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A diagnostic study on *folium* and *orchil* dyes with non-invasive and micro-destructive methods

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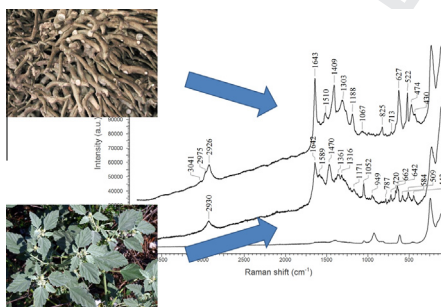
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HIGHLIGHTS

- Non-invasive and micro-invasive techniques used for folium/orchil identification.
- Diagnostic information on these dyes strongly increased.
- Historical reconstructions performed in order to have reliable standards.
- Evidence that bromine is not a key marker exclusive for Tyrian purple.
- Most folium spectral features presented for the first time in a scientific work.

GRAPHICAL ABSTRACT



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ABSTRACT

Folium and *orchil* are dyes of vegetal origin. *Folium* is obtained from *Chrozophora tinctoria* (L.) A. Juss., whereas *orchil* is obtained from *Roccella* and other genera of lichens. These dyes were used in the past to impart purple hue to paintings and textiles as substitutes for the more prized Tyrian purple dye, obtained from shellfish. Despite several citations in ancient technical treatises dating back at least to the Greek-Roman age, the identification of these dyes in artworks is rare. In the case of *folium*, an additional drawback is that its composition is presently unknown.

In this work different non-invasive (FT-IR, FT-Raman, fibre optic reflectance spectrophotometry, spectrofluorimetry, X-ray fluorescence spectrometry) and micro-invasive (surface enhanced Raman spectroscopy, matrix assisted laser desorption/ionisation-time of flight-mass spectrometry, inductively coupled plasma-mass spectrometry) techniques were used in order to increase the diagnostic information available on these dyes. Measurements were carried out on the dyes extracted from raw materials and on painted or dyed parchments. The possibility to distinguish between *folium* and *orchil* by chemical analysis is discussed.

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Introduction

The names *folium* and *orchil* are used today to indicate two kinds of dyes of vegetal origin, obtained from *Chrozophora tinctoria* (L.) A. Juss. and from several genera of lichens, respectively. These dyes have been in use for a long time to impart purple hue to

artworks, either dyed or painted; they were in fact mostly used as substitutes for the more priced Tyrian purple, the famous dye obtained from shellfish. For several centuries, though, folium and orchil were hardly considered as different materials, as the historical terminology used for their description in the technical and artistic literature was confusing. In many medieval manuscripts, similar names were given to lichen dyes and to dyes obtained from *Chroophora* species. As an example, lichens of *Roccella* species were also known as *tournesol* which is a traditional name used for *C. tinctoria* (L.) A. Juss. plant. It is only in the 19th century that ambiguity was resolved [1,2] and the different origin of these dyes was ascertained.

The term *orchil* refers primarily to red and purple dyes obtained from various lichen species upon fermentation in ammonia. Although the most renowned orchil-producing lichen species are those of *Roccella* genus (e.g. *Roccella tinctoria*, *Roccella montagnei*, *Roccella fuciformis*, etc.), *Dendrographa*, *Diploschistes*, *Evernia*, *Lasallia*, *Lecanora*, *Ochrolechia*, *Parmelia*, *Pertusaria*, *Umbilicaria* and *Variolaria* genera are other sources of orchil [3–5]. *Orchil* has been intended as a general name for purple lichen dyes at least since the end of 19th century. Nevertheless, in the past this name (and the dialectic variants *archil*, *orchall*, *orseille*, *oricello*, etc.) referred mainly to the dye obtained from *Roccella* species, while different names were used for regional varieties: *cudbear* was the name given to a purple dye obtained from *Lecanora* species widely common in Scotland (e.g. *Ochrolechia tartarea*), while in Scandinavia a similar dye, known as *korkje*, was extracted also from *Lasallia* and *Umbilicaria* lichens; another purple dye, obtained from lichen species living in Central France, was known as *parelle*. Purple dyes from lichens were in use at least since Greek-Roman times: literary citations from Theophrastus, Dioskurides and Pliny the Elder are known [6,7] which highlight their role as a substitute of Tyrian purple in dyeing. Pliny the Elder, in particular, suggested that orchil could be used to dye wool textiles as a background where a small amount of Tyrian purple was subsequently applied, a procedure known as top-dyeing [8]. Moreover, several recipes in the Greek manuscript known as *Stockholm Papyrus* (3rd century A.D.) recommended the use of dyes obtained from lichens to imitate purple [9]. As for painting, medieval treatises cited orchil as a suitable colourant, such as the manuscript *Ut auro scribatur* [10] where its use is suggested as a paint (not as a dye) to impart purple colour to parchment in purple codices. The composition of orchil, though complex, has been elucidated and reviewed in several studies [11,12]. Lichens contain depsides and depsidones which are precursors of the dye. Precursors can change from species to species but, in the case of purple-generating species, the main precursors are erythrin, evernic acid, gyrophoric acid and lecanoric acid; after extraction, these compounds are hydrolysed to orsellinic acid and decarboxylated to colourless orcinol, which is oxidised to orcein upon introduction of ammonia. Orcein is actually made up of a mixture of phenoxazone derivatives such as hydroxyorceins, amino-orceins and amino-orceimines.

Folium is extracted from *C. tinctoria* (L.) A. Juss, a plant known as *turnsole* or *morella*, native of coastal Mediterranean countries. Interestingly, the German name for turnsole has been for long *lackmuskraut*, a term meaning litmus-herb, where litmus is another dye produced from *R. tinctoria* lichens differing from orchil in reason of its polymeric structure. Literary citations concerning the use of *folium* in artworks are found later than those concerning orchil. The first recipes for the preparation of folium date back to 11th century A.D. but its use in painting can be probably referred to the early Middle Ages [13] since it is cited in the 9th century *Map-pae Clavicular* treatise [14]. However, it is highly probable that turnsole was already in use in Roman times. Pliny the Elder [8] in his *Naturalis historia*, book XXII, chapter 29, mentions in fact a vegetal species which he called *Heliotropium tricoccum*. This may indicate

the presence of three cells in the capsule of the plant, as the characteristic tri-lobed fruits yielding folium dye. Among others, Theophilus in his famous *De diversis artibus* treatise [15] and the anonymous author of the *De arte illuminandi* treatise [16] highlighted the fact that this plant can produce a red, violet or blue dye if berries are extracted respectively with an acid, neutral or alkaline solution: the so-called *folium rubeum*, *folium purpureum* and *folium saphireum*. The name folium, however, is historically referred to the purple-violet phase. The scientific knowledge on the composition of folium is relatively scarce if compared to orchil. Early studies [17] suggested that, according to its properties of changing colour on varying pH, folium could contain anthocyanin compounds. Other studies [18–20] suggested instead the similarity between folium and orchil from a compositional point of view. Guineau showed results from time-of-flight mass spectrometry (ToF-MS) analysis in his detailed historical and diagnostic study [13], which highlighted the presence of orcinol, a compound also present in lichen dyes.

