathway. *Biochem. Pharmacol.* 2007, 73, 504–514. [CrossRef]
- Chen, J.R.; Lazarenko, O.P.; Wu, X.; Kang, J.; Blackburn, M.L.; Shankar, K.; Badger, T.M.; Ronis, M.J.J. Dietary-induced serum phenolic acids promote bone growth via p38 MAPK/β-catenin canonical Wnt signaling. *J. Bone Miner. Res. Off. J. Am. Soc. Bone Miner. Res.* 2010, 25, 2399–2411. [CrossRef] [PubMed]
- 94. Guo, A.J.; Choi, R.C.; Cheung, A.W.; Chen, V.P.; Xu, S.L.; Dong, T.T.; Chen, J.J.; Tsim, K.W.K. Baicalin, a flavone, induces the differentiation of cultured osteoblasts: An action via the Wnt/beta-catenin signaling pathway. *J. Biol. Chem.* **2011**, *286*, 27882–27893. [CrossRef]
- 95. Patisaul, H.B.; Jefferson, W. The pros and cons of phytoestrogens. Front. Neuroendocrinol. 2010, 31, 400–419. [CrossRef]
- Xiao, H.H.; Gao, Q.G.; Zhang, Y.; Wong, K.C.; Dai, Y.; Yao, X.S.; Wong, M.S. Vanillic acid exerts oestrogen-like activities in osteoblast-like UMR 106 cells through MAP kinase (MEK/ERK)-mediated ER signaling pathway. *J. Steroid Biochem. Mol. Biol.* 2014, 144 Pt B, 382–391. [CrossRef]
- Abdel-Naim, A.B.; Alghamdi, A.A.; Algandaby, M.M.; Al-Abbasi, F.A.; Al-Abd, A.M.; Eid, B.G.; Abdallah, H.M.; El-Halawany, A.M. Rutin Isolated from *Chrozophora tinctoria* Enhances Bone Cell Proliferation and Ossification Markers. *Oxid. Med. Cell. Longev.* 2018, 2018, 5106469. [CrossRef]

- Rassi, C.M.; Lieberherr, M.; Chaumaz, G.; Pointillart, A.; Cournot, G. Modulation of osteoclastogenesis in porcine bone marrow cultures by quercetin and rutin. *Cell Tissue Res.* 2005, 319, 383–393. [CrossRef]
- 99. Lee, D.J.; Tseng, H.C.; Wong, S.W.; Wang, Z.; Deng, M.; Ko, C.-C. Dopaminergic effects on in vitro osteogenesis. *Bone Res.* 2015, 3, 15020. [CrossRef]
- Schneider-Stock, R.; Ghantous, A.; Bajbouj, K.; Saikali, M.; Darwiche, N. Epigenetic mechanisms of plant-derived anticancer drugs. Front. Biosci. 2012, 17, 129–173. [CrossRef]
- Howitz, K.T.; Bitterman, K.J.; Cohen, H.Y.; Lamming, D.W.; Lavu, S.; Wood, J.G.; Zipkin, R.E.; Chung, P.; Kisielewski, A.; Zhang, L.L.; et al. Small molecule activators of sirtuins extend Saccharomyces cerevisiae lifespan. *Nature* 2003, 425, 191–196. [CrossRef] [PubMed]
- 102. Shakibaei, M.; Shayan, P.; Busch, F.; Aldinger, C.; Buhrmann, C.; Lueders, C.; Mobasheri, A. Resveratrol mediated modulation of Sirt-1/Runx2 promotes osteogenic differentiation of mesenchymal stem cells: Potential role of Runx2 deacetylation. *PLoS ONE* 2012, 7, e35712. [CrossRef] [PubMed]
- Tseng, P.C.; Hou, S.M.; Chen, R.J.; Peng, H.W.; Hsieh, C.F.; Kuo, M.L.; Yen, M.L. Resveratrol promotes osteogenesis of human mesenchymal stem cells by upregulating RUNX2 gene expression via the SIRT1/FOXO3A axis. J. Bone Miner. Res. Off. J. Am. Soc. Bone Miner. Res. 2011, 26, 2552–2563. [CrossRef] [PubMed]
- 104. Lee, Y.M.; Shin, S.I.; Shin, K.S.; Lee, Y.R.; Park, B.H.; Kim, E.C. The role of sirtuin 1 in osteoblastic differentiation in human periodontal ligament cells. *J. Periodontal Res.* 2011, 46, 712–721. [CrossRef] [PubMed]
- 105. Shakibaei, M.; Buhrmann, C.; Mobasheri, A. Resveratrol-mediated SIRT-1 interactions with p300 modulate receptor activator of NF-kappaB ligand (RANKL) activation of NF-kappaB signaling and inhibit osteoclastogenesis in bone-derived cells. *J. Biol. Chem.* 2011, 286, 11492–11505. [CrossRef] [PubMed]
