

Università degli Studi del Piemonte Orientale “Amedeo Avogadro”

Dipartimento di Scienze del Farmaco

Dottorato di Ricerca in Biotecnologie Farmaceutiche ed
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EXPRESSION AND DISTINCT FUNCTIONAL ROLES OF AKT ISOFORMS IN MALIGNANT PLEURAL MESOTHELIOMA CELLS

Ester Borroni

Supervised by Prof. Laura Moro

PhD program co-ordinator Prof Menico Rizzi

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Chapter 1

Introduction

1 - MALIGNANT MESOTHELIOMA

1.1. Aetiology of Malignant Mesothelioma

Malignant Mesothelioma (MM) is a highly aggressive cancer arising from the mesothelial lining of the pleura, peritoneum and pericardium. Among these, Malignant Pleural Mesothelioma (MPM), that represents approximately 90% of cases, is the most common primary tumor of the pleura. MPM has a very poor prognosis; it typically spreads and invades locally, even though distant metastases to the contralateral lung, peritoneum, bone or liver can occur (1).

The median survival is less than 12 months, depending on the stage of the disease at the time of diagnosis (2). MPM is associated with exposure to asbestos. Asbestos is the generic term for a group of naturally occurring, fibrous minerals with high tensile strength and resistance to heat, chemicals, and electricity. Six minerals, defined as "asbestos", are divided in two classes: the serpentine and the amphibole. Chrysotile (white asbestos), is the only one member of the serpentine class, is made up of curled fibers and accounts for approximately 95% of all asbestos used worldwide. The amphiboles' group can be distinguished for its friable fibers and includes amosite (brown asbestos) and crocidolite (blue asbestos), extensively used in industries (3).

Between the 1940s and 1979 in the United States and Europe, asbestos was widely used in the shipbuilding and construction industries thanks to its

fire-resistant properties (4). The incidence of asbestos-related diseases has been rising worldwide since 1970, due to the exposure during the past and it is predicted to increase further in coming decades, especially in developing countries where asbestos has not yet been banned. The mean latency period between the time of initial exposure and diagnosis is about 30 years and ranges from 20 to 50 years; moreover, the disease latency has an inverse relationship with duration of exposure (5).

The first hypothesis about a link between cancer and asbestos, was proposed by Wagner et al. (6) in 1870, who described crocidolite asbestos exposure in 33 cases of mesothelioma in South Africa's North Western Cape, while further confirmations came during the following years when McDonald found a proportional mortality ratio for exposed subjects.

Currently rates of MPM are rising and estimates indicate that the incidence of MPM will peak within the next 10-15 years. Although the use of asbestos has been banned in many countries around the world, production of and potentially hazardous exposure to asbestos is still present with locally high incidences. Today a new man-made material, carbon nanotubes (CNTs), has arisen as a concern; CNTs may display 'asbestos-like' pathogenicity with mesothelioma induction potential (7).

Exposure mapping within countries reveals high regional variability in incidence and mortality: for instance, in Italy, significant municipal clusters of disease have been identified close to asbestos industries, shipyards, oil refineries and petrochemical industries (8); similarly in the UK, the highest mesothelioma mortality rates were recorded in areas with a history of ship building, such as Barrow-in-Furness, Plymouth, Portsmouth, Tyneside and Southampton (9). The correlation between amphibole, asbestos exposure and MPM development is generally accepted and crocidolite is considered the most oncogenic type of asbestos. Even if the relative oncogenicity of the specific asbestos fibers type is controversial, the evidence has been

considered sufficient by the World Health Organization (WHO) to conclude that all types of asbestos cause cancer in humans (10).

Asbestos fibers can be inhaled deeply and trapped in the lower lung where mechanically cause pleural irritation and induce an inflammatory response as consequence of repeated scratching of mesothelial surface. Moreover, phagocytosed fibers can initiate a cascade of events that includes activation of c-Myc and c-Jun oncogenes, binding with growth factor receptors (EGFRs) (11,12), or alternatively, mechanically interference with the mitotic process of the cell cycle by disrupting the mitotic spindle, with consequent chromosomal abnormalities and aneuploidy (13). Finally, another proposal is that asbestos can also induce reactive oxygen species (ROS) or reactive nitrogen species (RNS) (resulting in DNA damage), cytokines, growth factors -such as transforming growth factor- β (TGF- β)- as well as transcription factors -such as nuclear factor kappa B (NF- κ B) and activator protein-1 (AP-1) (14, 15).

1.2. MPM classification

According to the WHO classification, based on the histological analysis of tissue obtained by biopsies, MPM is divided in three different classes: epithelioid, sarcomatoid and biphasic.

Epithelioid

Around 60% of all mesothelioma cases comprises mesothelial cells with epithelial-like shape arranged in tubulo-papillary or trabecular formations and represents the most common histotype characterized by a better prognosis (16). The most frequent patterns encountered are tubulopapillary, adenomatoid (microglandular) and sheet-like, while less common patterns include small cells, clear cells and deciduoid. In particular, the tubule papillary form shows varying combinations of tubules with connective tissue

cores and clefts. Most epithelioid mesotheliomas are singularly bland in appearance and mitoses are scarce; usually a pattern predominates but often-different ones are commonly found in the same tumor, as much as sheets and nets of cells are frequently seen in association with other patterns (17). The fibrous stroma of epithelioid mesotheliomas can show varying degrees of cellularity from hyalinised acellular to highly cellular, merging with sarcomatoid; furthermore, these tumors may be difficult to distinguish from biphasic mesotheliomas.

Sarcomatoid

Sarcomatoid histotype is associated with less differentiated and more aggressive tumors in which more than 90% of the cells assume a spindle morphology and are arranged in fascicles or sheets, showing many mitotic figures, necrosis, and cytological atypia. Cells frequently tend to be irregular rather than uniform and overlap one other: their pattern often looks like fibrosarcoma, but marked anaplasia and uncommon multinucleate tumor cells make it more similar to malignant fibrous histiocytoma (18).

Biphasic

The biphasic histotype contains both epithelioid and sarcomatoid patterns and each component should represent at least 10% of the tumor to warrant the term biphasic. Cells of the biphasic variety form groups rather than displaying a uniform mixture (19).

1.3. Diagnosis, Staging and Prognosis

Malignant mesotheliomas are not ordinarily graded.

Patients with MPM commonly present with dyspnoea, chest wall pain and pleural effusion and diagnosis is often made at an advanced stage of the disease, so that in untreated patients median survival is less than one year.

Difficulty in diagnosing the disease, especially in its early stages, prevented the development of a generally accepted stage-related approach. In addition, MPM is a heterogeneous disease with a variable clinical course: the majority of patients (80%) are diagnosed in stage III/IV and these patients are not candidates for surgical cure (20). Thus, systemic therapy is the only treatment option for the majority of this kind of patients, but poor performance status and the low chemo and radio-sensitivity of the tumor result in a poor prognosis. For these reasons, advances in the understanding of the molecular biology of MPM is needed to establish diagnostic, therapeutic and prevention methods and allow the identification of promising new candidates for targeted treatments.

Knowledge of the biological basis of MPM progression and response may facilitate a personalized treatment approach involving early identification of poor prognostic indicators that will reduce the heterogeneity of the clinical response (1, 18).

One contributing factor for the diagnosis of malignant mesothelioma is the possibility to differentiate it from adenocarcinoma, a cancer originating in glandular tissue. The most important differential is metastatic or locally invasive (from lung or chest wall) tumor that covers the pleural surface, even if various localized tumors also exist in the pleura and some mimic mesothelioma microscopically. For this reason, knowledge of the distribution of tumor, whether obtained from radiographic studies, the operator's description of the findings at thoracotomy or thoracoscopy, or from a resected specimen, is crucial to make a proper diagnosis. However, beyond the distinction between epithelioid and sarcomatoid forms, these histopathologic features do not always correlate well with prognosis (17).

Staging

Several staging systems for mesothelioma have been used over the years, almost exclusively dealing with primary pleural mesothelioma. Peritoneal mesothelioma does not have its own staging system. The oldest staging system used for pleural mesothelioma is the Butchart system; it is still commonly used in some parts of the world (13). This method is based on a simple description of the extent of the disease without considering the histologic subtype: pleural contained (Stage I), chest wall or mediastinal invasion (Stage II), peritoneal or diaphragmatic penetration (Stage III), or distant metastases (Stage IV). Meanwhile, the Brigham system tries to define surgical resectability and lymph node involvement. Stage I disease is resectable without nodal spread while stage II is resectable with lymph node involvement. Stage III involves the chest wall, heart, diaphragm, or abdominal cavity, with or without lymph node involvement: stage III tumors are considered unresectable. Stage IV disease is distant metastases. This system is not utilized at present. The most practical and most commonly used system is the tumor-node-metastasis system developed by the International Mesothelioma Interest Group. This system is the currently accepted standard adopted by the American Joint Committee on Cancer (21, 22). Most patients present with advanced disease and are considered unresectable.

Histopathology and Survival

The prognosis for MPM is significantly affected by histopathologic variant. Most studies have found a clear survival benefit for patients with epithelioid subtype. A recently published study on 312 patients undergoing surgical therapy for MPM evaluated the effect of histology on survival (23). Patients with epithelioid subtype had the longest median survival of 15.3 months, and 1-year and 2-year survival of 63% and 32% respectively. Patients with

sarcomatoid subtype had a median survival of 5 months, and 1-year and 2-year survival of 4% and 0%, respectively. The biphasic subtype had intermediate survival rates. Multiple other studies have demonstrated extremely poor survival for patients with sarcomatoid-type MPM regardless of medical or surgical therapy, and most centres do not offer surgery to these patients (24).

1.4. Therapy

Mesothelioma is difficult to treat because many people are not diagnosed until their disease is advanced and the decisions about treatments are based on different tasks: where the tumor arose and its stage, the grade of malignancy, its location, whether it is resectable or not, symptoms, general health conditions of the patient and age (25). Treatment options are limited and the results with conventional therapies have been rather disappointing to this date. Moreover, chemotherapy is the only evidence-based treatment for mesothelioma patients in good clinical condition, with an increase in median survival of only 2 months: so finding out new different approaches to afford this malignancy became necessary.

Actually the therapy for mesothelioma is represented by surgery, radiotherapy, chemotherapy and the so-called “trimodality therapy” that consists in combination of chemotherapeutic drugs following surgery (26).

1.4.1. Surgery and Radiation

Surgery

Surgery for mesothelioma could be done essentially for two reasons: to try to cure cancer or to relieve pain and other symptoms carried on by the tumor. The curative surgery is an option for patients who are living with good health conditions because the cancer has not spread a lot and it could be removed completely also because it is thought to be localized. However

sometimes surgeons are not able to remove all of the cancer, some cells remain and can grow normally, divide allowing the cancer to come back after surgery. Otherwise, palliative surgery may be an option if the tumor has already spread beyond where it started and complete removal is not possible, or if the patient is too ill for a more extensive operation (27). The type of surgery done depends mainly on the location of the tumor, the stage of the cancer, the subtype of mesothelioma and other factors, such as patient overall health, also because chemotherapy and radiation represent additional therapies that are used in combination with surgery.

- Pleural mesothelioma surgery

There are two types of surgical treatments that may be offered to patients with pleural mesothelioma: pleurectomy/decortication (P/D) and extrapleural pneumonectomy (EEP). People with early stage or localized (stage I) pleural mesothelioma are undergone to P/D with removal of both the parietal pleura and visceral pleura together with the tumor, while the lung is not removed. In more advanced stages, a P/D can also be done to help control fluid build-up in the chest, improving breathing and relieve symptoms, such as chest pain caused by the tumor. Patients in stage I or II, and some with stage III pleural mesothelioma are treated with extrapleural pneumonectomy (EPP), only if the tumor is considered operable, is localized and has not spread into the diaphragm – this typically occurs in patients with resectable mesothelioma of the epithelioid type whose cancer has not reached the lymph nodes(28).

- Peritoneal mesothelioma surgery

Surgery is not often possible for people with peritoneal mesothelioma, but in people who are eligible candidates for surgery this treatment is often done to help relieve symptoms or to remove the tumor from the abdomen wall and organs; unfortunately as with pleural mesothelioma, these tumors often have spread too far to allow a complete removal (29). Essentially,

people with peritoneal mesothelioma may have a peritonectomy, in which the membrane that lines the abdomen (peritoneum) is removed, sometimes involving also other structures; or an omentectomy, a surgery that removes the layer of fatty tissue that covers the contents of the abdomen, when mesothelioma spread beyond the peritoneum (30).

Radiation

The pattern of spread of malignant pleural mesothelioma (MPM) poses unique challenges to a radiation oncologist. At first, patients are treated with conventional radiation techniques, when radiotherapy administered as adjuvant therapy following extrapleural pneumonectomy (EPP) or pleurectomy/decortication (P/D) (31). Anyway, with conventional radiotherapy there may be radiation underdosing near regions that are blocked: this has the potential to lead to increased risk of local failure in approximately 10-15% of patients. More recently, complex intensity-modulated radiation therapy (IMRT) techniques for the treatment of MPM have been explored, with early outcomes suggesting acceptable safety in appropriately selected patients. Multiple subsequent studies incorporated this technique into a multimodality approach combining chemotherapy, EPP and hemithoracic radiation (32).

1.4.2. Chemotherapy and Multimodal Therapy

Treatment modalities for MPM patients depend on age, performance status and disease stage at onset. It is now generally accepted that the future of cancer therapy lies in the combination of therapies with different mechanisms of action; actually the therapy for MPM is represented by surgery and radiotherapy, chemotherapy and the so-called “trimodality therapy” that consists in combination of chemotherapeutic drugs following surgery.

Chemotherapy

A phase 3 trial of combined therapy using pemetrexed and cisplatin has shown major efficacy compared with cisplatin monotherapy and consequently, current approved first line therapy now include cisplatin and antifolate agent, such as pemetrexed or raltitrexed (33).

Several other cytotoxic agents with definite activity in mesothelioma have recently been evaluated, including gemcitabine, vinorelbine and the antifolates pemetrexed and raltitrexed. Vinorelbine has been shown to have activity in a small phase II study; similar results have been achieved with vinflunine, a new semi-synthetic fluorinated vinca alkaloid. In contrast, taxanes, such as paclitaxel and docetaxel, showed very low or no activity. Platinum analogues, doxorubicin and some antimetabolites (methotrexate, raltitrexed, pemetrexed) have shown modest efficacy as single-agent. The combination of antifolates and cisplatin was proven to prolong survival over cisplatin alone in Phase III studies, while in other phase II studies, the combination of pemetrexed and gemcitabine was also effective as a first-line chemotherapy for patients with MPM (34).