Identifications of folium and orchil on artworks are rare, in particular when non-invasive analyses are considered. Orchil was identified by Clementi et al. [21,22] by means of fluorescence spectroscopy in some Renaissance tapestries and in purple details of the miniatures of the *Book of Kells* [23], a famous 8th–9th century A.D. manuscript. The same authors identified orchil on the parchment of the *Bible de Théodulfe* (9th century) using fluorescence spectroscopy and subtracted shifted Raman spectroscopy, [24] and Eveno et al. [25] gained a similar identification by HPLC. Aceto et al. [26] analysed the parchment of the *Codex Brixianus*, a 6th century A.D. purple manuscript, using UV-visible diffuse reflectance spectrophotometry, spectrofluorimetry and X-ray fluorescence spectrometry and suggested that both orchil and folium could be present. Recently Bicchieri [27] identified orchil by means of UV-visible diffuse reflectance spectrophotometry on the parchment of the precious *Codex Rossanensis*, another 6th century A.D. purple manuscript. Finally, the identification of litmus was carried out by Baraldi et al. [28] with Raman spectroscopy on a 17th century painted table. As regards folium, the number of identifications is very limited since it can be circumscribed to the pioneering work by Guineau [13] in which the author identified the dye in some 9th–11th century manuscripts by means of UV-visible diffuse reflectance spectrophotometry, to the tentative identification on the *Sinope Gospels* (a 6th century A.D. purple manuscript) by means of GC-MS by Thomas and Flieder [18] and to the tentative attribution to folium of blue areas in the de Brécy *Madonna and Child* tondo painting, analysed with FT-Raman spectroscopy by Edwards and Benoy [29].

From the artistic point of view, the use of folium and orchil in painting is certainly suitable for obtaining a range of hues from red to blue through purple, as described in several medieval artistic treatises. Therefore, despite the very low number of identifications on artworks, the number of instances in which these dyes could have been used is possibly much larger than the number of actual identifications. Moreover, the overview on the literature reported above highlights that the diagnostic information concerning these dyes is very limited or, as in the case of folium, almost absent. In the present work we aim to increase the diagnostic information available for the detection of folium and orchil by means of the spectroscopic techniques that are normally used in the analysis of painted artworks, with particular concern to illuminated manuscripts; therefore in this study folium and orchil have been subjected to a deep analytical investigation with a particular focus on the use of a non-invasive or a micro-invasive approach. The following non-invasive techniques were considered: Fourier transform Raman spectroscopy (FT-Raman), Fourier transform infrared spectrophotometry (FT-IR) both in transmission mode and in attenuated total reflection (ATR) mode, spectrofluorimetry,

UV-visible diffuse reflectance spectrophotometry with optical fibres (FORS) and X-ray fluorescence spectrometry (XRF). In order to assess the accuracy of the non-invasive approach and to gain further information on the dyes, micro samples were analysed by means of surface enhanced Raman spectroscopy (SERS) and matrix-assisted laser desorption-ionisation time-of-flight mass spectrometry (MALDI-ToF MS). Finally, additional elemental analysis on lichens and *C. tinctoria* (L.) A. Juss. samples was performed by means of inductively coupled plasma-mass spectrometry (ICP-MS). For all the above cited techniques, measurements were carried out both on raw powders and on standard paints and dyes on parchment; the results were compared with non-invasive analyses on some purple and violet painted areas on illuminated manuscripts.

Materials and methods

FT-IR spectrophotometry

Solid-state FT-IR spectra were collected on a FT-IR Bruker Equinox 55 spectrophotometer (Bruker Optics Inc., Ettlingen, Germany), at 2 cm^{-1} resolution, in anhydrous KBr discs (average of 50–100 scans). Measurements in ATR mode were carried out with a Thermo Scientific Nicolet (Madison, Wisconsin, USA) iN10™ model FT-IR spectrometer equipped with an iZ10 external module bearing a Smart iTR™ diamond ATR Sampling Accessory.

FT-Raman spectroscopy

FT-Raman measurements were performed with a Bruker Vertex 70 spectrometer (Bruker Optics Inc., Ettlingen, Germany) equipped with a RAM II accessory, a 1064 nm Nd/YAG laser source and a Ge diode detector. Spectral parameters were as follows: 100 mW laser power, 500 scans, and 4 cm^{-1} resolution.

Surface enhanced Raman spectroscopy (SERS)

SERS analysis was performed by means of Ag colloidal pastes, according to the procedure described by Idone et al. [30]. All the materials employed (e.g. nitric acid, hydrochloric acid, methanol, formic acid, silver nitrate and sodium citrate dihydrate) were purchased from Carlo Erba reagents (Arese, Italy), while ultra high quality (UHQ) water was obtained by a Millipore (Darmstadt, Germany) Direct-q 3 system. Citrate-reduced Ag nanoparticles were synthesized by modifying the procedure of Lee and Meisel [31]. Raman measurements were performed with a Renishaw (Stonehouse, Great Britain) inVia micro-Raman spectrometer equipped with a 633 nm laser, 1800 lines/mm grating and a 100x Leica (Wetzlar, Germany) microscope objective to focus the laser beam onto the sample. Power at the samples was kept very low (never exceeding $300\text{ }\mu\text{W}$) by a series of neutral density filters in order to avoid any thermal damage. Analysis of samples of dyed parchment was performed both directly and upon extraction of the dye. In the first case, $0.5\text{ }\mu\text{L}$ of silver colloidal paste were dropped on the parchment. In the last case, $50\text{ }\mu\text{L}$ of concentrated formic acid were added to a 1 mm^2 fragment of parchment and kept at $40\text{ }^\circ\text{C}$ for three hours; then, $2\text{ }\mu\text{L}$ of extract were mixed with $2\text{ }\mu\text{L}$ of Ag colloidal paste.