- 106. Li, Y.; Wang, J.; Chen, G.; Feng, S.; Wang, P.; Zhu, X.; Zhang, R. Quercetin promotes the osteogenic differentiation of rat mesenchymal stem cells via mitogen-activated protein kinase signaling. *Exp. Ther. Med.* **2015**, *9*, 2072–2080. [CrossRef]
- Moon, H.J.; Ko, W.K.; Han, S.W.; Kim, D.S.; Hwang, Y.S.; Park, H.K.; Kwon, I.K. Antioxidants, like coenzyme Q10, selenite, and curcumin, inhibited osteoclast differentiation by suppressing reactive oxygen species generation. *Biochem. Biophys. Res. Commun.* 2012, 418, 247–253. [CrossRef] [PubMed]
- 108. Oryan, A.; Alidadi, S.; Moshiri, A.; Maffulli, N. Bone regenerative medicine: Classic options, novel strategies, and future directions. *J. Orthop. Surg. Res.* 2014, *9*, 18. [CrossRef] [PubMed]
- Liu, Y.; Lim, J.; Teoh, S.-H. Development of clinically relevant scaffolds for vascularised bone tissue engineering. *Biotechnol. Adv.* 2013, 31, 688–705. [CrossRef]
- 110. Zhu, G.; Zhang, T.; Chen, M.; Yao, K.; Huang, X.; Zhang, B.; Li, Y.; Liu, J.; Wang, Y.; Zhao, Z. Bone physiological microenvironment and healing mechanism: Basis for future bone-tissue engineering scaffolds. *Bioact. Mater.* **2021**, *6*, 4110–4140. [CrossRef]
- Ogay, V.; Mun, E.A.; Kudaibergen, G.; Baidarbekov, M.; Kassymbek, K.; Zharkinbekov, Z.; Saparov, A. Progress and Prospects of Polymer-Based Drug Delivery Systems for Bone Tissue Regeneration. *Polymers* 2020, 12, 2881. [CrossRef]
- 112. Cojocaru, F.-D.; Botezat, D.; Gardikiotis, I.; Uritu, C.-M.; Dodi, G.; Trandafir, L.; Rezus, C.; Rezus, E.; Tamba, B.-I.; Mihai, C.-T. Nanomaterials Designed for Antiviral Drug Delivery Transport across Biological Barriers. *Pharmaceutics* **2020**, *12*, 171. [CrossRef]
- 113. Newman, M.R.; Benoit, D.S. Local and targeted drug delivery for bone regeneration. *Curr. Opin. Biotechnol.* **2016**, 40, 125–132. [CrossRef]
- 114. Bosetti, M.; Bianchi, A.E.; Zaffe, D.; Cannas, M. Comparative in vitro study of four commercial biomaterials used for bone grafting. *J. Appl. Biomater. Funct. Mater.* **2013**, *11*, e80–e88. [CrossRef] [PubMed]
- Ceresa, C.; Fracchia, L.; Marchetti, A.; Rinaldi, M.; Bosetti, M. Injectable Scaffolds Enriched with Silver to Inhibit Bacterial Invasion in Tissue Regeneration. *Materials* 2019, 12, 1931. [CrossRef] [PubMed]
- 116. Susmita, B.; Naboneeta Sa Dishary, B. Natural medicine delivery from biomedical devices to treat bone disorders: A review. *Acta Biomater.* **2021**, *126*, 63–91. [CrossRef]
- 117. Tadic, D.; Epple, M. A thorough physicochemical characterisation of 14 calcium phosphate-based bone substitution materials in comparison to natural bone. *Biomaterials* **2004**, 25, 987–994. [CrossRef]
- 118. Yuan, H.; Yang, Z.; Li, Y.; Zhang, X.; De Bruijn, J.D.; De Groot, K. Osteoinduction by calcium phosphate biomaterials. *J. Mater. Sci. Mater. Med.* **1998**, *9*, 723–726. [CrossRef]
- 119. Ramseier, C.A.; Rasperini, G.; Batia, S.; Giannobile, W.V. Advanced reconstructive technologies for periodontal tissue repair. *Periodontology* 2000 **2012**, *59*, 185–202. [CrossRef]
- 120. Reynolds, M.A.; Aichelmann-Reidy, M.E.; Branch-Mays, G.L. Regeneration of periodontal tissue: Bone replacement grafts. *Dent. Clin. N. Am.* **2010**, *54*, 55–71. [CrossRef] [PubMed]
- Kao, R.T.; Nares, S.; Reynolds, M.A. Periodontal regeneration—Intrabony defects: A systematic review from the AAP Regeneration Workshop. J. Periodontol. 2015, 86 (Suppl. 2), S77–S104. [CrossRef]