Recently a study on 409 patients with MPM was conducted assigning them randomly to one of three treatments for 12 weeks: active symptom control (ASC) alone, ASC plus MVP (mitomycin vinblastine, and cisplatin), or ASC plus vinorelbine. The median survival was 7.6 months in the ASC alone group and 8.5 months in the ASC plus chemotherapy group. Considering that ASC group included treatments with steroids, analgesic drugs, bronchodilators or palliative radiotherapy, the addition of chemotherapy to ASC did not offered significant benefits in terms of the overall survival or quality of life (35).

Chemotherapies consisting of doxorubicin, doxorubicin plus cyclophosphamide, oxaliplatin-raltitrexed or ZD 0473 (a platinum analogue) appeared ineffective. Some interesting response rates were noted with

pemetrexed alone, or the combination of carboplatin and pemetrexed and of cisplatin, irinotecan and mitomycin C, used as second line therapy (36). Weekly vinorelbine administration demonstrated useful clinical activity in the second-line treatment of MPM, resulting in a median overall survival of 9.6 months. Gemcitabine and vinorelbine in pemetrexed-pretreated patients with MPM resulted in a 10.9-month median survival time. Moreover, retreatment of pemetrexed-pretreated patients appeared effective, with 1-year survival rates of 71.0% for retreated patients versus 18.8% without retreatment.

In patients receiving second-line chemotherapy after the phase 3 trial of pemetrexed plus cisplatin versus cisplatin alone, a positive impact on survival was evident. MPM patients receiving second-line therapy with pemetrexed exhibited significant anti-tumor activity, improved disease control rates, and longer progression-free survival than patients receiving the best supportive care alone, although survival was not improved (37, 38).

- Immunotherapy

Immunotherapy is one of the more recent approaches to cancer therapy: it represents a new class of agents in the treatment of mesothelioma and it is based on the generally-accepted hypothesis that the immune system is the best tool humans have for fighting disease. This therapy is sometimes used by itself to treat cancer, but it is most often used in combination with traditional treatments like radiation, chemotherapy, and surgery in order to enhance their effects. One of the possible benefits of immunotherapy is that it has the potential not to be as toxic as radiation or chemotherapy, and less invasive than surgery.

In the case of cancer, the immune system alone often fails to effectively fight the tumor; normally the immune system does not recognize tumor cells because they are derived from the body's own cells, so the body “thinks” of

the tumor as “self” and creates a phenomenon known as “tumor tolerance”. Otherwise the immune system may also recognize cancer cells, but the response is not strong enough to destroy the cancer (because the response is insufficient in amount, it does not occur in the right place in the body, or it does not send the right “immune cells”). Moreover, the tumor has the ability to defend itself and cancer cells may secrete substances that keep the immune system in check. Thus, in the case of cancer, the immune system needs something to afford it more efficiently (39).

The purpose of cancer immunotherapy is to promote (activating or intensifying) the antitumor effects of the immune system of the patient; several approaches for immunotherapy have been developed over the years and some are in various stages of pre-clinical research. Immunotherapy can be divided in two main categories, passive and active immunotherapy. Passive immunotherapy uses in-vitro produced effectors that are able to influence tumor cell growth: the most common form is based on the monoclonal antibody therapy, which consists in humanized monoclonal antibodies targeting cells directly or indirectly, which efficacy may be often enhanced by linking a toxin (e.g., radionucleotides or anticancer drugs) (40). Active immunotherapy approaches aim at inducing or boosting immune effector cells *in vivo* against tumor cells, through the administration of immune mediators capable of activating the immune system (i.e. cytokines capable recruiting specific immune cells for enhance antitumor immunity, as IL-2, IL-12, IL-15, TNF- α , GM-CSF). These cytokines can be used as single agent or in combination with other immunotherapeutic strategies (41).

- Molecular-Targeted Therapy

Epidermal growth factor receptor (EGFR), platelet-derived growth factor receptor (PDGFR), vascular endothelial growth factor receptor (VEGFR), SRC, hepatocyte growth factor (HGF), transforming growth factor (TGF),

insulin like growth factor I receptor (IGF-1R), mammalian target of rapamycin (mTOR), histone deacetylases (HDAC), and the proteasome have been investigated and can be targeted in MPM treatment (42, 43).

Several clinical studies have been conducted. Phase 2 clinical trials based on EGFR tyrosine kinase inhibitor (erlotinib and gefitinib) therapy have shown limited or no activity in MPM; probably, the lack of EGFR mutations that confer sensitivity to TK-inhibitors in NSCLC could also explain the resistance to EGFR tyrosine kinase inhibitors in this cancer.

The phase 2 studies with the anti-EGFR monoclonal antibody, cetuximab, with platinum/pemetrexed are ongoing. In three phase 2 trials involving imatinib as a PDGFR inhibitor, no objective response was obtained; moreover, a combination of imatinib with gemcitabine, cisplatin/gemcitabine, or cisplatin/pemetrexed has also been investigated without good outcomes (44,45). Bevacizumab, a recombinant human anti-VEGF monoclonal antibody that blocks the binding of VEGF to its receptors, is under evaluation in a double-blind, randomized phase II trial in combination with cisplatin and gemcitabine.

Other novel agents under investigation include sorafenib, an inhibitor of VEGFR-2, PDGFR-b, and B-Raf tyrosine kinase; in a phase I trial, vorinostat, a histone deacetylase inhibitor, has produced objective responses in 20% of MPM patients, and a phase III double-blind, placebo-controlled trial is under way (46).

Multimodal Management of MPM

Failure of single-modality treatments to increase survival in MPM patients has led to a variety of multimodality approaches.

The typical curative surgical treatment is extrapleural pneumonectomy (EPP) that involves a total en-bloc resection of the lung, pleura, pericardium, and diaphragm, while pleurectomy/decortication (P/D) involves

resection of the parietal and visceral pleura, and pericardium. This cytoreductive surgery has been added to systemic and/or intrapleural chemotherapy and to external-beam or intraoperative RT, with the main aim of improving local control. Early studies of neoadjuvant chemotherapy followed by EPP and hemithoracic RT have been published, and confirmatory trials on patients are ongoing.

Several studies applying gemcitabine and cisplatin as an induction chemotherapy in combination with EPP and radiation in a combined-modality approach for resectable MPM have been also reported.

Patients completing trimodality therapy displayed a median survival of 29.1 months and a 2-year survival rate of 61.2%.

Patients who underwent trimodality therapy involving EPP and adjuvant chemoradiotherapy displayed a median overall survival of 13– 23.9 months. It was concluded that selected patients with MPM may benefit from EPP, particularly when combined with induction or adjuvant chemotherapy and adjuvant radiotherapy. Due to its toxicity, is difficult to administer EPP as adjuvant chemoradiotherapy; thus, a trimodality therapy consisting of induction chemotherapy and EPP followed by radiotherapy was preferred. Finally, surgery may remove macroscopic diseases and multimodality therapy with radiation and chemotherapy should be included to eradicate residual microscopic disease (47, 48).

2 - AKT

2.1. AKT Family: Isoforms and Roles

AKT/PKB (Protein Kinase B) regulates many cellular functions such as cell growth and proliferation, cell survival, apoptosis, energy metabolism, migration and resistance to anticancer therapeutics (49). Despite the

growing amount of researches demonstrating the existence of isoform-specific regulation, many papers still draw generalized conclusions about AKT functions without considering the unique function of each AKT isoform. AKT family consists of three isoforms, AKT1, AKT2 and AKT3 (also known as PKB α , PKB β and PKB γ , respectively) (50). Although these three isoforms are encoded by separate genes, they share a common NH₂-terminal pleckstrin homology (PH) domain (of 110 amino acids), a catalytic (kinase) domain in the middle (of about 260 amino acids) separated by a hinge region of 39 amino acids (“Linker”) and a C-terminal regulatory domain (with an identity in amino sequence of about 80%), which contains an hydrophobic motif, characteristic of the cAMP-dependent protein kinase A/protein kinase G/protein C (AGC) superfamily of protein kinases (51).

They are differentially expressed in various tissues: AKT1 is most abundant in brain, heart, and lung; AKT2 is predominantly expressed in skeletal muscle and embryonic brown fat, while AKT3 is mainly expressed in kidney, brain and embryonic heart, although it is also present in adipose tissue, mammal glands and lungs.

AKT isoform-specific knockout mice suggest that different AKT kinases are not completely overlapping and that isoform-specific signaling contributes to the diversity of their biological function.

- AKT1 knockout mice are smaller than littermate controls and show increased rates of apoptosis in some tissues, reflecting its role in cell survival (52).
- AKT2 null mice develop type 2 diabetes and impaired glucose utilization, suggesting its importance in the insulin receptor-signaling pathway (53).
- The precise role of AKT3 is less understood: AKT3 is a key modulator of several tumors like glioma, ovarian cancer and melanoma in which an increase of active this isoform promotes

tumorigenesis by decreasing apoptosis; moreover, mice lacking AKT3 show impaired brain development (54).

2.2. PI3K/AKT/mTOR Pathway

AKT activity is modulated downstream of phosphatidylinositol 3 (PI3) kinase via a multistep process that starts in response to various growth factors and cytokines such as platelet-derived growth factor receptor (PDGF-R), epidermal growth factor (EGF) and insulin-like growth factor 1 (IGF-1). In the absence of growth factor stimulation in quiescent cells, all three isoforms of the AKT kinases are catalytically inactive (55).

The PI3Ks constitute a family of intracellular lipid kinases that phosphorylate the 3'-hydroxyl group of phosphatidylinositols (PIs) and phosphoinositides. Based on structure and their specificity for substrates, PI3Ks are classified into three groups (classes I, II and III).

Class I PI3Ks are the primary lipid kinases that generate PIP3 from PIP2, and are subdivided into two classes (a, activated by growth factor TK-receptor, and b, activated by G protein-coupled receptors); class II binds clathrin and are while both (class II and III) use PI to generate PI-3-P (56).

After ligand-induced activation of specific receptors, a phosphorylated Y residue on receptor serves as docking site for regulatory p85 sub-unity of PI3K that recruits p110 sub-unity to the complex. PI3K activated localizes to the cytoplasmic face of the plasma membrane, where it converts PIP2 (phosphatidylinositol 4,5-bisphosphate) to PIP3 (phosphatidylinositol 3,4,5-trisphosphate), which promotes membrane localization of molecules such as AKT and 3-phosphoinositide-dependent kinase 1 (PDK1) by binding to their PH domain.

The C-terminal PH domain of PDK1 binds phospholipids, keeping it constitutively localized at the plasma membrane; upon GF stimulation AKT interacts with these phospholipids, causing its translocation to the inner

membrane, where PDK1 is located. The interaction of the AKT PH domain with 3'-phosphoinositides makes conformational changes in AKT with the exposure of its main phosphorylation sites, T308 and S473 (57).

Direct homodimerization of the two PH domains between AKT and PDK1 might phosphorylate Thr-308 in AKT, which stabilizes the activation loop in an active conformation and renders Ser473 phosphorylation by mTORC2 (mammalian target of rapamycin complex 2), resulting in full activation of the kinase. However, other molecules, including integrin-linked kinase (ILK) and mitogen-activated protein kinase-activated protein kinase-2 (MAPK-APK2), mTOR and AKT itself have also been described to phosphorylate Ser473 (58). Full activation of AKT is a multistep process that leads to AKT phosphorylation on two sites, Thr308 (for human AKT1) and Ser473 (for AKT1).

In particular, to optimize the kinase activity of all AKT isoforms, the concomitant phosphorylation of threonine and serine residues is required: curiously, they are these two amino acids are positioned in marginally different locations in the three isoforms.

For instance, the most essential regulatory amino acid residues are threonine 308 and serine 473 in AKT1, whereas the amino acid residues are threonine 309 and serine 474 in AKT2, while in the case of AKT3, the regulatory residues are threonine 305 and serine 472 (59).

AKT can be activated in a PI3K-independent manner: for example, agonists of the PKA pathway, such as cAMP elevating agents, can activate AKT: calcium binds to calmodulin, and the Ca²⁺/calmodulin complex activates the calcium/calmodulin dependent kinase kinase (CaMKK), which directly phosphorylates T-308. Finally, some non-kinase interactors such as Hsp90/Cdc37 (molecular chaperone and co-chaperone), Hsp27, Tcl1 (T-cell leukemia family), Grb10 (growth factor receptor binding protein10), Ft1 and many others can positively regulate AKT catalytic activity (60).

2.3. Downstream effectors of AKT

AKT activation is primarily implicated in cell survival and cell growth through regulation of several substrates. Among these substrates there are:

- regulators of cell survival or death, such as BAD, caspase-9, ASK1 (apoptosis signal-regulating kinase 1), forkhead box O transcription factors (FoxOs), Bim1, FasL, inhibitor of nuclear factor- κ B kinase (IKK-NF κ B), and p53;
- regulators of angiogenesis, such as mTOR and hypoxia-inducible factor-1 (HIF-1);
- regulators of cell metabolism, such as glucose transporter 1 (Glut1), GSK3, and a Ras homologue enriched in brain (Rheb).

AKT also regulates cell growth acting on tuberous sclerosis complexes 1 and 2 (TSC1/2), mTOR, elongation-initiation factor 4E binding protein-1 (4E-BP1) and mTOR pathway; it also modulates cell cycle progression through direct actions on the p21 and p27 (CDK inhibitors) and indirect effects on the levels of cyclin D1 and p53 (61).

2.4. Inactivation of AKT

AKT cascade can be blocked by cellular inhibitors as the phosphatase and tensin homolog (PTEN) and INPP4B, which directly antagonize PI3K functions, dephosphorylating PIP3 and revoking its downstream events. PTEN has been also demonstrated to be one of the most commonly altered genes in human malignancies. In contrast, gain-of-function AKT mutations are uncommon, but frequently occur at residue 17, within the PH domain (E17K-Lys 17 alters the electrostatic interactions of the pocket and forms new hydrogen bonds with a phosphoinositide ligand). This mutation activates AKT leading its localization to the plasma membrane, stimulates downstream signaling, transforms cells and induces leukemia in mice, indicating a direct role of AKT in cancer; moreover, the E17K mutation

does not change AKTs sensitivity to ATP-competitive inhibitors, but it modify change its sensitivity to the allosteric ones (62).

Negative regulation of PI3K/AKT pathways is mainly accomplished by PTEN (phosphatase and tensin homologue deleted). It was originally identified as a tumor-suppressor gene and was found frequently mutated in a wide variety of solid tumors, including endometrial, breast, prostate carcinomas, and glioblastomas. As a dual lipid and protein phosphatase, the primary physiological target of PTEN is considered to be the PI3K/AKT pathway.