UV-visible diffuse reflectance spectrophotometry with optic fibres (FORS)

FORS analysis was performed with an Avantes (Apeldoorn, The Netherlands) AvaSpec-ULS2048XL-USB2 model spectrophotometer and an AvaLight-HAL-S-IND tungsten halogen light source; detector and light source were connected with fibre optic cables to an

FCR-7UV200-2-1.5x100 probe. In this configuration, light is sent and retrieved with a single fibre bundle positioned at 45° with respect to the surface normal, in order not to include specular reflectance. The spectral range of the detector was 200–1160 nm; depending on the features of the monochromator (slit width $50\text{ }\mu\text{m}$, grating of UA type with 300 lines/mm) and of the detector (2048 pixels), the best spectra resolution was 2.4 nm calculated as FWHM (full width at half maximum). Diffuse reflectance spectra of the samples were referenced against the WS-2 reference tile provided by Avantes and guaranteed to be reflective at 98% or more in the investigated spectral range. Blank correction was not efficient on both the extremes of the spectral range, therefore the regions 200–250 and 900–1160 were not considered in the discussion. The diameter of the investigated area on the sample was 1 mm. In all measurements the distance between the probe and the sample was kept constant at 1 mm. The instrumental parameters were as follows: 10 ms integration time, 100 scans for a total acquisition time of 1.0 s for each spectrum. The system was managed by means of AvaSoft v. 8™ dedicated software, running under Windows 7™.

Spectrofluorimetry

An Ocean Optics (Dunedin, Florida, USA) Jaz model spectrophotometer was employed to record molecular fluorescence spectra. The instrument is equipped with a 365 nm Jaz-LED internal light source; a QF600-8-VIS/NIR fibre fluorescence probe is used to drive excitation light on the sample and to recover the emitted light. The spectrophotometer works in the range 191–886 nm; according to the features of the monochromator ($200\text{ }\mu\text{m}$ slit width) and detector (2048 elements), the spectral resolution available is 7.6 nm calculated as FWHM. The investigated area on the sample is 1 mm in diameter. In all measurements the sample-to-probe distance was kept constant to 1 mm (corresponding to the focal length of the probe) with aid of a small black cylinder inserted on top of the probe, which also shields external light. Instrumental parameters were as follows: 2 s integration time, 3 scans for a total acquisition time of 6 s for every spectrum. The system is managed with SpectraSuite™ software under Windows 7™.

Matrix-assisted laser desorption-ionisation time-of-flight mass spectrometry (MALDI-ToF MS)

MALDI-ToF-MS experiments were performed in positive-ion mode on a time of flight (ToF) mass spectrometer Voyager DE-PRO model (Applied Biosystems Italia, Monza, Italy). Desorption/ionisation was obtained by using a 337-nm nitrogen laser and the accelerating voltage of +20 kV. To obtain good resolution and signal-to-noise (S/N) ratios, laser power was adjusted slightly above the threshold and each mass spectrum was generated by averaging 100 laser pulses. The calibration of mass spectra was performed externally using Sequazime Peptide Mass Standard, Calibration mixture 1 (AB Sciex Italia, Brugherio, Italy) and matrix peaks. Sample preparation was carried out as follows:

- Matrix solution: 5 g/L of sinapinic acid solution was obtained with a 1:1 volumetric ratio of acetonitrile to 0.1% trifluoroacetic acid in ultrapure water.
- Sample solution: 2 mg of dye powder were dissolved in 0.5 ml of matrix diluent (1:1 acetonitrile/water with 0.1% trifluoroacetic acid).
- Parchment samples: analysis of samples of dyed and painted parchment was performed after extraction of the dye with $50\text{ }\mu\text{L}$ of concentrated formic acid.

The same amounts of matrix and sample mother solution were mixed and then deposited as 1 µl drop on a stainless steel 96-well target and allowed to dry before MALDI-ToF-MS analysis.

All organic solvents (HPLC grade), formic acid and ultrapure water were purchased from VWR, Milan, Italy). Sinapinic acid recrystallized matrix was purchased from LaserBio Labs (Sophia-Antipolis, France).

X-ray fluorescence spectrometry (XRF)

XRF measurements were performed with an EDXRF Thermo (Waltham, USA) NITON spectrometer XL3T-900 GOLDD model, equipped with an Ag tube (max. 50 kV, 100 µA, 2 W), a large area SDD detector, energy resolution of about 136 eV at 5.9 keV. Analysed spot had an average diameter of 3 or 8 mm and was focused by a CCD camera, with a working distance of 2 mm. Total time of analysis was 240 s. The instrument is held in position with a moving stage allowing micrometric shifts, in order to reach the desired probe-to-sample distance; the stage is laid on a tripod. The obtained spectra have been processed with the commercial software WinAxil, derived by the academic software QXAS from IAEA.

Inductively coupled plasma-mass spectrometry (ICP-MS)

ICP-MS was used to determine the amount of bromine and iodine in raw materials, i.e. in scraps of lichens and in parts of *C. tinctoria* (L.) A. Juss. fruits. For this task, 50 mg of sample were subjected to microwave assisted acid digestion with 2 ml of concentrated HNO₃ TraceSelect grade (Sigma-Aldrich, Milan, Italy). The dissolved sample was diluted to 100 ml with high purity water. The analytical conditions used for ICP-MS were the same as described in [32]; ⁷⁹Br, ⁸¹Br and ¹²⁹I were the isotopes used for quantification.

Extraction of the dyes from raw materials

Folium was obtained by extraction of fruits of *C. tinctoria* (L.) A. Juss. from Turkey in cold water at neutral pH for 1 h; extract was filtered and allowed to dry. Orchil, following the indications by Kok [3], was obtained by extraction of scraps of *R. tinctoria* from

Canary Islands in 30% v/v ammonia, with frequent stirring to favour introduction of air and oxidation of orcinol to orcein; after 3 weeks the extract was filtered and left at room temperature until dryness.

Preparation of painted and dyed parchment

Paints and dyes of folium and orchil were prepared following the recipes indicated in ancient treatises and applied on parchment. In particular, a solution with 1 g/ml in gum Arabic and 2 g/ml in sucrose was used as painting medium. Folium and orchil (ca. 0.25 g/ml) were dissolved in the medium and applied on parchment by means of a brush. Parchment was dyed according to the procedures employed for dyeing textiles with mordant dyes. The parchment was soaked for 1 h in a solution containing the dye and alum (30% by weight with respect to the weight of parchment).

Results and discussion

The results will be presented and discussed indicating whether they refer to raw powdered dyes or to painted/dyed samples. It must be considered that all the spectral information presented results from complex and, with particular concern to folium, largely unexplored matrices, which actually contain the molecules responsible for the colour but can also contain several other metabolites or compounds resulting from the production of the dyes; it is possible, therefore, that the spectral profiles from pure colouring molecules (e.g. orceins in the case of orchil) could differ from those of the extracted dyes. Moreover, as stated before, this work aims at identifying folium and orchil on artworks rather than characterising their structure or composition, which will be the subject of a subsequent work.