- 122. Houde, V.; Grenier, D.; Chandad, F. Protective effects of grape seed proanthocyanidins against oxidative stress induced by lipopolysaccharides of periodontopathogens. *J. Periodontol.* **2006**, 77, 1371–1379. [CrossRef] [PubMed]
- 123. Hirasawa, M.; Takada, K.; Makimura, M.; Otake, S. Improvement of periodontal status by green tea catechin using a local delivery system: A clinical pilot study. *J. Periodontal Res.* 2002, 37, 433–438. [CrossRef] [PubMed]
- 124. Kushiyama, M.; Shimazaki, Y.; Murakami, M.; Yamashita, Y. Relationship between intake of green tea and periodontal disease. *J. Periodontol.* **2009**, *80*, 372–377. [CrossRef] [PubMed]

- 125. Diaz-Rodriguez, P.; Sánchez, M.; Landin, M. Drug-Loaded Biomimetic Ceramics for Tissue Engineering. *Pharmaceutics* 2018, 10, 272. [CrossRef] [PubMed]
- 126. Morra, M.C.C.; Bollati, D.; Iviglia, G. Inventor Compositions for Filling Bone and Periodontal Defects WO 2015/014872, 5 February 2015.
- 127. Cazzola, M.; Corazzari, I.; Prenesti, E.; Bertone, E.; Vernè, E.; Ferraris, S. Bioactive glass coupling with natural polyphenols: Surface modification, bioactivity and anti-oxidant ability. *Appl. Surf. Sci.* **2016**, *367*, 237–248. [CrossRef]
- 128. Zhou, K.; Ren, X.; Zhao, M.; Mei, X.; Zhang, P.; Chen, Z.; Zhu, X. Promoting proliferation and differentiation of BMSCs by green tea polyphenols functionalized porous calcium phosphate. *Regener. Biomater.* **2018**, *5*, 35–41. [CrossRef]
- 129. Preethi Soundarya, S.; Sanjay, V.; Haritha Menon, A.; Dhivya, S.; Selvamurugan, N. Effects of flavonoids incorporated biological macromolecules based scaffolds in bone tissue engineering. *Int. J. Biol. Macromol.* **2018**, *110*, 74–87. [CrossRef] [PubMed]
- Joseph, J.; Sundar, R.; John, A.; Abraham, A. Phytochemical Incorporated Drug Delivery Scaffolds for Tissue Regeneration. *Regen. Eng. Transl. Med.* 2018, 4, 167–176. [CrossRef]
- 131. Rekulapally, R.; Udayachandrika, K.; Hamlipur, S.; Nair, A.S.; Pal, B.; Singh, S. Tissue engineering of collagen scaffolds crosslinked with plant based polysaccharides. *Prog. Biomater.* **2021**, *10*, 29–41. [CrossRef]
- 132. Rambhia, K.J.; Ma, P.X. Controlled drug release for tissue engineering. J. Control. Release 2015, 219, 119–128. [CrossRef]
- 133. Santin, M.; Morris, C.; Standen, G.; Nicolais, L.; Ambrosio, L. A new class of bioactive and biodegradable soybean-based bone fillers. *Biomacromolecules* 2007, *8*, 2706–2711. [CrossRef]
- 134. Merolli, A.; Nicolais, L.; Ambrosio, L.; Santin, M. A degradable soybean-based biomaterial used effectively as a bone filler in vivo in a rabbit. *Biomed. Mater.* 2010, *5*, 015008. [CrossRef] [PubMed]
- 135. Wang, W.; Sun, L.; Zhang, P.; Song, J.; Liu, W. An anti-inflammatory cell-free collagen/resveratrol scaffold for repairing osteochondral defects in rabbits. *Acta Biomater.* **2014**, *10*, 4983–4995. [CrossRef] [PubMed]
- 136. Li, Y.; Dånmark, S.; Edlund, U.; Finne-Wistrand, A.; He, X.; Norgård, M.; Blomén, E.; Hultenby, K.; Andersson, G.; Lindgren, U. Resveratrol-conjugated poly-ε-caprolactone facilitates in vitro mineralization and in vivo bone regeneration. *Acta Biomater.* 2011, 7, 751–758. [CrossRef] [PubMed]