PTEN specifically catalyses dephosphorylation of the 3-phosphate of the inositol ring in PIP3, resulting in the biphosphate product PIP2 and inhibition of AKT activity. Inactivating mutations or loss of PTEN expression leads to a permanent increase in the basal level of PI3K/AKT signaling, generally resulting in increased cell proliferation and resistance to apoptosis. Recently, a family of novel protein phosphatases, namely, PHLPP (PH domain and leucine-rich repeat protein phosphatase), have been identified as important regulators of AKT kinases and protein kinase C (PKC). Two isoforms, PHLPP1 and PHLPP2, directly dephosphorylate Ser473 and therefore inactivate AKT. It has been shown that PHLPP differentially terminates AKT signaling through particular regulation of AKT isoforms. PHLPP2 dephosphorylates AKT1 and AKT3, whereas PHLPP1 is specific for AKT2. PHLPP expression is commonly lost in cancer, including colon, breast, ovarian, Wilms tumors, and prostate cancer, while its co-deletion, together with PTEN is strongly associated with metastatic prostate cancer, suggesting the role of PHLPP as a tumor suppressor (63).

2.5. Post-translational Modifications of AKT

A part phosphorylation other post-translational modifications has been described to modify AKT activity/stability.

AKT acetylation

The level of protein acetylation is regulated by enzymes that increase or decrease acetylation. When the balance between these mechanisms is altered, it results in protein hyperacetylation or deacetylation. As phosphorylation, the reversible acetylation of cellular proteins plays an important role in cellular processes; whereas different amino acids (i.e., threonine, serine, and tyrosine) can be phosphorylated, acetylation occurs only on lysine residues (64, 65).

Deacetylases are enzymes that remove acetyl groups: they can be divided in four classes based on their structural and functional similarities, and among these, the major family is represented by sirtuin enzymes, that consist in seven components designated as SIRT1 through SIRT7, exhibiting different tissue distributions and subcellular localizations.

Activity of AKT requires tight control, as evidenced by the multiple means by which the protein is regulated: phosphorylation, ubiquitination and binding to phospholipids.

Recently, Sundaresan et al. reported the regulation of AKT and PDK1 by reversible acetylation. Their work showed that p300 and pCAF acetylate AKT and PDK1, while SIRT1 deacetylates them. The acetylation regulated the ability of AKT to bind PIP3, offering a new sight of AKT regulation through reversible acetylation: AKT was acetylated in murine cells in a manner inversely related to its activation. The kinase was associated with the histone deacetylase SIRT1 and deacetylation enhanced its ability to be activated. In contrast, activation of AKT after growth factor stimulation was inhibited in cells lacking SIRT1 (66, 67); moreover, cells effects on cardiac hypertrophy in mice lacking or overexpressing SIRT1 were associated with altered regulation of AKT. Thus, acetylation of AKT (regulated by SIRT1) appears to be an important aspect of control of different biological activities of the enzyme (68). Another elegant work conducted by Ramakrishnan and

colleagues, focused the attention on SIRT2, the primary cytoplasmic sirtuin, to which it have been attributed tumor suppressor functions and a role in maintaining genome integrity and in programmed necrosis. There is substantial crosstalk between the insulin-PI3K-AKT-metabolic pathways and sirtuins: SIRT1 and SIRT2 can deacetylate and regulate the function of FoxO transcription factors, which are direct AKT targets.

In particular, they showed that the main sirtuin that binds and regulates AKT activation in insulin-responsive cells is SIRT2 rather than SIRT1. While AKT associates with both SIRT1 and SIRT2 in cells with constitutive PI3K activation, it exclusively binds SIRT2 in cells with normal regulation of the PI3K-AKT pathway. The SIRT2-AKT binding is induced by glucose and nutrient deprivation and PI3K inhibition, while insulin treatment induces the dissociation of the complex.

Finally, the study identifies SIRT2 as a new positive AKT regulator that potentiates insulin responsiveness in normal cells, while both SIRT1 and SIRT2 play a role in maintaining AKT activation in cancer cells with constitutive PI3K activation; all these observations lead to considerate that SIRT2 activators could be useful in the treatment of obesity-associated metabolic syndrome and type-2 diabetes, while SIRT1/2 inhibitors may have therapeutic use in cancers with constitutive PI3K-AKT pathway activation (69).

Ubiquitin-dependent AKT degradation

The ubiquitin proteasome system (UPS) regulates many cellular functions such as cell cycle, growth and cell polarity, through degradation of some proteins. The UPS acts involving the sequential activation of three enzymes: ubiquitin activating enzymes (E1), ubiquitin-conjugating enzymes (E2), and ubiquitin ligases (E3), which determine the specificity of substrates; activation of E1, E2, and E3 results then in the conjugation of

ubiquitin to the lysine residues of target proteins, that are tagged with poly-ubiquitin and degraded by the proteasome complex (70, 71, 72).

It is demonstrated that E3 ligases are involved in AKT degradation by UPS: Kim et al. reported that mannitol-induced degradation of AKT can be blocked by IGF-1, while in other studies reported that deprivation of vascular endothelial growth factor (VEGF) or blockade of the VEGF cascade with the tyrosine kinase inhibitor PTK787/ZK222584 resulted in a specific decrease of AKT protein level and the subsequent cellular stimulation with VEGF rescued AKT stability in endothelial cells (73, 74). In addition, the mTOR inhibitor also neutralized the VEGF-protective effect in proteolysis-dependent reduction of AKT protein, suggesting that this signaling is involved in VEGF-protected AKT degradation (75). Moreover, mTORC2 is generally rapamycin insensitive, while the rapamycin-sensitive mTORC1 lies downstream of AKT; other groups reported that selective AKT degradation by the UPS in dendrites is required for generating neuronal polarity under physiological conditions, and those findings indicate that local protein degradation of AKT mediated by the UPS is important in determining neuronal polarity (76).

Caspase-mediated AKT cleavage

Differently from the UPS mediated AKT degradation, the mechanism by which AKT protein is cleaved by caspases is not completely clear. As a critical survival factor downstream of the receptor protein tyrosine kinases, AKT-transmitted survival signals have protective effects against apoptosis induced by a variety of stimuli. AKT protein is degraded when cells undergo apoptosis. Rokudai et al described that after incubation with active caspase, AKT was cleaved at three sites to produce 40- and 44-kDa fragments (77). As expected, the loss of the C-terminal domain of the AKT protein reduced its kinase activity; overexpression of AKT fragment with

deletion of the N-terminal domain or deletion of the C-terminal domain increased cell sensitivity to apoptosis-inducing stimuli, indicating that caspase-dependent cleavage of anti-apoptotic AKT turns off survival signals and increases apoptotic cell death (78). In interleukin-3 (IL-3)-dependent 32D cells, Xu et al discovered that cytokine withdrawal resulted in AKT degradation by caspases as well: in this study, the authors identified the Asp462 residue of AKT1 as the primary cleavage site for caspase-3. Mutation of this site (AKT1-D462N) prevented caspase cleavage. Similar to previous description of C-terminal deletion fragment, the AKT truncation mutant mimicking the caspase cleavage product lost its kinase activity, functioning as a dominant negative mutant and promoting cell death. These results showed that the balance between AKT and caspase activity controls cell survival in 32D cells: moreover, upon survival factor withdrawal, caspases are able to render AKT inactive, inhibiting AKT pathway to allow apoptosis to occur (79).

SUMOylation and AKT

SUMO (Small Ubiquitin-like modifier) conjugation or “SUMOylation” is a post-translational modification (PTM), transient and reversible, that consists in the attachment of SUMO to specific lysine residues of target proteins, mainly nuclear proteins. SUMOylation affects activity, structure and sub-cellular localization of the target proteins, promotes radical changes and plays a critical role in various cellular processes, including transcription, DNA repair/replication and cell cycle progression. Moreover, SUMOylation is involved in human disease such as neurodegenerative disorders (associated with huntingtin, ataxin-1, tau, PARK-7 or alpha-synuclein) and has been associated with cancer development, due to its cancer-related targets such as p53, pRB and Mdm2 (82).

SUMO conjugation shows some similarities with the ubiquitin pathway: SUMO needs to be cleaved to expose the glycine-glycine (GG) motif in its c-terminal (and this is performed by SUMO-specific proteases); mature SUMO is activated by the heterodimer SAE I/SAE II(E1), then it is transferred to the Ubc9 (E2) and finally Ubc9 transfers SUMO to the lysine in the target protein. While no role in proteolytic targeting has been observed, recently, SUMOylation has been demonstrated to function as a secondary signal that mediates the ubiquitin-dependent degradation (83).

During years, most of the studies about the regulation of AKT activity have been focused on its phosphorylation, and in a second moment, acetylation and ubiquitynation have been considered as regulatory modifications of this protein. Recently, Risso and colleagues described AKT as target protein of SUMOylation: in particular, SUMOylation occurs at Lys276 and Lys301 within the kinase domain of AKT and both residues are crucial for SUMO modification. They also demonstrated that different post-translational modifications don't overlap, in fact diminished sumoylation of AKT didn't prevent its phosphorylation at T308 or S473, and inhibition of PI3K did not alter basal (or Ubc9-stimulated) SUMO conjugation (84).

However, downstream activities of AKT are drastically impaired in the SUMO-deficient mutant, indicating that this PTM does not regulate the activation of the kinase, but it might regulate its interaction with downstream targets. Moreover, considering the role of AKT in the balancing between cell survival and apoptosis they demonstrated that SUMO conjugation to AKT is not only relevant for cell cycle progression at the G1/S transition, but has relevance also for the production of different mRNA splice variants, associated with cell proliferation and survival (83).

Finally, in another work, Li et al. found that this PTM influences AKT activity in cell proliferation and tumor development: they showed that diminishing

SUMO conjugation to the cancer-associated mutant AKT1 E17K reduces its oncogenic capacity (84).

Heat shock proteins and AKT stability

Molecular chaperones are a group of proteins involved in the maintenance of other “clients” proteins in folded and active conformations. The term is usually related to proteins that play a role in protein folding and refolding and derived from a large family of genes, originally known as heat shock protein (HSP) genes; within this family we can find HSPA (Hsp70-Hsc70), HSPB (small HSP), HSPD (Hsp60), HSPC (Hsp90) and HSPH (large HSP) (86, 87).

These proteins conduct the folding of much of the proteome, with the formation of proteins (or complexes) able to direct several functions in the cell. Some of these proteins are also expressed at high levels after proteotoxic stresses such as exposure to high temperatures, heavy metals, alcohols or chemical agents: thus, they are mostly recognized as heat shock proteins.

As a result of different proteotoxic stresses levels of unfolded, aggregated and ubiquitinated proteins increase and cells respond to insults through abundant synthesis of HSPs. These proteins are also known to be involved in cell survival after stress in two way: both through direct chaperoning of misfolded proteins, as well as inhibition of programmed cell death. Moreover, alterations in molecular chaperones regulation are also associated with human diseases (88).

Among molecule chaperones, Hsp90 is one of most important proteins for the cancer cell survival. It contains a unique nucleotide binding-domain at its amino terminal pocket that binds ATP and has ATP hydrolysing activity. Hsp90 forms the basis of a super-chaperone machine that promotes the

proper folding of client proteins so that they can respond to a stimulus or bind a ligand. ATP hydrolysis and ADP/ATP nucleotide exchange drive the cycling of the Hsp-90–based chaperone machine: in fact, the half-life of a client protein depends by the period that it remains in association with the complex Hsp-90/Hsp-70, because during this time, the client protein is susceptible to ubiquitination and delivery to the proteasome where it is degraded (89, 90, 91). Client proteins of Hsp90 consist in various key components of multiple signaling pathways active in cancer cells to maintain growth and/or survival, and among these, PI3K/AKT/mTOR (92, 93). Hsp90 also stabilizes AKT and prevents AKT from PP2A-mediated inactivation. When cells are exposed to heat shock, AKT is activated: the heat shock-induced AKT activation is PI3K dependent, as blocking PI3K leads to rapid dephosphorylation of AKT and detachment of AKT from Hsp90. This event increases AKT sensitivity to dephosphorylation mediated by PPA and subsequent degradation (94). Moreover, blockade of the AKT-HSP-90 complex formation by the specific inhibitor 17-AAG (a geldanamycin analogue that is undergoing clinical testing in malignant melanoma and breast and prostate cancers) induces AKT dephosphorylation and degradation. In addition, 17-AAG can block the intrinsic ATPase activity of Hsp90 and subsequently block the formation of a multi-chaperone complex, including the AKT-Hsp90 complex. However, it is not yet clear in which patho-physiological conditions the ubiquitin proteasome pathway will recycle or degrade Hsp90-bound AKT (95).

2.6. AKT in cancers

The AKT/PKB (protein kinase B) kinases, which include AKT1, AKT2, and AKT3, are known to play an important role as intermediates of signaling pathways that regulate cellular processes as proliferation, survival, glucose metabolism, genome stability and neo-vascularization.

When deregulated they can contribute in development of cancer: moreover, AKT has been found mutated or hyperactivated in various solid tumors and haematological malignancies, underlining its crucial role in cancer progression, where AKT activation correlates with advanced disease or poor prognosis. Some studies evidenced that approximately 40% of breast and ovarian cancers and more than 50% of prostate carcinomas had increased AKT1 activity, while in other studies, the activation of the AKT2 kinase was observed in 30–40% of pancreatic and ovarian cancers. Further, elevated AKT3 activity has been reported in oestrogen receptor-deficient breast cancer and androgen-insensitive prostate cancer cell lines, suggesting that also AKT3 may contribute to the aggressiveness of these carcinomas (96).

2.6.1. AKT in malignant mesothelioma

As in many other cancers, the PI3K/AKT pathway plays a critical role for the cell cycle progression in human MPM cells and it has been reported that inhibition of the PI3K activity leads to significant cell cycle arrest and suppression of proliferation of different MPM cell lines.

Activation of AKT, as described by Altomare et al. was observed in 65% of MPM specimens but no frequent genetic alterations were found for PI3K/AKT activation in mesothelioma cells (97).

Recently our lab has firstly described that mesothelioma cells express the two isoforms of AKT, AKT1 and AKT3, while they completely lack of AKT2. The role and the relevance of AKT kinases in MPM are described, but no data are available on AKT1/3 expression and function in this kind of tumor.

The PI3K/AKT signaling pathway, aberrantly active and involved in MPM cell cycle progression and chemo-resistance, represents a novel good therapeutic target. Various studies have also demonstrated that growth of tumor cells with elevated AKT activity is more sensitive to targeted

disruption of AKT signaling; moreover, these evidences suggest that targeting AKT activity sensitizes MPM cells to conventional drugs such as cisplatin, and that sensitivity after AKT inhibition might be more effective in cells with an elevated AKT activity. In a work recently published by our group, AKT was investigated as a relevant target of perifosine; it has been described that perifosine inhibits both AKT1 and AKT3 phosphorylation and activation in cells growing exponentially in serum-containing medium and after growth factor stimulation (98).