FT-IR spectrophotometry analysis

The FT-IR spectra of raw orchil (above) and folium (bottom) are reported in Fig. 1; for sake of comparison, an offset has been applied along Y axis. The lichen dye is provided with strong hydrophilic features, noticed by three main broad absorptions set at ca. 3400, 1630 and 1000 cm⁻¹, which are diagnostic of hydroxyl

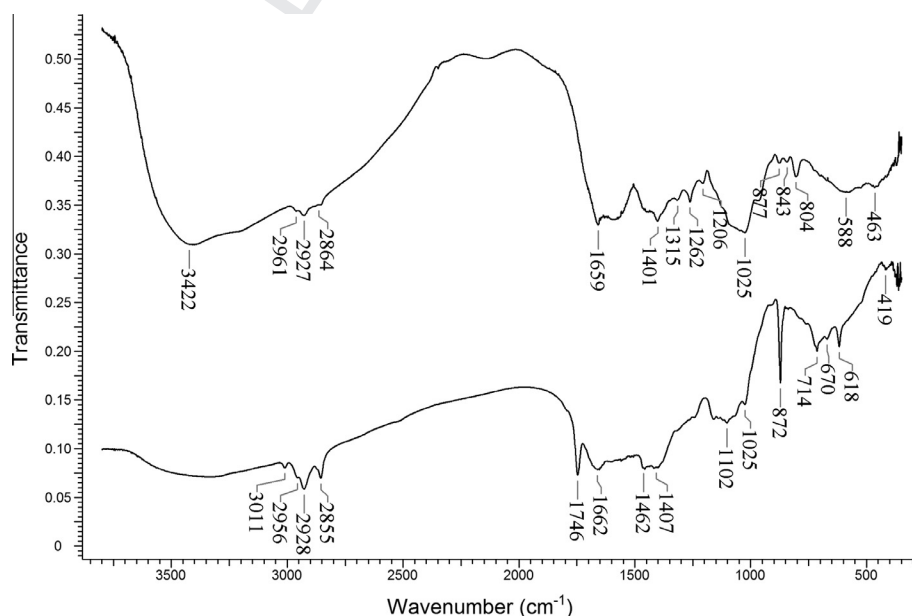


Fig. 1. FT-IR spectra in transmittance coordinates of folium (bottom line) and orchil (top line).

390 –OH moiety, whilst folium retains a more lipophilic nature, proba-
391 bly determined by an amphiphilic structure, as suggested by the
392 fact that folium is easily extracted in water. In both spectral pat-
393 terns, peaks in the fingerprint regions can be correlated to the nor-
394 mal vibrational modes of aromatic and polycyclic aromatic skeletal
395 frames [33,34]. In particular, the 3050–3000 cm^{-1} region is pat-
396 terned with the –CH aromatic stretching modes, the 1650–
397 1580 cm^{-1} region with the sp^2 C=C aromatic stretching modes,
398 the 1450–1200 cm^{-1} region with the coupled C=C stretching and
399 in-plane CH bending modes and the 900–700 cm^{-1} region with
400 the out-of-plane CH bending modes. In this perspective, reported
401 modes can be observed in both spectral patterns. Noteworthy,
402 the out-of-plane –CH deformation modes, i.e. the $\gamma(\text{CH})$ modes,
403 are provided with very strong infrared intensities (the highest or
404 among the highest of the entire aromatic pattern), and they are
405 by far the most distinctive region of the spectra [33,35,36]. Hence,
406 the pattern of folium, featured with a strong sharp absorption at
407 872 cm^{-1} , can be associated with a mainly aromatic molecular
408 frame, whilst the pattern of orchil may be correlated to a mixture
409 of different products. In this pattern, sp^2 aromatic –CH stretching
410 modes, set at 3000 cm^{-1} or higher wavenumbers, are not observed.
411 However, the experimental result is coherent with the relative
412 intensities of $\gamma(\text{CH})$ modes, embedded in a spectral profile with
413 the very strong broad absorptions determined by the highly-polar
414 –OH groups. Therein, the peak at 1659 cm^{-1} may be correlated to a
415 conjugated ketonic moiety. The weak shoulder set at about
416 3200 cm^{-1} can be determined by ring-conjugated N–H modes.
417 The –CH stretching mode peaks under 3000 cm^{-1} , with the related
418 –CH in-plane deformation modes in the 1450–1200 cm^{-1} spectral
419 region, are associated to aliphatic ring-substituents. In this context,
420 the overlapping broad profile of peaks between 1450 and
421 1200 cm^{-1} supports the presence of a mixture of different prod-
422 ucts. As a whole, the comprehensive spectral pattern matches in
423 appropriate results with orcein-like molecular frames that can be
424 actually extracted and purified from lichen substrates [11]. A sim-
425 ilar substance is provided by folium, with less polar groups and
426 with specific hydrophobic features, although a peak at 1746 cm^{-1}
427 is observed which can be determined by ketonic moieties.

428 The reported spectrum of orchil is substantially in agreement
429 with that shown by Beecken et al. [11] while the spectrum of
430 folium, to the authors' knowledge, is the first ever published.

431 Neither painted nor dyed samples of folium and orchil on
432 parchment yielded a significant FT-IR spectrum in transmittance
433 and ATR modes. Indeed, the corresponding spectra (not reported)
434 were dominated by the spectral features of the parchment and it
435 was not possible to recognise any useful features from the dyes.

FT-Raman spectroscopy analysis

437 FT-Raman spectra of raw orchil (above) and folium (bottom) are
438 reported in Fig. 2; for sake of comparison, an offset has been
439 applied along Y axis. Both Raman patterns are generally coherent
440 with results obtained from infrared spectroscopy. In the orchil pat-
441 tern, different partially overlapping peaks fill the 1450–1200 cm^{-1}
442 region of in-plane –CH deformation modes, which can be associ-
443 ated to a coexisting mixture of different products. As in the infrared
444 pattern, in the FT-Raman spectrum of orchil –CH aromatic stretch-
445 ing mode peaks are barely observed. Noteworthy, the strong peaks
446 at 2926 and at 2927 cm^{-1} (with a shoulder at 2864 cm^{-1}) for orchil
447 and folium, respectively, supported by the strong signals observed
448 in the in-plane –CH deformation mode region, have to be corre-
449 lated to aliphatic ring-substituents. In both spectra, sharp peaks
450 (at 1075 and at 1087 cm^{-1}) can be associated to –C–O–C– ether
451 groups, whilst weak peaks at 3250 cm^{-1} can be associated to –
452 NH groups.