- 137. Kamath, M.S.; Ahmed, S.S.; Dhanasekaran, M.; Santosh, S.W. Polycaprolactone scaffold engineered for sustained release of resveratrol: Therapeutic enhancement in bone tissue engineering. *Int. J. Nanomed.* **2014**, *9*, 183–195. [CrossRef]
- 138. Riccitiello, F.; De Luise, A.; Conte, R.; D'Aniello, S.; Vittoria, V.; Di Salle, A.; Calarco, A.; Peluso, G. Effect of resveratrol release kinetic from electrospun nanofibers on osteoblast and osteoclast differentiation. *Eur. Polym. J.* **2018**, *99*, 289–297. [CrossRef]
- 139. Wang, C.C.; Wang, C.H.; Chen, H.C.; Cherng, J.H.; Chang, S.J. Combination of resveratrol-containing collagen with adipose stem cells for craniofacial tissue-engineering applications. *Int. Wound J.* **2018**, *15*, 660–672. [CrossRef]
- 140. Wang, Z.; Li, C.; Xu, J.; Wang, K.; Lu, X.; Zhang, H.; Qu, S.; Zhen, G.; Ren, F. Bioadhesive Microporous Architectures by Self-Assembling Polydopamine Microcapsules for Biomedical Applications. *Chem. Mater.* **2015**, *27*, 848–856. [CrossRef]
- Ko, E.; Yang, K.; Shin, J.; Cho, S.-W. Polydopamine-Assisted Osteoinductive Peptide Immobilization of Polymer Scaffolds for Enhanced Bone Regeneration by Human Adipose-Derived Stem Cells. *Biomacromolecules* 2013, 14, 3202–3213. [CrossRef]
- Lee, J.S.; Lee, J.S.; Lee, M.S.; An, S.; Yang, K.; Lee, K.; Yang, H.S.; Lee, H.; Cho, S.W. Plant Flavonoid-Mediated Multifunctional Surface Modification Chemistry: Catechin Coating for Enhanced Osteogenesis of Human Stem Cells. *Chem. Mater.* 2017, 29, 4375–4384. [CrossRef]
- 143. Pasche, S.; Vörös, J.; Griesser, H.J.; Spencer, N.D.; Textor, M. Effects of ionic strength and surface charge on protein adsorption at PEGylated surfaces. *J. Phys. Chem. B* 2005, *109*, 17545–17552. [CrossRef] [PubMed]
- 144. Verma, A.; Stellacci, F. Effect of surface properties on nanoparticle-cell interactions. Small 2010, 6, 12–21. [CrossRef] [PubMed]
- 145. Bertrand, N.; Wu, J.; Xu, X.; Kamaly, N.; Farokhzad, O.C. Cancer nanotechnology: The impact of passive and active targeting in the era of modern cancer biology. *Adv. Drug Deliv. Rev.* **2014**, *66*, 2–25. [CrossRef] [PubMed]
- 146. He, L.; Deng, D.; Zhou, X.; Cheng, L.; ten Cate, J.M.; Li, J.; Li, X.; Crielaard, W. Novel tea polyphenol-modified calcium phosphate nanoparticle and its remineralization potential. *Biomed. Mater. Res. Part B Appl. Biomater.* 2015, 103, 1525–1531. [CrossRef] [PubMed]
- 147. Wang, M.; Wang, L. Plant polyphenols mediated synthesis of gold nanoparticles for pain management in nursing care for dental tissue implantation applications. *J. Drug Deliv. Sci. Technol.* **2020**, *58*, 101753. [CrossRef]
- 148. Felice, F.; Zambito, Y.; Belardinelli, E.; D'Onofrio, C.; Fabiano, A.; Balbarini, A.; Di Stefano, R. Delivery of natural polyphenols by polymeric nanoparticles improves the resistance of endothelial progenitor cells to oxidative stress. *Eur. J. Pharm. Sci.* **2013**, *50*, 393–399. [CrossRef] [PubMed]
- 149. Luisa, D.P.-A.M.; Griselda, R.-M.; Valentín, M.-L.; Carmina, O.-S.; Cristina, V.-M.; J., M.; Maykel, G.T.; David, Q.G.; Sánchez-Sánchez, R.; Leyva-Gómez, G. Curcumin-loaded poly-ε-caprolactone nanoparticles show antioxidant and cytoprotective effects in the presence of reactive oxygen species. *J. Bioact. Compat. Polym.* 2020, 35, 270–285. [CrossRef]
- 150. Malathy, S.; Iyer, P.R. Naringin Loaded Chitosan Nanoparticle for Bone Regeneration: A Preliminary in vitro Study. *J. Nanomed. Nanotechnol.* **2018**, *9*, 1–7.