2.6.2. AKT in ovarian cancer

The involvement of AKT in cancer was firstly described in 1992 when Cheng et al. showed the hyperactivation of AKT2 in ovarian cancer cell lines. Ovarian cancer is the fifth most common cause of death due to cancer in women; therapy is successful at first, but the recurrence of the disease makes the rate of survival low. In ovarian cancer, the PI3K/AKT/mTOR pathway is frequently mutated or amplified, underlining its role in progression, cell proliferation, migration and chemoresistance of the tumor (99).

Different studies report the expression of AKT in ovarian cancer: in particular, AKT1 and AKT2 were found activated and/or over-expressed, while AKT3 was detected up-regulated in over 20% of primary ovarian tumors and involved in the regulation of the G2/M transition.

Tang et al. reported that AKT1 is frequently activated in ovarian cancer, while AKT2 has been shown to be amplified and overexpressed in human ovarian carcinoma cell lines and primary ovarian tumors, as described by Bellacosa and colleagues. Moreover, amplification of AKT2 was particularly frequent in undifferentiated ovarian tumors, suggesting a correlation between AKT alterations and tumor aggressiveness (100).

The main biological functions of AKT activation are anti-apoptotic and pro-proliferative in cancer cells; proliferation and invasion are also affected when AKT is directly targeted as well: silencing of AKT1 isoform reduces proliferation and invasion of OVCAR-3 cells, while targeting the AKT2 isoform has been shown to increase the activation of apoptosis (101).

2.6.3. AKT in prostate cancer

The PI3K/AKT/mTOR pathway has a pivotal role in prostate cancer and is estimated to be up-regulated in 30-50% of the cases, often associated with PTEN loss. Furthermore, phospho-AKT levels were also significantly greater in high-grade prostate tumors if compared to low/intermediate grade tumors, as different studies showed (102). In fact, according to the Gleason score, a grading assigned to a prostate tumor with values between 2 and 10, when lowest numbers indicate slow-growing tumor and the highest ones indicate an aggressive tumor, phospho-AKT was detected mostly in tumor samples with Gleason score ≥ 8 (103). Levels of phospho-AKT were significantly increased in cancer cells relative to normal prostate epithelium and benign prostatic hyperplasia. In normal tissue AKT isoforms are present in different percentage, with the prevalence of AKT2 and AKT3, while changes in expression and activation of AKT have been reported in prostate cancer. Recent studies showed that silencing of AKT2 led to an increasing of migration, allowing epithelial-mesenchymal transition (EMT); on the contrary, silencing of other two isoforms did not evolve in such consequences. Moreover, down-regulation of AKT1 and AKT2, but not AKT3, induced activation of cell surface $\beta 1$ -integrins and enhanced adhesion, migration and invasion; while silencing of AKT1 and AKT2 resulted in increased focal adhesion size (104).

2.6.4. AKT in lung cancer

Lung cancer is one of the most common human cancers and non-small cell lung cancer (NSCLC) represents around 80% of deaths caused by all primary lung cancers. Due to diagnosis at late stages of the disease and despite advances in surgery and other treatments, the 5-year rate of survival for patients with NSCLC is only 15%. The annual incidence worldwide is more than 1.3 million lung cancer cases, while more than 1.1 million lung cancer deaths per year are estimated (105). Researches based on molecular cancer biology and biochemical alterations evidenced that deregulation of some pathways involved in cellular proliferation and survival are responsible for cancer progression and among these, a pivotal role is played by the phosphoinositide-3-kinase/protein kinase B (AKT) signaling pathway.

NSCLC cells express all the three AKT isoforms; in accordance with results published by different groups, there is a prevalence of AKT1 activation in all NSCLC subtypes. Regarding AKT1 and AKT3 activation, no significant difference between expression in tumor cores and control cores was observed, whereas PTEN and PI3K showed a significant difference in expression; in addition, high expression of PI3K was observed in 29% of all tumor core (106). Recently new findings studies have reported a lack of association between activated AKT and survival or clinic-pathological variables such as stage of disease and metastasis. In 102 NSCLC cases, Tang et al. found that patients with concomitant p-AKT Ser473 expression and loss of PTEN had a significantly reduced 5-year survival rate, while other research groups have reported the prevalence of phosphorylated AKT and its relevance to prognosis in NSCLC. Moreover, others studies observed high non-phosphorylated AKT2 expression in epithelial cells and considered it a positive prognosticator (107).

2.6.5. AKT in breast cancer

Breast cancer is the most common malignancy among women, and it is the illness most frequently diagnosed in developed countries. Mutations in a number of genes are now known to cause susceptibility to breast cancer. In high-risk families, the most significant of these are the BRCA1 and BRCA2 genes, while the human epidermal growth factor receptor 2 gene (HER2) is overexpressed and/or amplified in ~15% of breast cancer patients and has been identified as a marker of poor prognosis (108). Other genes are involved in the genesis of breast cancer as well; these are tumor suppressor genes as p53 and PTEN: in particular, mutations in components of PI3K pathway and AKT hyper-activation are found in 70% of breast cancers. Studies in transgenic mice have shown that AKT promotes mammary tumor progression by increasing cell survival. The majority of the currently available in vivo data agree with the overall model whereby AKT1 and AKT2 kinases play opposite roles in breast cancer migration, invasion and metastatic dissemination (events that ultimately determine the clinical outcome): as several studies reported, AKT1 functions as an inhibitor of migration and invasion in breast, while AKT2 acts in a positive manner (109). Moreover, AKT activation is associated with poor outcome in endocrine-treated breast cancer, whereas high levels of cytoplasmic AKT2 are associated with an improved overall survival. AKT signaling pathway regulates also IKK ϵ , an inducible kinase and is amplified and overexpressed in breast, and in particular AKT2, but not the AKT1 or AKT3 isoform, is responsible for IKK ϵ induction (110).

2.6.6. AKT in skin cancer

Malignant melanoma is a devastating tumor of the skin, with poor prognosis for patients (cause metastasis spread early) and resistant to all current treatments including chemo-, immuno-, or radiation therapy. In melanoma,

both the Ras/Raf/MEK/ERK (MAPK) and the PI3K/AKT (AKT) signaling pathways are constitutively activated and represent a crucial point for cancer development and progression; in particular, PTEN deletion leads to AKT activation that promote cell survival and can results in downstream loss of differentiation or senescence, through Raf-ERK cascades.

Members of the PI3K and AKT3 signaling cascades have been implicated in initiation, progression, invasive, and drug resistance phenotypes of melanomas. Increased phospho-AKT expression in melanoma is associated with tumor progression and shorter survival: recently, has been also described that AKT3 in early melanocytic lesions can phosphorylate ^{V600E}BRAF reducing its activity and promoting proliferation to overcome the senescence block (111).

Furthermore, the discovery of an AKT3 mutation in melanoma underlined the critical role of this isoform. Stahl et al (2004) described that AKT3 protein, but not AKT1 or AKT2, was increased in melanoma cell lines when compared to normal melanocytes, while other studies reported that PTEN loss cooperates in increasing melanoma cell and non-transformed melanocytes capabilities of invasion and migration, by enhancing AKT2 activity and through E-Cadherin down-regulation. Interestingly, targeting PTEN differentially regulates AKT3- mediated cell survival and AKT2-mediated metastasis in melanomas (112).

Expression of AKT3 has been detected elevated in cell lines derived from primary melanoma tumors at the radial and vertical stages of cell growth compared to normal melanocytes; on the contrary in the same samples, no significant changes were observed in the levels of AKT1 and AKT2, indicating that AKT3 expression is predominant in melanoma development. Although a central role of the AKT3 isoform in melanoma maturation is well established, this evidence still remain controversial: a recent report using biopsy of melanoma found AKT2 as a predominantly activated isoform in

melanomas, while a different study demonstrated that loss of PTEN promoted melanoma cell metastasis via activating AKT2 but not AKT1 or AKT3 (113, 114).

2.6.7. AKT in thyroid cancer

The Cowden's syndrome, an autosomal dominant disorder characterized by the onset of various type of cancer affecting breast, colon, skin and thyroid, has been connected to the inactivation of the tumor suppressor PTEN, a phosphatase that regulates negatively the PI3K signaling, with the consequent constitutively activation of AKT. Although these observations, PTEN inactivation resulted to be sporadic in thyroid cancer, while AKT might be a central regulator of many thyroid oncogenes.

In normal thyroid cells AKT activation is the major mediator of cell growth and inhibition of apoptosis; overexpression of an active form of AKT1 in thyroid cells results in serum-independent growth and leads to resistance to cell death, although it seems insufficient to transform the thyroid cells: this evidence suggests that additional signaling pathways are needed for developing thyroid cancer. Other recent study demonstrated that AKT signaling is enhanced in human thyroid cancer, in particular that AKT1 and AKT2 protein levels are increased in follicular thyroid cancer if compared with normal thyroid tissue as well as the level of phospho-AKT. Finally, among the isoforms, AKT2 results having an important role in the pathogenesis and/or progression of thyroid cancer (115).

2.6.8. AKT in colon cancer

Colorectal cancer (CRC) represents one of the most common malignancies in the world; it undergoes a multistage carcinogenesis pathway from adenomatous polyps to carcinoma and despite advances in chemotherapy, this tumor remains a major cause of death. Genetic events that lead to

neoplastic processes include alterations in tumor suppressor genes that in normal conditions regulate cell cycle progression, or somatic mutations in the TGFb receptor and the K-ras oncogene (116).

Activation of AKT signaling and impaired expression of phosphatase and tensin homolog (PTEN) has been reported in 60-70% of human CRC and leads to inhibition of the pro-apoptotic GSK3b, with resulting increase in the levels of the anti-apoptotic beta catenin protein. AKT is up regulated in 57% of colon cancer, rate markedly higher than in other malignancies; moreover, AKT can act synergically with RAS and RAF cascades, both critical in colorectal carcinogenesis, and has been shown to impact also on WNT signaling, to regulate expression of the anti-apoptotic COX2 protein and to modulate cellular motility enhanced by prostaglandin E2. Although mutations in AKT genes are not frequently found in CRC, a somatic missense mutation of AKT1 (E17K) in the PH domain has been described and resulted in constitutive association of AKT1 with the plasma membrane, with its prolonged activation and mTOR deregulation. Finally, AKT proto-oncogene is overexpressed in cancer suggesting that its overexpression may be crucial in earliest stages of carcinogenesis process (117, 118).

2.6.9. AKT in glioma

Malignant glioma is the most common primary brain tumor in adults; it originates from the supportive cells of the brain (glial cells) and carries a poor prognosis. The median survival of patients with high-grade glioma (World Health Organization [WHO] grade III or IV) is 10-30 months despite multimodal treatment with surgery, radiation, and chemotherapy.

Malignant gliomas develop because of the consequent accumulation of alterations in genes regulating cell proliferation, differentiation, and apoptosis. The amplification and mutation of tyrosine kinase receptors (i.e.

EGFR) activates several signaling pathways including the PI3K/AKT pathway that delivers major survival signals to human malignant glioma.

AKT is activated in 70% of gliomas and is usually associated with PTEN mutations. Gliomas express all the AKT isoforms in different ratios; according to the WHO classification, the expression of AKT1 protein was not associated with the pathological glioma grade, while AKT2 expression was notably higher in tissues from grade III and IV than in normal tissues. On the contrary, AKT3 protein diminished with increasing glioma grades (119).

Moreover, AKT3 was found down-regulated in malignant glioma cell lines overexpressing AKT2. In a recent study, Mure et al. observed the induction of apoptosis via the activation of caspase-9 and caspase-3 in glioma cell lines, after silencing of AKT2 or AKT3, while the combined knock-down of AKT2 and AKT3 effectively inhibited the growth of malignant cells, demonstrating the pivotal role of the two isoforms in the biology of this cancer (120).

2.7. AKT Inhibitors

Based on the evidence that AKT pathway has a pivotal role in various cellular functions and is one of the most frequently activated pathway in several type of cancers, the advent of targeted agents represents a new possibility to overcome the plateau achieved by current chemotherapeutic treatments.

The three isoenzymes of AKT are not functionally identical, but their sequence is still similar to other kinases of the AGC family (>90% homology among AKT isoforms and >50% with PKA- α and PKC- α). Moreover, when considering the homology between ATP-binding site in AKT1 and AKT2 we reach almost the 100% (96% of homology with AKT2/3 and more than 70% with PKA- α / PKC- α). All these observations underline how the

development of specific inhibitors could represent an open challenge for researchers (121, 122).

AKT inhibitors can be divided into four major classes based on their mechanisms of action and target regions.

- 1) INHIBITORS OF THE KINASE DOMAIN (ATP-competitive inhibitors); it seems to be the most obvious target for therapeutics, but it is really the most difficult one to reach. Each kinase domain of AKT ends in a regulatory hydrophobic motif, but because of the high homology, the majority of them occupy entirely the ATP-binding site and acts as pan-AKT inhibitors. Numerous studies of AKT kinase inhibitors with reasonable selectivity and promising efficacy have been carried on. Luo et al., from a computational approach based on structure, generated a group of indazole-pyridine based pan-AKT inhibitors; among those, the most potent is A-443654 and targets the ATP-binding pocket with equal potency against AKT1, AKT2, or AKT3 within cells, but is more selective against AKT than other kinases in the AGC family (123). This class includes also compounds such as CCT128930 and GDC-0068 that inhibit respectively AKT2 and AKT1 (the latter entered in Phase II trials) and the pan-AKT inhibitor GSK2110183 (afuresertib, itself in Phase II trials).

GlaxoSmithKline created another AKT inhibitor, **GSK690693** (AKT1: IC₅₀= 2nM, AKT2: IC₅₀= 13nM and AKT3: IC₅₀= 9nM); it is aminofurazan-derived, ATP-competitive inhibitor. This molecule showed selective anti-proliferative effects in malignant cells and also downstream AKT phosphorylation events were reduced. In murine xenograft models, this agent led to a significant growth inhibition of tumors SKOV-3 cells (ovarian), LNCaP cells (prostate) and BT474 cells (breast). Despite these encouraging preclinical data, clinical development of the agent was halted due to results from a phase I

trial testing an IV formulation of the agent, where studies on mice indicate that GSK690693 may inhibit glycogen synthesis and activate glycogenolysis, suggesting also that the inhibitor may induce peripheral insulin resistance (124, 125).