453 It is difficult to find in the literature a suitable comparison for
454 the here reported spectra. The spectrum of orchil shows limited
455 resemblance to those reported by Edwards et al. [37–39] in their
456 works on the characterisation of substances obtained from lichens
457 (lecanoric acid, parietin, gyrophoric acid, etc.) but these com-
458 pounds are actually the precursors of orchil and therefore they
459 may not be a correct reference to compare with. A more proper
460 comparison can be carried out with the spectrum of orcein recently
461 published by Zaffino et al. [40] which shows similar spectral fea-
462 tures. In the case of folium, there is no reliable reference to com-
463 pare with; the spectrum of a blue area, tentatively attributed to
464 folium in a work by Edwards et al. [29], largely differs from those
465 obtained here.

466 Also in this case, the spectra of painted and dyed samples were
467 dominated by the spectral features of parchment, although very
468 few characteristic features of the dyes could be singled out. Peaks
469 occurring at 1271 and 1248 cm^{-1} , which can be attributed to
469

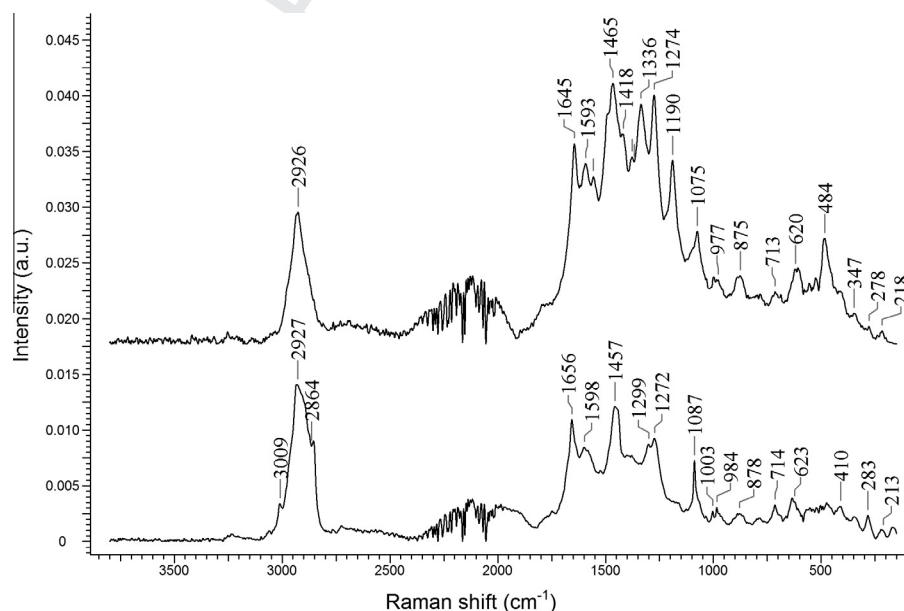


Fig. 2. FT-Raman spectra of folium (bottom line) and orchil (top line).

470 in-plane ring stretching and to $-\text{CO}$ aromatic ether stretching
471 respectively, appear in folium, orchil and parchment itself, but in
472 the case of orchil the peak at 1271 cm^{-1} is definitely higher. In
473 the spectrum of folium a distinctive peak is the one occurring at
474 975 cm^{-1} , due to ring breathing or to $-\text{CH}$ out-of-plane bending;
475 this peak is weak in orchil and it is absent in the spectrum of
476 parchment.

477 **SERS analysis**

478 The SER spectra obtained from application of Ag colloidal pastes
479 to the raw powdered dyes are shown in Fig. 3. They support FT-
480 Raman results with some differences. In the spectrum of orchil
481 the modes at 1643, 1409, 1312, 626 and 522 cm^{-1} are clearly
482 enhanced, while the SER spectrum of folium appears more similar
483 to its FT-Raman spectrum. Noteworthy, the deposition of dye mole-
484 cules on the heterogeneous surface of colloidal Ag nanoparticles
485 determines broad overlapping peaks and a smoothened vibrational
486 profile [41].

487 Similar results were obtained by analysing raw dyes and sam-
488 ples of dyed parchment, either as such or upon extraction of the
489 dye with formic acid. Silver colloidal pastes directly spread onto
490 the parchment dyed with orchil were effective in enhancing the
491 signals of the dye, even though their intensity was lower than
492 those observed for the powder sample. In particular, peaks below
493 800 cm^{-1} were more intense, while weaker signals were found in
494 the $1000\text{--}1700\text{ cm}^{-1}$ region. The position of the peaks was in quite
495 good accordance with that of powder orchil: 419 (w), 461 (w), 476
496 (w), 522 (s), 602 (sh), 619 (s), 630 (sh), 658 (w), 818 (m), 1186 (w),
497 1410 (m), 1526 (w) and 1645 (m). SER spectra recorded on the
498 parchment dyed with folium presented the stronger signals at
499 1467, 1483 (sh) and 1641 cm^{-1} , with medium peaks at 503, 572
500 and 640 cm^{-1} and weak peaks at 370, 583 (sh), 595 (sh), 684,
501 1000, 1033, 1068, 1117, 1289, 1319, 1555 cm^{-1} , most of which cor-
502 responding to the SER peaks observed for powdered folium. SERS
503 analysis allowed to establish a reliable micro-invasive and micro-
504 destructive procedure for identification of these dyes.

505 The SER spectrum of orchil is in good agreement with those
506 reported by Leona et al. [42] and by Doherty et al. [43], while some
507 differences arise upon comparison with the one reported by Rosi
508 et al. [24] which was obtained, at any rate, with Subtracted Shifted

509 Raman spectroscopy. On the other hand, the only comparison
510 available in the literature for folium is the above mentioned FT-
511 Raman spectrum obtained by Edwards from the blue areas in de
512 Brécy *Madonna and Child* tondo painting [29]. To the authors'
513 knowledge, this is the very SER spectrum of folium ever published,
514 together with the FT-Raman spectrum reported above.

515 **FORS analysis**

516 FORS spectra were recorded in reflectance mode and trans-
517 formed in $\text{Log}(1/R)$ in order to obtain *apparent absorbance* coordi-
518 nates which better highlight the absorption spectral features
519 (Fig. 4). The spectra from painted and dyed parchment samples
520 were identical, as already evidenced before [26]. FORS spectra of
521 folium and orchil are rather similar and characterised by a large
522 absorption band structured in two sub-bands. The absorption max-
523 ima of folium are located at ca. 546 and 577 nm, while those of
524 orchil occur at ca. 544 and 588 nm.

525 As regards folium, the spectral features are in good agreement
526 with those reported by Guineau [13] and by Clarke [44] which
527 are, to the authors' knowledge, the only references available in
528 the literature. Noteworthy, folium extracts with metal ions (e.g.
529 aluminium, copper, iron, lead, tin, zinc) can provide alterations of
530 the absorption profile, reasonably due to metal chelation (Unpub-
531 lished results); these metals, with particular concern to iron from
532 preparation tools, might have been available during the procedure
533 of extraction and preparation of the dye. Even in the case of orchil
534 we have scarce references to compare with: Clementi et al. [21]
535 published spectra of orchil in acetonitrile, ethanol and aqueous
536 solutions; in the last case they reported a marked red shift of the
537 maxima in alkaline solution, which is hardly comparable to those
538 found in solid-state spectra considered here.