- 2) INHIBITORS OF PIP3 BINDING-SITE; this class contains lipid-based AKT inhibitors which act by preventing the generation of PIP3 by PI3K. This mechanism is used by phosphatidylinositol analogues such as Calbiochem AKT Inhibitors I, II and III or other PI3K inhibitors such as PX-866. This class comprises Triciribine-phosphate (TCN-P) that inhibits phosphorylation of AKT at Thr308 and Ser473, as well as downstream p70S6K: it is highly selective for AKT and does not inhibit the activation of other kinases such as PI3K, PDK-1, PKC and PKA. In order to overcome the little specificity of inhibitors caused by the homology in ATP-binding pocket, targeting the PH domain to interfere with its binding to PIP3 and membrane translocation became a good alternative to therapeutic compounds. AKT can be activated through binding to 3' - OH phosphorylated phosphatidylinositols via the PH domain. In order to block this interaction, a series of structurally modified phosphatidylinositol ether lipid analogues (PIAs) were drawn by computer-based modelling and synthesized. The best-characterized inhibitor of AKT is **perifosine** [octadecyl-(1,1-dimethyl-piperidinio-4-yl)-phosphate], a synthetic novel alkylphospholipid (ALP) agents, which targets cell membranes: *in vitro*, perifosine interferes with the recruitment of AKT to the cell membrane and inhibits the growth of melanoma, lung, prostate, colon, and breast cancer cells in association with inhibition of AKT activity (126, 127).

Important, perifosine does not directly affect activity of PI3-K or PDK1; it induces p21^{Cip1}, associated with G₂/M phase cell accumulation, in head and neck squamous cell carcinoma cells. Most recently, combination treatment of human leukemia cells with perifosine and histone deacetylase inhibitors (HDACs) inhibited both AKT and MEK/ERK, induced ROS production and mitochondrial injury, and triggered apoptosis; moreover, perifosine was shown to sensitize tumor cells to conventional therapeutic antitumor agents and radiation(128, 129).

- 3) ALLOSTERIC INHIBITORS OF AKT KINASE DOMAIN; **MK-2206** is an investigational allosteric inhibitor of AKT that requires the PH domain of AKT for activity, but does not interact with the ATP binding pocket. Studies reported that MK-2206 is highly selective for AKT inhibition, has higher potency against recombinant human AKT1 and AKT2 isoforms than AKT3, has little off-target kinase activities, and is less vulnerable to feedback activation of AKT compared with ATP-competitive inhibitors(130). In preclinical models, MK-2206 used in association with other targeted therapies showed synergy: for example, combination of MK-2206 with *erlotinib* in NSCLC cell lines and with *lapatinib* in breast cancer cell lines led to an important synergistic inhibition of growth. In addition, in xenograft studies utilizing mice bearing the A2780 ovarian cancer cell line, treatment with MK-2206 led to strong 60% growth inhibition together with sustained inhibition of all three AKT isoforms (occurring at nanomolar concentrations - AKT1=8nM, AKT2=12nM, AKT3=65nM) (131).

- 4) **PSEUDO-SUBSTRATE INHIBITORS**; this class includes compounds that bind the sites of catalytic domain in peptide/protein substrate. They are selective for many kinases and this characteristic derives from the much larger peptide-kinase contact region involved to discriminate various substrates. One of this compounds is **AKTide-2T**, that binds to the substrate binding site of AKT1 and inhibits its activity. Another hybrid peptide was recently generated this sequence and a sequence of the forkhead transcription factor FOXO3. Replacement of a putative phosphorylation site serine (Ser) in the sequence with an alanine (Ala) resulted in a further 10-fold improvement of potency. Fusion peptides were also constructed to allow cell uptake and demonstrated dose-dependent inhibition of GSK3b phosphorylation: although these peptides are selective inhibitors of AKT, their size makes them poor leads for consequent development (132).

Another peptide-based approach is the development of a single-chain antibody (scFv) against AKT such as **GST-antiAKT1-MTS**. This antibody is the first genetically engineered scFv against AKT with inherent cell membrane translocation activity and detention of AKT inhibitory function associated with induction of apoptosis in vivo. Although the concept of peptide-based inhibitors seems promising, their development is currently hampered by their low bioavailability and stability in vivo (133).

Apart from these, other molecules such as **RX-0201**, have it also yielded favorable results: RX-0201 represents an antisense oligonucleotide (AO) to mRNA encoding AKT1. In vitro, nanomolar concentrations of resulted in growth inhibition of various human cancer cell lines. When tested in two in vivo models (glioblastoma cells U251 or pancreatic cells MIA were implanted in nude mice), strong growth inhibition was also observed. In the radiation resistant H1299 NSCLC cell line, treatment with AKT1-AO

resulted in cytotoxicity. These encouraging studies led to further clinical development of the agent: a preliminary report of a Phase I trial with RX-0201 included 17 patients with advanced solid tumors, in which the outcome was positive: moreover, phase II clinical trials are planned and a liposomal formulation is under development (134). Considering the wide quantity of new drugs discovered and under development, is necessary to define if they encounter the actual medical management; several current studies are assessing the safety of AKT inhibitors alone or in combination with both traditional chemical agents and other targeted therapies and these new findings could allow novel AKT inhibitors to complete existing treatments.

References:

1. Robinson BM 2012. Malignant pleural mesothelioma: an epidemiological perspective. *Ann Cardiothorac Surg*1(4):491-6
2. Pass HI, Vogelzang N, Hahn S, Carbone M 2004. Malignant pleural mesothelioma. *Curr Probl Cancer* 28:93-17
3. Robinson BW, Musk AW, Lake RA 2005. Malignant mesothelioma. *Lancet* 366:397-408
4. Ismail-Khan R, Robinson LA, Williams Jr CC, et al 2006. Malignant pleural mesothelioma: a comprehensive review. *Cancer Contr* 13:255-63
5. Heintz NH, Janssen-Heininger Yvonne MW, Mossman BT 2010. Asbestos, lung cancers, and mesotheliomas: from molecular approaches to targeting tumor survival pathways. *American J of Res Cell and Mol Biol* 42(2):133-9
6. Wagner JC, Sleggs CA, Marchand P 1960. Diffuse pleural mesothelioma and asbestos exposure in the North Western Cape Province. *Br J Ind Med* 17:260-71

7. Nagai H and Toyokuni S 2012. Differences and similarities between carbon nanotubes and asbestos fibers during mesothelial carcinogenesis: shedding light on fiber entry mechanism. *Cancer Sci* 103(8):1378-90
8. Fazzo L, De Santis M, Minelli G, et al 2012. Pleural mesothelioma mortality and asbestos exposure mapping in Italy. *Am J Ind Med* 55:11-24
9. Mc Elvenny DM, Darnton AJ, Price MJ, et al 2005. Mesothelioma mortality in Great Britain from 1968 to 2001. *Occup Med (Lond)* 55:79-87
10. Fennell DA, Gaudino G, O'Byrne KJ, et al 2008. Advances in the systemic therapy of malignant pleural mesothelioma. *Nat Clin Pract Oncol* 5 (3):136-47
11. Carbone M, Kratzke RA, Testa JR 2002. The pathogenesis of mesothelioma. *Semin Oncol* 29:2-17
12. Bocchetta M, Di Resta I, Powers A, et al 2000. Human mesothelial cells are unusually susceptible to simian virus 40-mediated transformation and asbestos cocarcinogenicity. *Proc Natl Acad Sci USA* 97:10214–9
13. Frank E Mott 2002. Mesothelioma: A Review. *The Ochsner Journal* 12:70-9
14. Yoshitaka Sekido 2008. Molecular biology of malignant mesothelioma. *Environ Health Prev Med* 13:65–70
15. Yang H, Bocchetta M, Kroczyńska B et al 2006. TNF- α inhibits asbestos-induced cytotoxicity via a NF- κ B dependent pathway, a possible mechanism for asbestos induced oncogenesis. *Proc Natl Acad Sci USA* 103:10397-402
16. Hammar SP 1992. Controversies and uncertainties concerning the pathologic features and pathologic diagnosis of asbestosis. *Semin Diagn Pathol* 9(2):102-9
17. Van Zandwijk N, Clarke C, Henderson D, et al 2013. Guidelines for the diagnosis and treatment of malignant pleural mesothelioma. *J Thorac Dis* 5(6): E254-E307

18. Travis W, Brambilla E, Muller-Hermelink H, Harris C 2004. World Health Organization Classification of Tumours. *Pathology and Genetics. Tumours of the Lung, Pleura, Thymus, and Heart*. IARC Press: Lyon, France, 125-44
19. William D. Travis 2010. Sarcomatoid Neoplasms of the Lung and Pleura. *Arch Pathol Lab Med* 134:1645-58
20. Ceresoli GL, Gridelli C and Santoro A 2007. Multidisciplinary Treatment of Malignant Pleural Mesothelioma. *The Oncologist* 12:850-863
21. Butchart EG, Ashcroft T, Barnsley WC, et al 1976. Pleuropneumectomy in the management of diffuse malignant mesothelioma of the pleura. Experience with 29 patients. *Thorax* 31:15-24
22. Sugarbaker DJ, Strauss GM, Lynch TJ, et al 1993. Node status has prognostic significance in the multimodality therapy of diffuse, malignant mesothelioma. *J Clin Oncol* 11:1172-78
23. Cao C, Tian D, Manganas C, et al 2012. Systematic review of trimodality therapy for patients with malignant pleural mesothelioma. *Ann Cardiothorac Surg* 1(4):428-37
24. Rusch VW and Giroux D 2012. Do we need a revised staging system for malignant pleural mesothelioma? Analysis of the IASLC database. *Ann Cardiothorac Surg* 1(4):438-48
25. Bearz A, Talamini R, Rossoni G, et al 2012. Re-challenge with pemetrexed in advanced mesothelioma: a multi-institutional experience. *BMC Res Notes* 5:482
26. American Joint Committee on Cancer. Pleural mesothelioma. *AJCC Cancer Staging Manual*. 7th ed. New York, NY: Springer; 2010: 271-4.
27. Rusch VW 1997. Pleurectomy/decortication in the setting of multimodality treatment for diffuse malignant pleural mesothelioma. *Semin Thorac Cardiovasc Surg* 9:367-72

28. Ismail-Khan R, Lary A. Robinson LA, Williams CC Jr, et al 2006. Malignant Pleural Mesothelioma: A Comprehensive Review. *Cancer Control* 13(4):255-63
29. Turner KM, Varghese S, Alexander HR Jr 2011. Surgery for peritoneal mesothelioma. *Curr Treat Options Oncol* 2(2):189-200
30. Paul H. Sugarbaker 2013. Cytoreductive surgery using peritonectomy and visceral resections for peritoneal surface malignancy. *Transl Gastrointest Cancer* 2(2):54-74
31. Rimner A, Rosenzweig KE 2012. Novel radiation therapy approaches in malignant pleural mesothelioma. *Ann Cardiothorac Surg* 1(4):457-61
32. Zierhut D, Gutwein S, Munter MW, et al 2004. Radiation therapy of mesothelioma: the Heidelberg experience and future aspects. *Lung Cancer* 45: S85-S91
33. Vogelzang NJ, Rusthoven JJ, Symanowski J, et al 2003. Phase III Study of Pemetrexed in Combination With Cisplatin Versus Cisplatin Alone in Patients With Malignant Pleural Mesothelioma. *J Clin Oncol* 21 (14): 2636-44
34. Krug LM, Pass HI, Rusch VW, et al 2009. Multicenter phase II trial of neoadjuvant pemetrexed plus cisplatin followed by extrapleural pneumonectomy and radiation for malignant pleural mesothelioma. *J Clin Oncol* 27:3007-13
35. Nagio Takigawa, Katsuyuki Kiura and Takumi Kishimoto 2011. Medical Treatment of Mesothelioma: Anything New? *Curr Oncol Rep* 13:265-71
36. Rea F, Favaretto A, Marulli G, et al 2013. Phase II trial of neoadjuvant pemetrexed plus cisplatin followed by surgery and radiation in the treatment of malignant pleural mesothelioma. *BMC Cancer* 13:22
37. Zucali PA, Ceresoli GL, Garassino I, et al 2008. Gemcitabine and vinorelbine in pemetrexed-pretreated patients with malignant pleural mesothelioma. *Cancer* 112:1555-61

38. Jassem J, Ramlau R, Santoro A, et al 2008. Phase III trial of pemetrexed plus best supportive care compared with best supportive care in previously treated patients with advanced malignant pleural mesothelioma. *J Clin Oncol* 26:1698-704
39. R Cornelissen, ME Heuvers, AP Maat, et al 2012. New Roads Open Up for Implementing Immunotherapy in Mesothelioma. *Clin Dev Immunol* 2012:1-13
40. Tol J, Punt CJA 2010. Monoclonal antibodies in the treatment of metastatic colorectal cancer: a review. *Clinical Therapeutics* 32(3): 437-53
41. Powell A, Creaney J, Broomfield S, et al 2006. Recombinant GM-CSF plus autologous tumor cells as a vaccine for patients with mesothelioma. *Lung Cancer* 52(2):189-97
42. Pasello G, Favaretto A 2009. Molecular targets in malignant pleural mesothelioma treatment. *Curr Drug Targets* 10:1235-44
43. Paik PK, Krug LM 2010. Histone deacetylase inhibitors in malignant pleural mesothelioma: preclinical rationale and clinical trials. *J Thorac Oncol* 5:275-9
44. Agarwal V, Lind MJ, Cawkcwell L 2011. Targeted epidermal growth factor receptor therapy in malignant pleural mesothelioma: Where do we stand? *Cancer Treatment Reviews* 37:533-42
45. Govindan R, Kratzke RA, Herndon JE, et al 2005. Gefitinib in patients with malignant mesothelioma: a phase II study by the Cancer and Leukemia Group B. *Clin Cancer Res* 11:2300-4
46. Jackman DM, Kindler HL, Yeap BY, et al 2008. Erlotinib plus bevacizumab in previously treated patients with malignant pleural mesothelioma. *Cancer* 113(4):808-14
47. Zellos LS, Sugarbaker DJ 2002. Multimodality treatment of diffuse malignant pleural mesothelioma. *Semin Oncol* 29:41-50
48. Stahel R, Weder W 2005. Neoadjuvant chemotherapy in malignant pleural mesothelioma. *Lung Cancer* 49(suppl 1):S69-S70