539 FORS analysis appears to be the most reliable, among the totally
540 non-invasive methods considered in this work, for the identifica-
541 tion and discrimination of folium and orchil.

542 **Spectrofluorimetry analysis**

543 The fluorescence spectra were registered using a 365 nm LED
544 source. Emission spectra are shown in Fig. 5; the spectrum of the
545 underlying parchment is also reported for comparison. Again, we

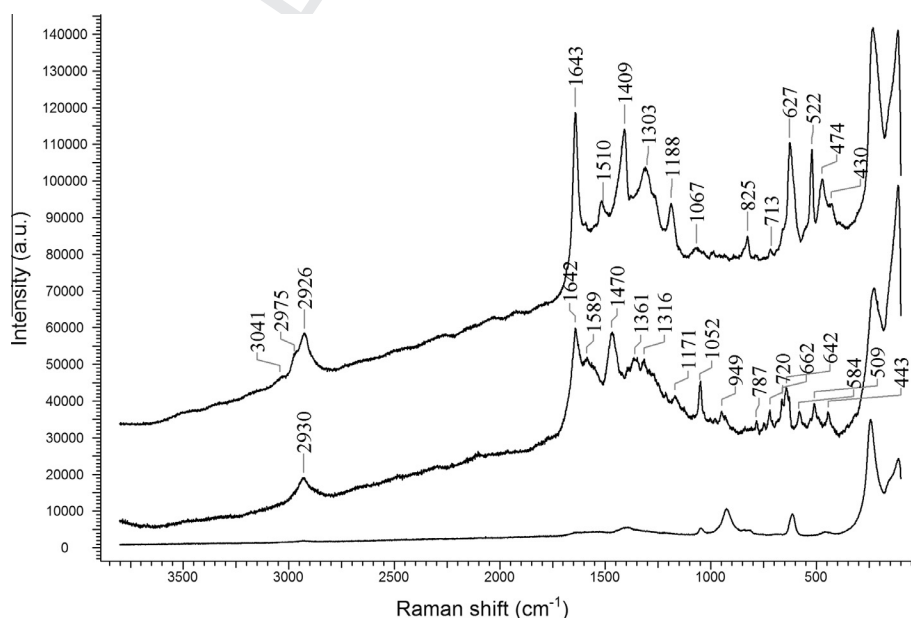


Fig. 3. SER spectra of folium (middle line) and orchil (top line); the spectrum of Ag colloidal paste is also reported (bottom line).

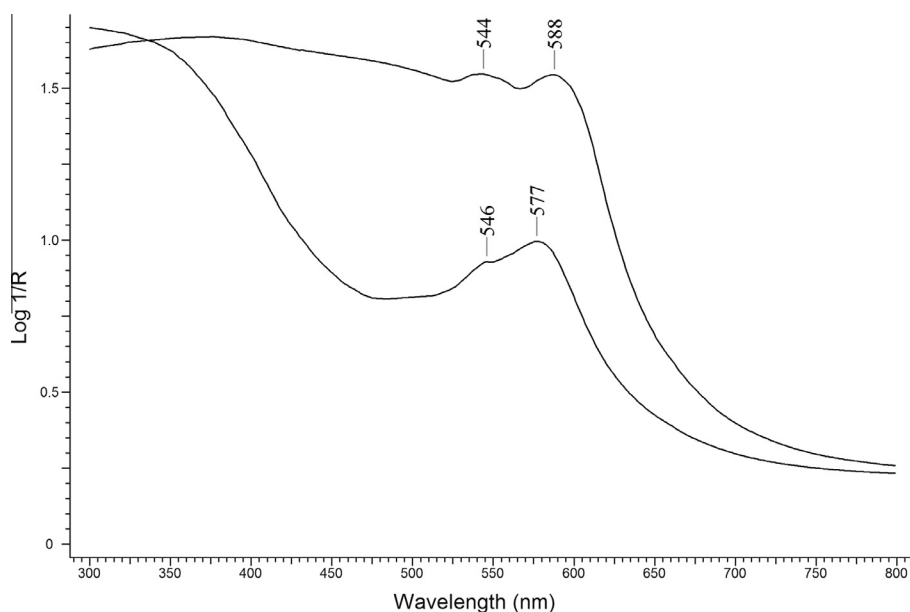


Fig. 4. FORS spectra of folium (bottom line) and orchil (top line).

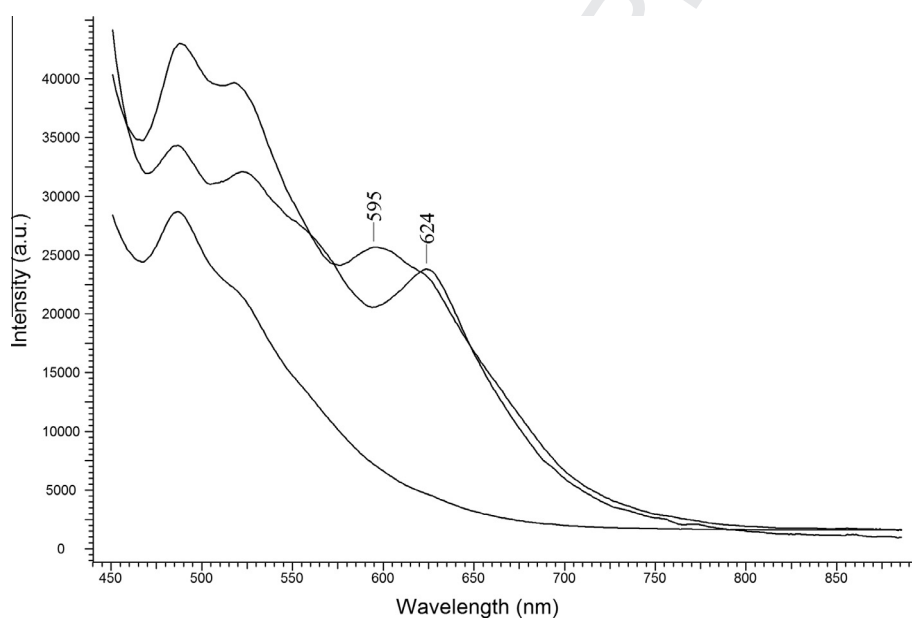


Fig. 5. Fluorescence spectra of folium (medium line) and orchil (top line); the spectrum of parchment is also reported (bottom line).