49. Carnero A 2010. The PKB/AKT Pathway in Cancer. *Current Pharmaceutical Design* 16:34-44
50. Datta SR, Brunet A, Greenberg ME 1999. Cellular survival: a play in three Akts. *Genes Dev* 13:2905-27
51. Kandel ES, Hay N 1999. The regulation and activities of the multifunctional serine/threonine kinase Akt/PKB. *Exp Cell Res* 253:210-29
52. Yang ZZ, Tschopp O, Hemmings-Mieszczak M, et al 2003. Protein kinase B alpha/Akt1 regulates placental development and fetal growth. *J Biol Chem* 278:32124-31
53. Cho H, Kim JK, et al 2001. Insulin resistance and a diabetes mellitus-like syndrome in mice lacking the protein kinase Akt2 (PKB beta). *Science* 292(5522):1728-31
54. Easton RM, Cho H, Roovers K et al 2005. Role for Akt3/protein kinase B gamma in attainment of normal brain size. *Mol Cell Biology* 25:1869-78
55. Chan TO, Rittenhouse SE and Tsichlis PN 1999. AKT/PKB and other D3 phosphoinositide regulated kinases: kinase activation by phosphoinositide-dependent phosphorylation. *Annu Rev Biochem* 68: 965-1014
56. Yong Liao and Mien-Chie Hung 2010. Physiological regulation of Akt activity and stability. *Am J Transl Res* 2(1):19-42 Review Article
57. Yuan TL and Cantley LC 2008. PI3K pathway alterations in cancer: variations on a theme. *Oncogene* 27:5497-510
58. Alessi DR, Pearce LR and Garcia-Martinez JM 2009. New insights into mTOR signaling: mTORC2 and beyond. *Sci Signal* 2:pe27
59. Yang J, Cron P, Thompson V, et al 2002. Molecular mechanism for the regulation of protein kinase B/Akt by hydrophobic motif phosphorylation *Mol Cell* 9:1227-40
60. Manning BD and Cantley LC 2007. AKT/PKB signaling: navigating downstream. *Cell* 129:1261-74

61. Huang J and Manning BD 2009. A complex interplay between Akt, TSC2 and the two mTOR complexes. *Biochem Soc Trans* 37:217-22
62. Carpten JD, Faber AL, Horn C, et al 2007. A transforming mutation in the pleckstrin homology domain of AKT1 in cancer. *Nature* 448: 439-44
63. Brognard J, Sierrecki E, Gao T and Newton AC 2007. PHLPP and a second isoform, PHLPP2, differentially attenuate the amplitude of Akt signaling by regulating distinct Akt isoforms. *Mol Cell* 25:917-31
64. Freiman RN, Tjian R 2003. Regulating the regulators: lysine modifications make their mark. *Cell* 112:11-17
65. Yang XJ, Seto E 2008. Lysine acetylation: codified crosstalk with other posttranslational modifications. *Mol Cell* 31:449-61
66. Pillai VB, Sundaresan NR, Samant SA, et al 2011. Acetylation of a Conserved Lysine Residue in the ATP Binding Pocket of p38 Augments Its Kinase Activity during Hypertrophy of Cardiomyocytes. *Mol Cell Biol* 31(11):2349-63
67. Sundaresan NR, Pillai VB, Wolfgeher D, et al 2001. The deacetylase SIRT1 promotes membrane localization and activation of Akt and PDK1 during tumorigenesis and cardiac hypertrophy. *Sci Signal* 4: ra46
68. Zhao Zhong Chong, Yan Chen Shang, Shaohui Wang, Kenneth Maiese 2012. SIRT1: new avenues of discovery for disorders of oxidative stress. *Expert Opin Ther Targets* 16(2):167-78
69. Ramakrishnan G, Davaakhuu G, Kaplun L, et al 2014. Sirt2 Deacetylase Is A Novel AKT Binding Partner Critical For AKT Activation By Insulin. *J Biol Chem* Jan 20
70. Yong Liao, Mieu-Chien Hung 2010. Physiological regulation of Akt activity and stability. *Am J Trans Res* 2(1):19-42 Review Article
71. Herrmann J, Lerman LO and Lerman A 2007. Ubiquitin and ubiquitin-like proteins in protein regulation. *Circ Res* 100:1276-91

72. Thompson SJ, Loftus LT, Ashley MD and Meller R 2008. Ubiquitin-proteasome system as a modulator of cell fate. *Curr Opin Pharmacol* 8:90-95
73. Jung T, Catalgol B and Grune T 2009. The proteasomal system. *Mol Aspects Med* 4:191-296
74. Kim B and Feldman EL 2002. Insulin-like growth factor I prevents mannitol-induced degradation of focal adhesion kinase and Akt. *J Biol Chem* 277:27393-00
75. Riesterer O, Zingg D, Hummerjohann J, et al 2004. Degradation of PKB/Akt protein by inhibition of the VEGF receptor/mTOR pathway in endothelial cells. *Oncogene* 23:4624-35
76. Yan D, Guo L and Wang Y 2006. Requirement of dendritic Akt degradation by the ubiquitin-proteasome system for neuronal polarity. *J Cell Biol* 174:415-24
77. Bachelder RE, Wendt MA, Fujita N, et al 2001. The cleavage of Akt/protein kinase B by death receptor signaling is an important event in detachment-induced apoptosis. *J Biol Chem* 276:34702-07
78. Rokudai S, Fujita N, Hashimoto Y and Tsuruo T 2000. Cleavage and inactivation of antiapoptotic Akt/PKB by caspases during apoptosis. *J Cell Physiol* 182:290-96
79. Xu J, Liu D and Songyang Z 2002. The role of Asp-462 in regulating Akt activity. *J Biol Chem* 277:35561-66
80. KA Wilkinson and JM Henley 2010. Mechanisms, regulation and consequences of protein SUMOylation. *Biochemical Journal* 428(2):133-45
81. Hochstrasser M 2001. SP-RING for SUMO: new functions bloom for a ubiquitin-like protein. *Cell* 107:5-8
82. Da Silva-Ferrada E, Lopitz-Otsoa F, Lang V, et al 2012. Strategies to Identify Recognition Signals and targets of SUMOylation. *Biochem Res Intern* 2012:875148 Review Article

83. Gareauand JR andLima CD 2010. The SUMO pathway: emerging mechanisms that shape specificity, conjugation and recognition. *Nat Rev Mol Cell Biol* 11(12):861-71
84. Risso G, Pelisch F, Pozzi B, et al 2012. Modification of Akt by SUMO conjugation regulates alternative splicing and cell cycle. *Cell Cycle* 19:3165-74
85. Li R, Wei J, Jiang C, et al 2013. AKT SUMOylation regulates cell proliferation and tumorigenesis. *Cancer Res* 73(18):5742-53
86. Houry WA 2009. Chaperone-assisted protein folding in the cell cytoplasm. *Curr Protein Pept Sci* 2:227-44
87. Kampinga HH, Hageman J, Vos MJ, et al 2009. Guidelines for the nomenclature of the human heat shock proteins. *Cell Stress Chaperones* 14(1):105-11
88. Hayes D, Napoli V, Mazurkie A, et al 2009. Phosphorylation dependence of multimeric size and molecular chaperone function. *J Biol Chem* 284:18801-7
89. Whitesell L, Lindquist SL 2005. HSP90 and the chaperoning of cancer. *Nat Rev Cancer* 5:761-72
90. Isaacs JS, Xu W and Neckers L 2003. Heat shock protein 90 as a molecular target for cancer therapeutics. *Cancer Cell* 3:213-7
91. Niu G and Chen X 2009. From protein-protein interaction to therapy response: molecular imaging of heat shock proteins. *Eur J Radiol* 70:294-304
92. Neckers L and Ivy SP. Heat shock protein 90. *Curr Opin Oncol.* 2003; 15: 419-24
93. Davenport EL, Morgan GJ and Davies FE 2008. Untangling the unfolded protein response. *Cell Cycle* 7:865-69
94. Power MV and Workman P 2006. Targeting of multiple signaling pathways by heat shock protein 90 molecular chaperone inhibitors. *Endocrine-Related Cancer* 13:S125-35

95. Georgakis GV, Younes A 2005. Heat-shock protein 90 inhibitors in cancer therapy: 17AAG and beyond. *Future Med* 1:273-81
96. Bellacosa A, Kumar CC, Di Cristofano A, Testa JR 2005. Activation of Akt kinases in cancer: Implications for therapeutic targeting. *Adv Cancer Res* 94:29-86
97. Altomare DA, Huihong You, Guang-Hui Xiao, et al 2005. Human and mouse mesotheliomas exhibit elevated AKT/PKB activity, which can be targeted pharmacologically to inhibit tumor cell growth. *Oncogene* 24:6080-9
98. Cheng J, Godwin A, Bellacosa A, et al 1992. AKT2, a putative oncogene encoding a member of a subfamily of protein serine/threonine kinases, is amplified in human ovarian carcinomas. *Proc Natl Acad Sci USA* 89:9267-71
99. Dobbin ZC and Landen CN 2013. The Importance of the PI3K/AKT/MTOR Pathway in the Progression of Ovarian Cancer. *Int J Mo Sci* 14: 8213-27. Review
100. Huai-Jing Tang, Xiaohong Jin, Shaomeng Wang, et al 2006. A small molecule compound inhibits AKT pathway in ovarian cancer cell lines. *Gynecologic Oncology* 100:308-17
101. Huang J, Zhang L, Greshock J, et al 2011. Frequent genetic abnormalities of the pi3k/akt pathway in primary ovarian cancer predict patient outcome. *Genes Chromosom Cancer* 50:606-18
102. Morgan TM, Koreckij TD, Corey E 2009. Targeted Therapy for Advanced Prostate Cancer: Inhibition of the PI3K/Akt/mTOR Pathway. *Curr Cancer Drug Targets* 9(2): 237-49
103. Malik SN, Brattain M, Ghosh PM, et al 2002. Immunohistochemical demonstration of phospho-Akt in high gleason grade prostate cancer. *Clin Cancer Res* 8:1168-71
104. Virtakoivu R, Pellinen T, Juha K, et al 2012. Distinct roles of AKT isoforms in regulating β 1-integrin activity, migration, and invasion in prostate cancer. *Mol Biol Cell* 23(17):3357–69

105. Peters S, Adjei AA, Gridelli C, et al 2012. Metastatic non-small-cell lung cancer (NSCLC): ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Annals of Oncology* 23(7): vii56-vii64
106. Al-Saad S, Donnem T, Al-Shibi K, et al 2009. Diverse Prognostic Roles of Akt Isoforms, PTEN and PI3K in Tumor Epithelial Cells and Stromal Compartment in Non-small Cell Lung Cancer. *Anticancer Res* 29:4175-84
107. Jian-Min Tang, Quan-Ying He, Rui-Xia Guo, et al 2006. Phosphorylated Akt overexpression and loss of PTEN expression in non-small cell lung cancer confers poor prognosis. *Lung Cancer* 51:181-91
108. Cianfrocca M and Goldstein LJ 2004. Prognostic and Predictive Factors in Early-Stage Breast Cancer. *The Oncologist* 9(6): 606-16
109. Wickenden JA and Watson CJ 2010. Key signaling nodes in mammary gland development and cancer. Signaling downstream of PI3 kinase in mammary epithelium: a play in 3 Akts. *Breast Cancer Res* 12(2): 202
110. Soumya Krishnamurthy, Alakananda Basu 2011. Regulation of IKK ϵ expression by Akt2 isoform. *Genes Cancer* 2(11):1044-50
111. Chudnovsky Y, Adams AE, Robbins PB, et al 2005. Use of Human Tissue to Assess the Oncogenic Activity of Melanoma-Associated Mutations. *Nat Genet* 37(7):745-49
112. Mikhail M, Velazquez E, Shapiro R, et al 2005. PTEN expression in melanoma: relationship with patient survival, Bcl-2 expression, and proliferation. *Clinical Cancer Research* 11(14): 5153-7
113. Stahl JM, Sharma A, Cheung M, et al 2004. Deregulated Akt3 activity promotes development of malignant melanoma. *Cancer Research* 64(19):7002-10
114. Madhunapantula Subba Rao V, Robertson GP 2011. Therapeutic Implications of Targeting AKT Signaling in Melanoma. *Enzyme Res* 2011:327923

115. Ringel MD, Hayre N, Saito J, et al 2001. Overexpression and Overactivation of Akt in Thyroid Carcinoma. *Cancer Res* 61:6105-11.
116. Soung YH, Lee JW, Nam SW et al 2006. Mutational analysis of AKT1, AKT2 and AKT3 genes in common human carcinomas. *Oncology* 70(4):285-9
117. Roy HK, Olusola BF, Clemens DL, et al 2002. AKT proto-oncogene overexpression is an early event during sporadic colon carcinogenesis. *Carcinogenesis* 23(1):201-5
118. Francipane MG, Lagasse E 2013. mTOR pathway in colorectal cancer: an update. *Oncotarget* 5(1):49-66
119. Wen PY and Kesari S 2008. Malignant gliomas in adults. *N Engl J Med* 359:492-507
120. Hideo Mure, Kazuhito Matsuzaki, Keiko T Kitazato 2010. Akt2 and Akt3 play a pivotal role in malignant gliomas. *Neuro Oncol* 12(3): 221-32
121. Bhutani J, Sheikh A, Khan Niazi A 2013. Akt inhibitors: mechanism of action and implications for anticancer therapeutics. *Infectious Agents and Cancer* 8:49
122. Song G, Ouyang G, Bao S 2005. The activation of Akt/PKB signaling pathway and cell survival. *J Cell Mol Med* 9(1):59-71
123. Luo Y, Shoemaker AR, Liu X, et al 2005. Potent and selective inhibitors of Akt kinases slow the progress of tumors *in vivo*. *Mol Cancer Ther* 4(6):977-86
124. Rhodes N, Heerding DA, Duckett DR, et al 2008. Characterization of an Akt kinase inhibitor with potent pharmacodynamic and antitumor activity. *Cancer Res* 68(7):2366-74
125. Kozikowski AP, Dennis P, Sun H, et al 2004. WO2004022569. Akt Inhibitors, pharmaceutical compositions, and uses thereof.

126. Gills JJ, Dennis PA 2009. Perifosine: Update on a novel Akt inhibitor. *Curr Oncol Rep* 11(2):102-10
127. Hideshima T, Catley L, Yasui H, et al 2006. Perifosine, an oral bioactive novel alkylphospholipid, inhibits Akt and induces in vitro and in vivo cytotoxicity in human multiple myeloma cells. *Blood* 107(10):4053-4062
128. Ruiter GA, Zerp, SF, Bartelink H, et al 2003. Anti-cancer alkyllysophospholipids inhibit the phosphatidylinositol 3-kinase-Akt/PKB survival pathway. *Anti-Cancer Drugs* 14(2):167-173
129. Pinton G, Manente AG, Angeli G, et al 2012. Perifosine as a potential novel anti-cancer agent inhibits EGFr/MET-AKT axis in malignant pleural mesothelioma. *PLoS One* 7(5):1-7
130. Hirai H, Sootome H, Nakatsuru Y, et al 2009. An allosteric Akt inhibitor, MK-2206 enhanced anti-tumor efficacy by standard of care agents or molecular targeted drugs in vitro and in vivo. AACR Meeting Abstracts; p. 3707
131. Lu W, Defeo-Jones D, Davis LJ, et al 2004. In vitro and in vivo antitumor activities of MK-2206, a new allosteric Akt inhibitor. AACR Meeting Abstracts; p. 3714
132. Luo Y, Smith RA, Guan R, et al 2004. Pseudosubstrate peptides inhibit Akt and induce cell growth inhibition. *Biochemistry* 43:1254-63
133. Shin I, Edl J, Biswas S, Lin PC, et al 2005. Proapoptotic activity of cell-permeable anti-Akt single-chain antibodies. *Cancer Res* 65:2815-24
134. Marshall J, Posey J, Hwang J, et al 2007. A phase I trial of RX-0201 (AKT anti-sense) in patients with an advanced cancer. *J Clin Oncol* (Meeting Abstracts). 20.25(18_suppl):3564

Chapter 2

Outline of the thesis

Malignant Mesothelioma (MM) is an asbestos-related highly aggressive cancer that arises from mesothelial cells. It is characterized by a long latency period, poor prognosis and high chemo-resistance; its incidence has been predicted to increase dramatically worldwide during the next two decades.