546 found no differences in the spectra from painted and dyed parch- 561
547 ment samples [26]. The spectrum of folium is dominated by a band 562
548 at 595 nm with a shoulder at ca. 625 nm. In this case also, there are 563
549 no previous data to compare with. As for orchil, the spectrum 564
550 obtained has a neat peak at ca. 625 nm which well compares with 565
551 spectra reported in the literature, for example in the work by Rosi 566
552 et al. [24] and references therein. According to the spectral features 567
553 exhibited by the two dyes, the spectrofluorimetric analysis could 568
554 be selective enough to enable the distinction between folium and 569
555 orchil, at least with the instrumental setup used in this work. 3d 570
556 techniques, analysis in synchro mode or determination of half-life 571
557 times could possibly represent more reliable alternatives [45]. 572

558 MALDI-ToF-MS analysis

559 The application of MALDI-ToF-MS analysis enabled the 573
560 development of another interesting procedure for a micro- 574
575 576
577

561 invasive, micro-destructive procedure for the identification of 562
563 folium and orchil. The amount of sample requested was in fact less 564
565 than 1 mm² of parchment, which was subjected to hydrolysis with 566
567 formic acid as described in the Experimental section. The results of 568
569 MS analysis are shown in Fig. 6. Following a sort of *untargeted* 570
571 approach, once having obtained the mass spectra from the dyes 572
573 some peaks were identified as markers, setting aside the identifica- 574
575 tion of the exact chemical nature of the compounds involved to 576
577 further future research. It appears that two peaks, one for folium 578
579 and one for orchil, can be considered as markers. The mass spec- 580
581 trum of folium (Fig. 6, bottom) is dominated by a peak at 266 *m/z*. 582
583 The mass spectrum of orchil (Fig. 6, top) has its main feature 584
585 in a peak occurring at 672 *m/z*. The main coloured chemical species 586
587 known to be present in orchil according to the literature, i.e. 588
589 amino- and hydroxy-orceins, are barely detectable in the spec- 590
591 trum. The reason of this phenomenon is uncertain; it can be 592
593 hypothesised that, considering the spectral features of these 594
595 596
597

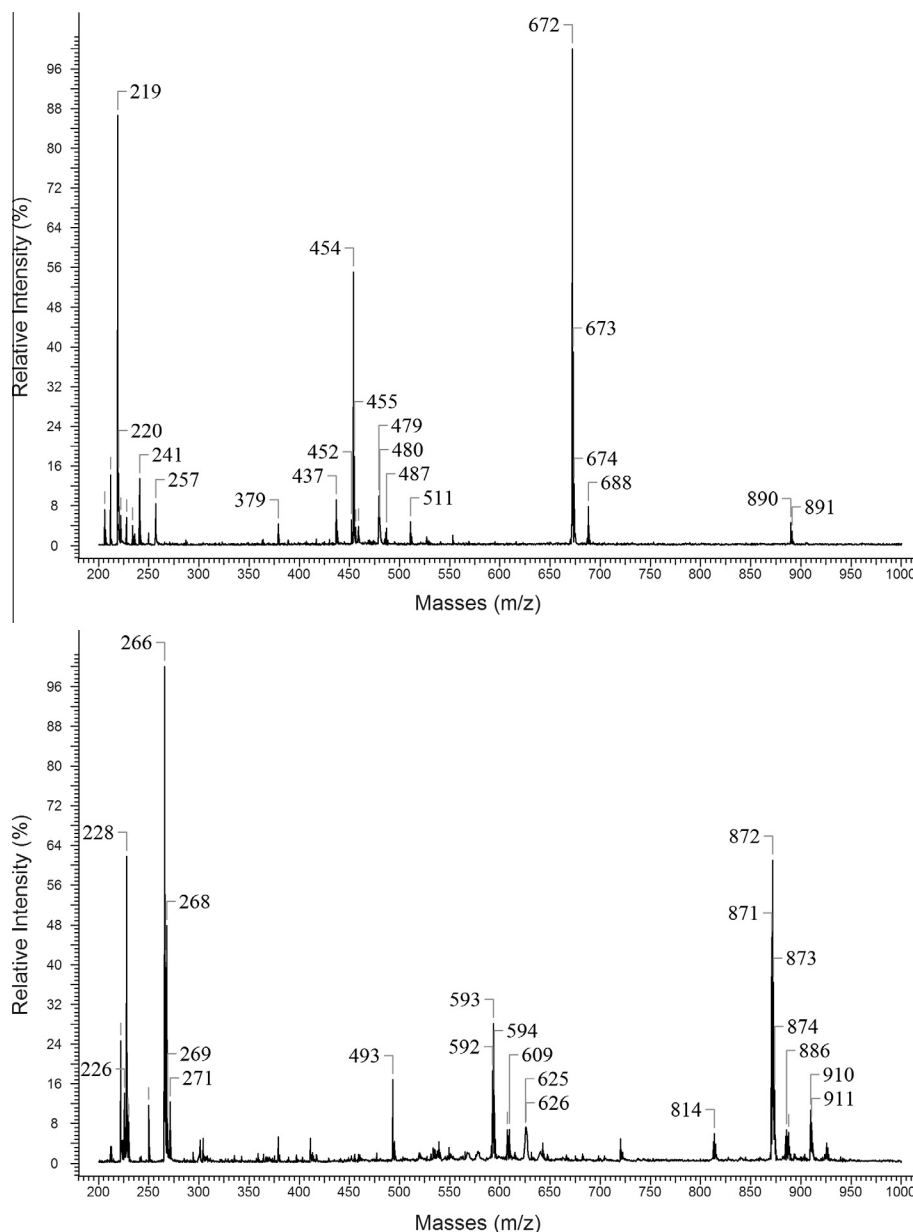


Fig. 6. MALDI-ToF-MS spectra of folium (bottom line) and orchil (top line).

Table 1
ICP-MS analysis of raw matters and dyes.

Sample	Bromine (mg/kg)	Iodine (mg/kg)
<i>Rocella tinctoria</i>	196.0	0.902
<i>Lasallia pustulata</i>	10.2	0.635
<i>Ocrolechia tartarea</i> , sample A	113.5	2.18
<i>Ocrolechia tartarea</i> , sample B	48.9	1.42
Orchil from <i>Rocella tinctoria</i>	159.7	n.d. ^a
<i>Chrozophora tinctoria</i> (L.) A. Juss. pericarp	19.4	0.127
<i>Chrozophora tinctoria</i> (L.) A. Juss. seeds	8.14	0.045
Folium	104.2	n.d. ^a

^a Not detected (below detection limit).

It must be considered that, in the present work, MALDI-ToF-MS analysis was focused on orchil obtained from *R. tinctoria*. However, the spectral profile of purple dyes obtained from different species (e.g. *Lasallia pustulata*, *O. tartarea*, etc.) could be different. This aspect will be examined in depth in a future work.