The PI3K/AKT signaling pathway is one of the major pathways involved in carcinogenesis and tumor progression; it is aberrantly active in several human cancers and plays an important role in human malignant pleural mesothelioma (MPM).

We firstly demonstrated that MPM cell lines express only two AKT isoforms (AKT1 and AKT3) and differently from lung adenocarcinoma derived cells they lack of AKT2; moreover, the two isoforms are expressed in different percentages in cells derived from tumors with different histotype.

Based on these data, the first aim of this study was to better characterize the expression and function of the two AKT isoforms in MPM cells, and focus on their role in chemo-resistance.

Our data show that treatment of MPM cells with the pan-AKT inhibitor MK2206 resulted in a significant reduction of cell viability and enhanced sensitivity to cisplatin, with an increase in apoptotic cell death. In addition, we compared the effect of silencing each AKT isoform in MSTO211-H cells to the inhibition of their kinase activity. Both AKT1 or -3 silencing reduced cell viability and increased apoptotic cell death in response to cisplatin treatment: this suggests that both isoforms play essential roles in cell survival, but probably they act through different mechanisms.

The roles of the different AKTs in cell migration and metastases have been described. Different and sometimes conflicting studies suggest either positive or negative regulatory roles of specific AKT isoforms.

The MSTO211-H cell line is derived from a tumor with biphasic histotype, shows a spindle morphology and expresses high level of both AKT1 and -3 isoforms. In this cell model we observed that AKT1 down-regulation caused a reversion versus a more epithelial phenotype when cell were cultured in monolayer, a reduction in spreading on Matrigel and inhibition of colony growth in soft agar. Conversely, AKT3 down-regulation did not influence these processes, neither modified cell morphology.

These results led us to establish a primary role for the endogenous AKT1 protein in affecting anchorage independent cell growth and underline the isoform-specific nature of this effect.

AKT1 inhibition with MK2206 did not cause a change in cell morphology, while overexpression of AKT1 in the epithelioid REN cells caused a change in their morphology towards a more spindle-shaped phenotype. These data suggest that these effects depend on the physical presence (and not on the activation status) of AKT1 and one can hypothesize that the drug-inhibited protein still interact with some partners or assemble into macromolecular complexes.

As previously described in our laboratory, AKT in basal condition is in part phosphorylated and in part acetylated; upon SIRT1 inhibition, it becomes more acetylated and associates with different proteins, as shown by immunoprecipitation experiments.

Based on this evidence, the last aim of my study was to explore the roles of these associated proteins and among them, we focused on HSC70, a chaperone protein constitutively expressed across tissues and cell types.

Obtained data demonstrate that silencing of HSC70 leads to a strong reduction in cell viability and results in cell morphological changes,

associated with a decrease in E-Cadherin expression. We describe that AKT1 phosphorylation was affected by silencing of HSC70, while the total protein amount increased. EGF stimulus was not able to rescue AKT1 activation. Furthermore, we observed that upon HSC70 silencing also EGFR phosphorylation and expression were compromised.

Chapter 3

Targeting different AKT isoforms in Malignant Mesothelioma

¹E Borroni, ¹G Pinton, ¹AG Manente, ²SG Gray, ^{2,3}KJ O'Byrne, ⁴L Mutti, ¹L Moro.

¹Dept. of Pharmaceutical Sciences, University of Piemonte Orientale A. Avogadro, Novara, Italy; ²Institute of Molecular Medicine, St James's Hospital, Dublin 8, Ireland; ³Cancer & Ageing Research Program, Queensland University of Technology, Brisbane, Australia, ⁴Dept. of Medicine, Vercelli Hospital, Vercelli, Italy.

Lung Cancer – in submission

ABSTRACT

The PI3K/AKT signaling pathway is aberrantly active and has an important biologic impact in human malignant pleural mesothelioma (MPM) progression and chemo-resistance.

AKT family consists of three isoforms, AKT1, AKT2, and AKT3. Despite the growing amount of research demonstrating the existence of isoform-specific roles, many papers still draw generalized conclusions about AKT, without focusing on functional specificity of each isoform. Here we describe that MPM cells express increased AKT1 and AKT3 levels compared to non-malignant mesothelial cells, with the highest expression in the biphasic subtypes. We show that both AKT1 and AKT3 play essential roles in the biphasic MPM cell survival and response to cisplatin treatment. Importantly, the knockdown of AKT1, but not of AKT3, results in a dramatic change in

cell-shape when cells are grown as monolayer in culture dishes or in a Matrigel matrix and in a near complete inhibition of colonies formation in soft agar. Our studies establish a role for the endogenous AKT1 protein in affecting anchorage independent MPM cell growth and highlight the is form-specific nature of this effect. These data could be helpful for designing new effective therapeutic strategies.

INTRODUCTION

Human malignant pleural mesothelioma (MPM) is a rapidly lethal cancer associated with exposure to asbestos that is increasing in incidence worldwide [1, 2]. MPM is typically refractory to current treatment options using chemotherapy. Indirectly poly-chemotherapy has been demonstrated to increase overall survival against controls (from median of just 6 months to between just 9-12 months) [3, 4]. In a first line setting, pemetrexed in combination with cisplatin has been accepted as an almost universal standard [5]. In the second line setting, various chemotherapy agents are used, either as monotherapy or as part of polytherapy, but none has been validated and actually, no approved drugs reverse disease progression [6, 7]. Therefore, there is a major unmet need for novel therapies for MPM.

The PI3K/AKT signaling pathway, aberrantly active and with an important biologic impact in MPM progression and chemo-resistance, represents a novel good therapeutic target [8, 9]. AKT (also known as PKB) serine-threonine kinases function as critical regulators of cell survival, proliferation, metabolism, and migration. Three isoforms of AKT have been identified in mammals: AKT1, AKT2 and AKT3 [11]. Although these three isoforms are encoded by separate genes, they share a conserved N-terminal pleckstrin homology (PH) domain, a central kinase domain, and a C-terminal regulatory domain, which contains the hydrophobic motif, a characteristic of

the cAMP-dependent protein kinase A/protein kinase G/protein C (AGC) superfamily of protein kinases. N-terminal PH domain, a catalytic domain in the middle, and a C-terminus. Findings from AKT isoform-specific knockout mice suggest that the functions of the different AKT kinases are not completely overlapping and that isoform-specific signaling contributes to the diversity of AKT activities.

Thus, AKT1 knockout mice are smaller than littermate controls and show increased rates of apoptosis in some tissues, reflecting the role of AKT1 in cell survival. By contrast, AKT2 null mice develop type 2 diabetes and impaired glucose utilization, suggesting that AKT2 function is more specific for the insulin receptor signaling pathway. The precise role of AKT3 is less clear, however, mice lacking AKT3 display impaired brain development [12].

Deregulation of AKT kinases is frequently associated with human diseases such as cancer [13]. All AKT isoforms possess in vitro transformational ability [14]; however, there may be isoform-specific functions in tumor cells where amplification or mutations have been detected [15].

Despite the growing amount of research demonstrating the existence of isoform-specific regulation, many papers still draw generalized conclusions about AKT function in cancer cells without considering the unique function of each AKT isoform.

Recently, our group published that MPM derived cell lines express both AKT1 and -3 isoforms but, differently from lung adenocarcinoma cells, not AKT2 [16].

Here, we better define the relevance of AKT1 and -3 expression and activation in MPM cells.

MATERIALS AND METHODS

Reagents and antibodies

The pan-AKT and phospho-AKT1, -2, -3(pSer473) polyclonal antibodies and the monoclonal antibodies specific for PARP1 and α -tubulin were from Santa Cruz Biotechnology (Santa Cruz, CA). The monoclonal antibodies specific for AKT1 and AKT3 were from Rockland Immunochemicals Inc. (Gilbertsville, PA).

Anti mouse and anti rabbit IgG peroxidase conjugated antibodies and chemical reagents were from Sigma-Aldrich (St Louis, MO). ECL was from Amersham Pharmacia Biotech (Uppsala, Sweden). Nitrocellulose membranes and protein assay kit were from Bio-Rad (Hercules, CA). Culture media, sera, antibiotics and LipofectAMINE transfection reagent were from Invitrogen (Carlsbad, CA).

Cell cultures and transfection

The epithelioid MPM derived REN cell line was isolated, characterized and kindly provided by Dr. S.M.Albelda (University of Pennsylvania, Philadelphia; PA, USA). The MM98 cell line derived from pleural effusions of patients with epithelioid MPM and the MESO-1 biphasic cell line were isolated, characterized and stabilized in culture [17, 18]. The H2596 cell line was produced by Dr. H.I. Pass from surgical specimens derived from patients with resected sarcomatoid MPM [19]. The biphasic MSTO-211H and the mesothelial MET5A cell lines were obtained from the Istituto Scientifico Tumori (IST) Cell-bank, Genoa, Italy. Cells were cultured in standard conditions. Mycoplasma infection was excluded by the use of Mycoplasma Plus TM PCR Primer Set kit from Stratagene (La Jolla, CA). Cells grown to 80% confluence in tissue culture dishes were transiently transfected using LipofectAMINE reagent as described by the

manufacturer. Gene silencing was achieved using Akt1 or Akt3 -specific siRNAs by QIAGEN (Hilden, Germany). As a control was used QIAGEN non specific siRNA. HA-Akt expression vector was kindly provided by Dr. P. Defilippi (University of Turin, Italy).

Cell cultures in a Matrigel matrix

Briefly, 10×10^3 cells suspended in 50 μ l of cell culture medium were seeded onto solidified Matrigel (BD-Biosciences, NJ) in 24-well plates. After incubation at 37°C, cells adhered to the surface of the gel and spread to form networks. These were observed under an inverted microscope and photographed live. The experiments were done in triplicate and repeated in independent conditions.

Assay for anchorage-independent cell growth

Anchorage-independent growth was determined using a modification of previously described methods [20]. Briefly, a base layer of 0.6% agar in complete medium was plated in six-well plates and allowed to solidify. Next, wells were overlaid with 5×10^3 cells per well in a 0.3% agar. The plates were incubated at 37°C, 5% CO₂ for 15 days and checked every 2 days for colony formation. At day 7, individual colonies (defined as clusters of 15 or more cells) were counted in 10 random fields.

Proliferation assay by cell count

Cells were seeded at a density of 10×10^4 cells/well on 6-well plates in complete growth medium and incubated over-night at 37°C in a humidified environment containing 5% CO₂ to allow adherence. After described treatments, cells were trypsinized and stained with Trypan blue. The number of cells considered viable was counted in a Bürker haemocytometer within 5 minutes after staining.

Cell lysis and immunoblot

Cells were extracted with 1% NP-40 lysis buffer (1% NP-40, 150 mM NaCl, 50 mM Tris-HCl pH 8, 5 mM EDTA, 10 mM NaF, 10 mM Na₄P₂O₇, 0.4 mM Na₃VO₄, 10 µg/ml leupeptin, 4 µg/ml pepstatin and 0.1 Unit/ml aprotinin). Cell lysates were centrifuged at 13.000 x g for 10 minutes and the supernatants were collected and assayed for protein concentration with the Bio-Rad protein assay method. Proteins were separated by SDS-PAGE under reducing conditions. Following SDS-PAGE, proteins were transferred to nitrocellulose, reacted with specific antibodies and then detected with peroxidase-conjugate secondary antibodies and chemiluminescent ECL reagent. Densitometric analysis was performed using the GS 250 Molecular Image (Bio-Rad).

RNA isolation and Quantitative Real-Time PCR

Total RNA was extracted from REN cells previously transfected with the plasmid, empty vector, siRNAs or nonspecific siRNA and, using the guanidiniumthiocyanate method [21]. Starting from equal amounts of RNA, cDNA used as template for amplification in the Real Time PCR (5 µg), was synthesized by the reverse transcription reaction using RevertAidTM Minus First Strand cDNA Synthesis Kit from Fermentas-Thermo Scientific (Burlington, Ontario, CDN), using random hexamers as primers, according to the manufacturer's instructions. As a PCR internal control, 18S was simultaneously amplified using the primers: Fw 5'-AAACGGCTACCACATCCAAG-3' and Rev 5'-CCTCCAATGGATCCTCGTTA-3'. The primers for Akt1 were Fw 5'-GCTGGACGATAGCTTGGA-3' and Rev 5'-GATGACAGATAGCTGGTG-3'. The primers for Akt3 were Fw 5'-GCAAGTGGACGAGAATAAGTCTC-3' and Rev 5'-ACAATGGTGGGCTCATGACTTCC-3'. These primers were designed to generate 383 (Akt1) and 329 (Akt3) bp products, respectively.

Primers for MTA1 were Fw 5'- AGCTACGAGCAGCACAACGGG GT -3', Rev 5'-CACGCTTGGTTTCCGAGGAT-3'and for CDH1 were Fw 5'-TGGGCTGGACCGAGAGAGTT-3' and Rev 5'-ATCTCCAGCCAGTTGGCAGT-3'.

Statistical analysis

Statistical evaluation of the differential analysis was performed by one way ANOVA and Student's t-test. The threshold for statistical significance was set at $P < 0.05$.

RESULTS

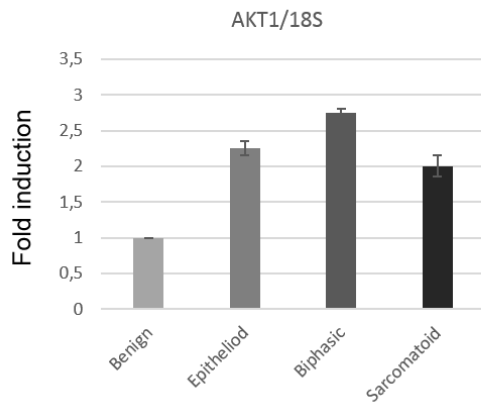
AKT isoforms are differentially expressed in MPM tissues

We recently published that MPM cells express AKT1 and -3 but not AKT2 [16]; in the present study we better investigated AKT1 and -3 expression levels in different MPM tissues and cell lines representative of the three MPM histotypes: epithelioid, biphasic and sarcomatoid. Figure 1A and B report AKT1 and -3 mRNA expression levels evaluated in clinical surgical specimens from normal mesothelia and mesotheliomas with different histotype. As shown, tumor tissues express higher levels of AKTs mRNA compared to normal mesothelium, with the biphasic tumors expressing the highest levels of both AKT1 and -3 isoforms. Data reported in Figure 1C and D show a strong correspondence between results obtained *in vivo* (Figure 1A and B) and what observed *in vitro* when the expression of both AKTs was evaluated in different MPM derived cell lines. AKT1 and -3 resulted increased in MPM derived cell lines compared to non-malignant mesothelial cells, with the highest expression of both isoforms in the MSTO-211H cell line established from a tumor with biphasic histotype.