XRF analysis

Being an elemental technique, XRF spectrometry was used in order to check whether heavy elements were present in the composition of the dyes. Surprisingly, it was found that both folium and orchil contained bromine at a detectable level. Scraps of *R. tinctoria* from Canary Islands, of other coastal lichen samples and a sample of orchil were analysed according to the conditions described in the Experimental section. A semi-quantitative determination of bromine indicated an approximate level of ca. 100 mg/kg in lichen samples. Moreover, this element is entirely or partially extracted from the lichen during the preparation of

molecules (see apparent absorbance spectrum in Fig. 4), they strongly absorbed laser radiation at 337 nm and resonance effect could led to degradation or internal rearrangement of the molecules.

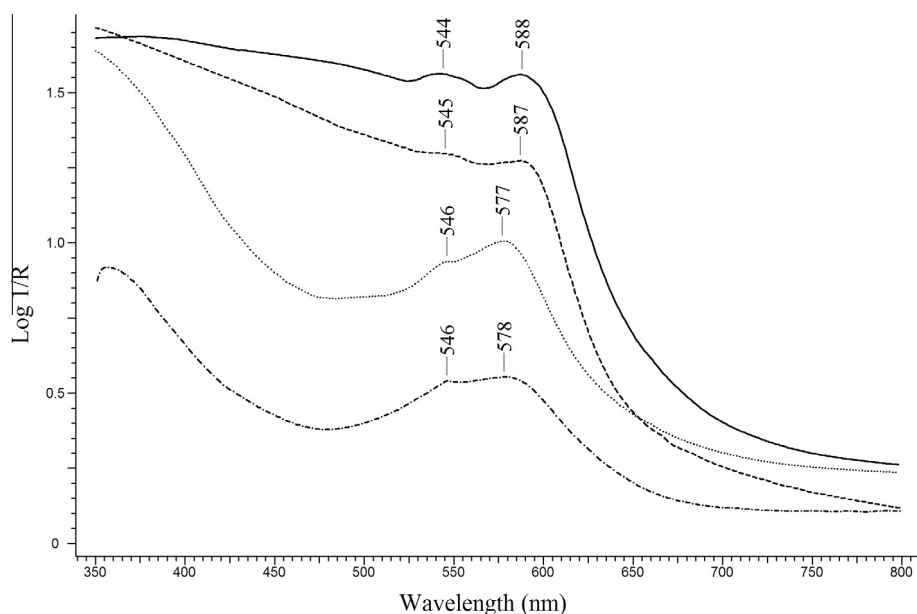


Fig. 7. FORS spectra of orchil (solid line), ms. CIV (dashed line), folium (dotted line) and ms. J.II.1 (dotted-dashed line).

the dye, therefore resulting in the final composition of orchil. Bromine in lichens may be due to their exposure to marine aerosol, since many of the cited lichen species grow on coastal lands. The enrichment of some elements in lichens compared to natural crustal composition has been well demonstrated [46] and bromine, together with chlorine and magnesium, is representative of the contribution of sea-spray. In a study on lichens from Azores and Madeira Archipelagos [47], enrichment factors between 10 and 100 were found for bromine. Bromine could come from low molecular weight organobromine compounds which are known to be produced by living organisms [48].

A similar response was obtained by analysing the fruits of *C. tinctoria* (L.) A. Juss. and the folium powder, even if the level of bromine was found to be lower than in lichens and close to the detection limit of the XRF instrument. The presence of bromine in this plant can also be connected to sea-spray exposure, as it mainly grows on coastal lands of the Mediterranean basin (e.g. Sardinia, Southern France, Turkey, etc.).

ICP-MS analysis

To improve the information obtained by XRF identification of bromine, a more accurate quantitative result was obtained by means of ICP-MS analysis. Samples of *R. tinctoria* from Canary Islands, *L. pustulata* from England and *O. tartarea* from Dartmoor (Southern England) were considered, along with a sample of raw orchil powder obtained from *R. tinctoria*. To evaluate the indication of sea-spray as the origin of bromine, a comparison was carried out among two samples of *O. tartarea*, collected respectively near the coast (sample A) and several kilometres far from the seaside (sample B) with the setup used in this work. As regards *C. tinctoria* (L.) A. Juss., analysis was carried out on the external pericarp (the part richest in purple dye), on the internal seeds and on the raw folium powder.

The results are shown in Table 1: it is apparent that lichens living on coastal lands (*R. tinctoria* and *O. tartarea*) show higher levels of bromine and of iodine, accordingly, than lichens living on internal lands (*L. pustulata*). This fact is confirmed by the analysis of the two samples of *O. tartarea*: the sample coming from the coast (sample A) has a level of bromine which is more than twice that

of sample B, and it has a higher amount of iodine too. In the case of *C. tinctoria* (L.) A. Juss., it is interesting to note that the concentration of bromine is higher in the pericarp than in the internal seeds, according to the hypothesis of the contribution from sea-spray.

From the diagnostic point of view, there is a significant consequence in the results of elemental analysis of orchil and folium: the identification of bromine in the analysis of purple artworks cannot be considered as a definite clue for the presence of Tyrian purple. Some studies on ancient manuscripts involving XRF analysis [26,49,50] led to the hypothesis that the precious dye obtained from shellfish had been used thanks to the identification of bromine, but the present study actually demonstrates that Tyrian purple, orchil and folium may all contribute bromine to the sample.

Analysis of purple and violet painted areas on illuminated manuscripts

To verify the possibility of identifying and distinguish folium and orchil on painted artworks, non-invasive analyses were performed on purple and violet painted areas of several illuminated manuscripts. As an example, Fig. 7 reports the FORS spectra obtained from two manuscripts held in Italian libraries. Ms. CIV or *Libri S. Augustini de Trinitate* is a 9th century codex held in the Archivio Capitolare at Vercelli (Piedmont), while ms. J.II.1, also known as *Beatus of Liébana-Turin Codex*, is a 12th century codex held in the Biblioteca Nazionale Universitaria in Torino (Piedmont). According to the spectral features, in the first case there is a very good match against orchil; the second manuscript, instead, appears to be decorated with folium.

Conclusions

The application of different techniques to the identification of folium and orchil allowed us to select the most suitable procedures of analysis. Besides in this work, the very first FT-IR, FT-Raman and SER spectra of folium have been obtained, FORS has shown to be the best technique for a totally non-invasive approach. It is also evident that micro-invasive techniques such as SERS and MALDI-ToF-MS allowed us to obtain more selective diagnostic information for the identification and discrimination of these ancient dyes,

which can be considered largely unexplored at present. A wider application of SERS and MALDI-ToF-MS is strongly recommended, since they can provide unique information at the expense of a very limited amount of sample.

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