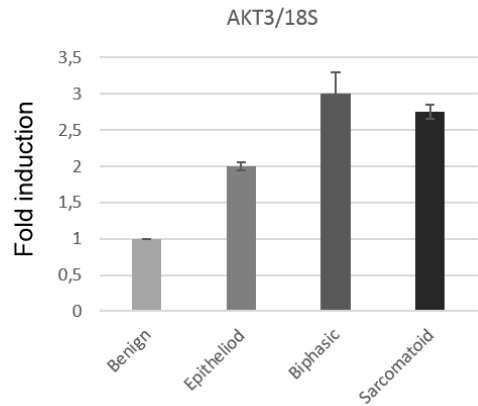
Based on these results, we decided to mainly use this cell line as a model to perform further *in vitro* experiments.

Tissue samples

A

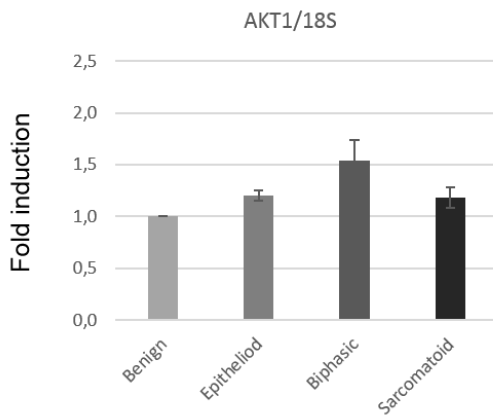


B



Cell lines

C



D

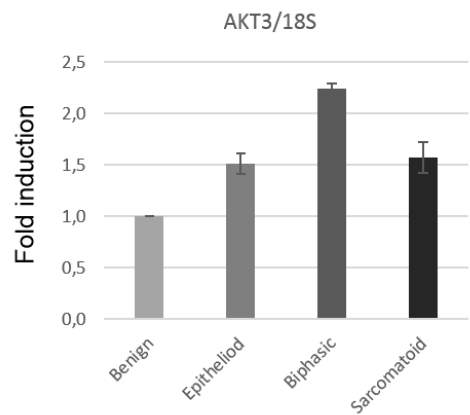
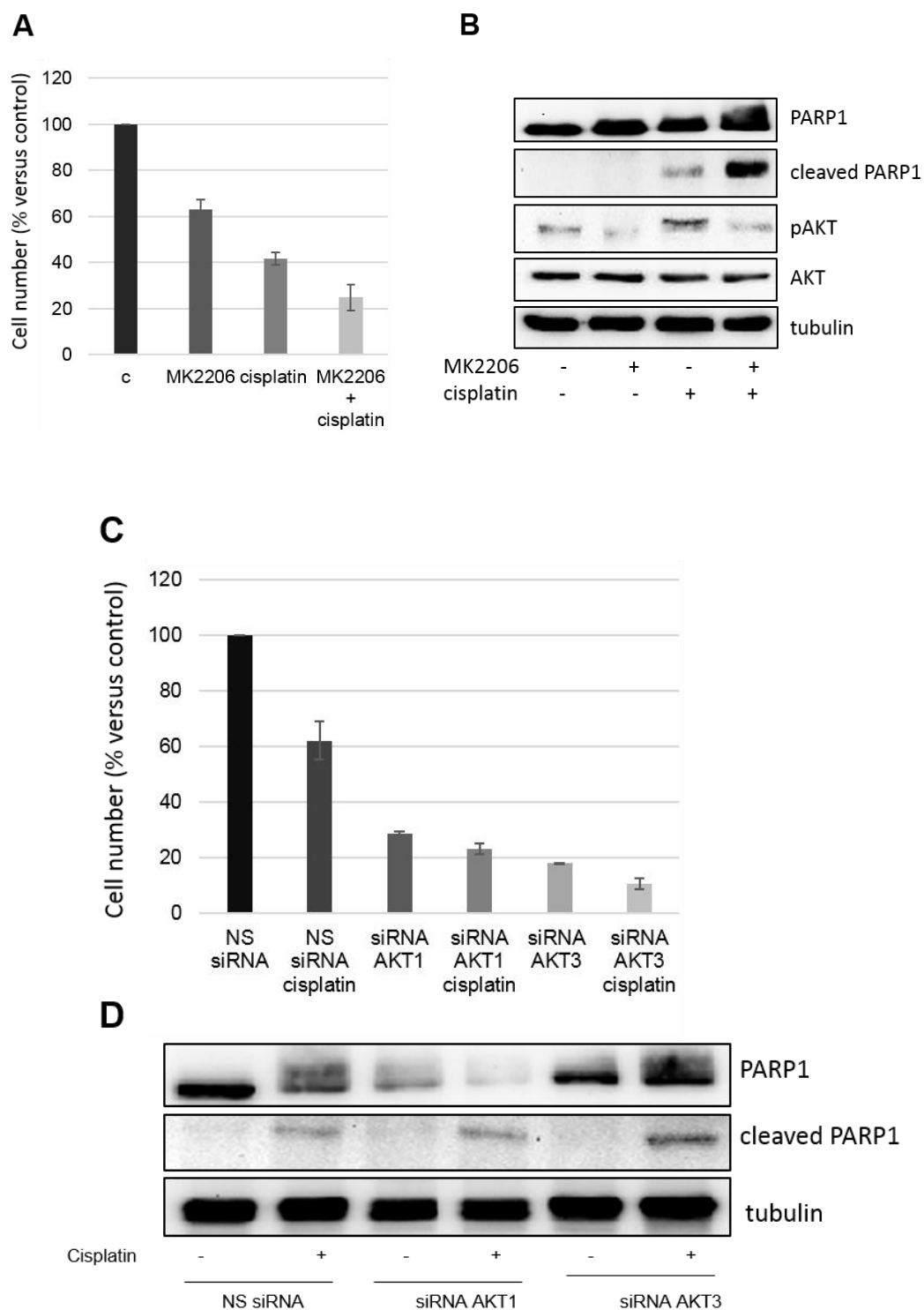


Figure 1. AKT isoforms are differentially expressed in MPM cell lines and tissues

A, B. Bar graphs representative of AKT1 and AKT3/18S mRNA ratio evaluated by RT-PCR in surgical specimens of tumors with different histotype from mesothelioma patients and in normal mesothelium. **C, D.** Bar graphs representative of AKT1 and AKT3/18S mRNA ratio in normal mesothelial cells and in different mesothelioma cell lines representative of the three histotypes. Data, shown as fold induction versus benign, are means \pm S.D. of three independent experiments.

AKTs inhibition affects MPM cell viability and enhances cisplatin induced apoptosis

Figure 2B shows that MSTO-211H cells constitutively express phosphorylated AKTs. We demonstrated that 24 hours treatment of MSTO-211H cells with MK2206, an highly selective inhibitor of AKT1, -2 and -3, markedly decreased AKTs phosphorylation (Figure 2B) and significantly compromised cell viability (Figure 2A). As it has been described a role of AKT in cisplatin resistance, we tested the effect of AKTs inhibition in response of MSTO-211H cells to this chemotherapeutic. Concomitant treatment of cells with MK2206 and cisplatin resulted in enhanced cisplatin induced apoptotic cell death as evidenced by cell count (Figure 2A) and increased PARP1 cleavage (Figure 2B). Thus, we explored the effect on cell viability and cisplatin response of specific knockdown of AKT1 and -3 isoforms by specific siRNA. Non-specific siRNAs were used as control. As shown in Figure 1C, down-regulation of both AKT1 and -3 had a significant impact on MSTO-211H cells viability in monolayer culture, although the reduction due to the loss of AKT 3 was more significant. Cisplatin treatment resulted in increased apoptotic cell death in both AKT1 and -3 siRNA transfected cells compared with the non-specific control siRNAs (Figure 1C), indicating that both activated AKTs are essential for cell survival and play a role in response to chemotherapy. The AKT1 and -3 silencing efficiency was confirmed by mRNA (Figure 2E) and protein analysis (Figure 2F).



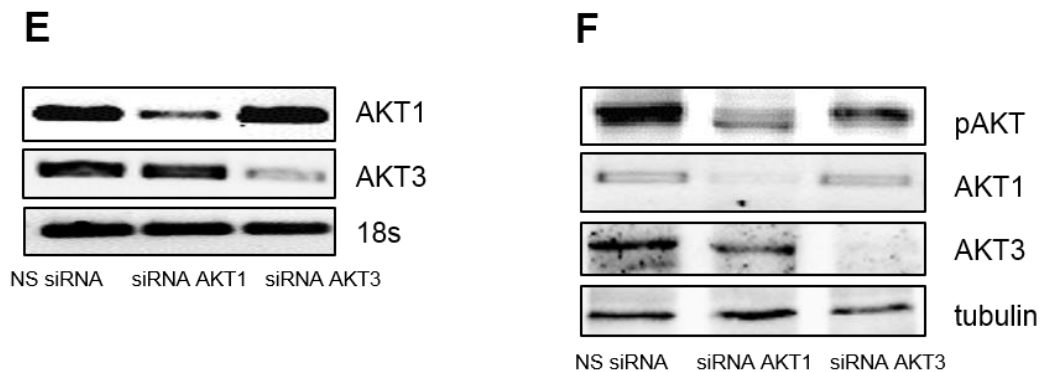


Figure 2. AKTs inhibition induces MPM cell death and enhances cisplatin induced apoptosis

A. The effect of MK2206 on the MSTO-211H cell viability was tested at 24 hours in the absence or in the presence of 25 μ M cisplatin. Points, means \pm SD of three individual measurements. **B.** Representative Western blot analysis that documents PARP1 cleavage and AKTs phosphorylation in response to 24 hours treatment with MK2206 \pm 25 μ M cisplatin. **C.** MSTO-211H cell viability was evaluated after 72 hours of transfection with specific siRNA for AKT1 and AKT3 in the absence or in the presence during the last 24 hours of 25 μ M cisplatin. Points, means \pm SD of three individual measurements. **D.** Representative Western blot analysis that documents PARP1 cleavage in cells silenced for AKT1 or -3 \pm 24 hours of 25 μ M cisplatin. **E, F.** Representative quantitative RT-PCR and Western blot analysis that confirm AKT1 and -3 silencing in MSTO-211H cells.

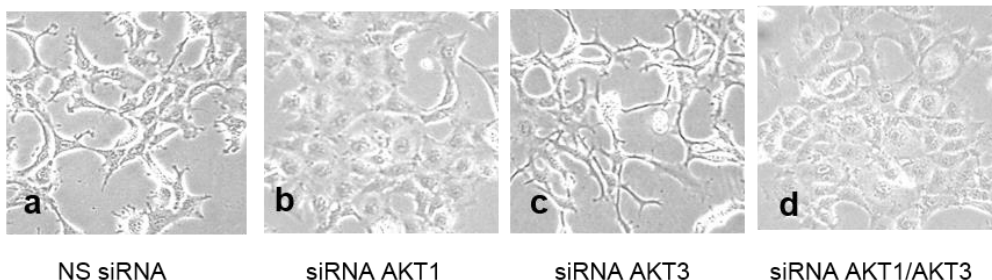
AKT1 silencing mediates MPM cell shape changes and affects clonogenic growth

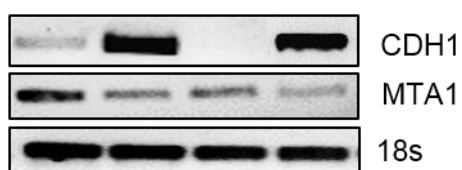
As described, down-regulation of both AKT1 and -3 had a significant impact on MSTO-211H cells viability in monolayer culture but, interestingly, only AKT1 down-regulation caused a dramatic change in MSTO-211H cell shape. Knockdown of AKT1 reverted the MSTO-211H spindle-shaped cell morphology to a more epithelial one (Figure3A-b); conversely AKT3 down-regulation exaggerated the spindle-shaped phenotype (Figure3A-c). By AKT1 and -3 double silencing experiments, we demonstrated that the epithelial phenotype, induced by AKT1 down-regulation, was maintained both in the presence and in the absence of expressed AKT3 (Figure 3A-d),

indicating that this change is due to reduced expression of the AKT1 isoform and does not depend from AKT3. Consistently with the observed change in cell morphology, an increase in CDH1 (E-Cadherin coding gene) expression was observed in AKT1 silenced cells and again this occurred independently from AKT3 expressed levels (Figure 3B). We further examined AKTs induced modifications by employing a Matrigel-overlay model. The silencing of AKT1 in MSTO-211H cells associated with reduced adhesion and cells that remained clumped together as compared to the cells treated with control siRNAs (Figure 3C-b) while cells silenced for AKT3 formed structures with more invasive protrusions (Figure 3C-d). Finally, AKT1 down-regulation significantly compromised the capability of MSTO-211H cells to form colonies in soft agar (Figure 3D-b, d) while, in contrast, AKT3 down-regulation did not result in significant variations in the number of colonies which, however, appeared larger if compared to controls (Figure 3D-c). These results support a role for AKT1 in promoting EMT (epithelial to mesenchymal transition) and invasiveness of MPM cells.

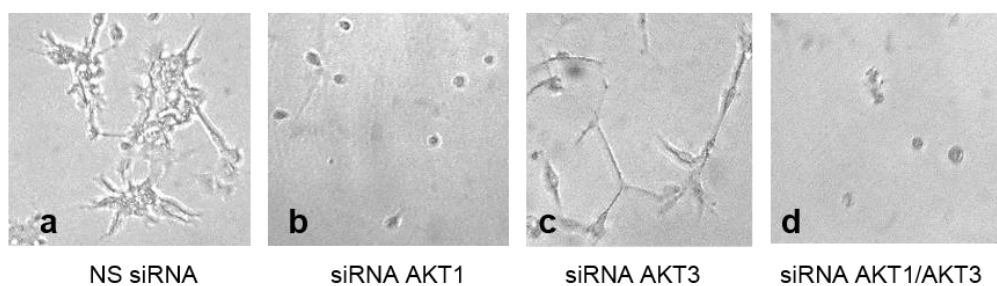
A

2D culture



B**C**

Matrigel culture

**D**

Colonies in soft agar

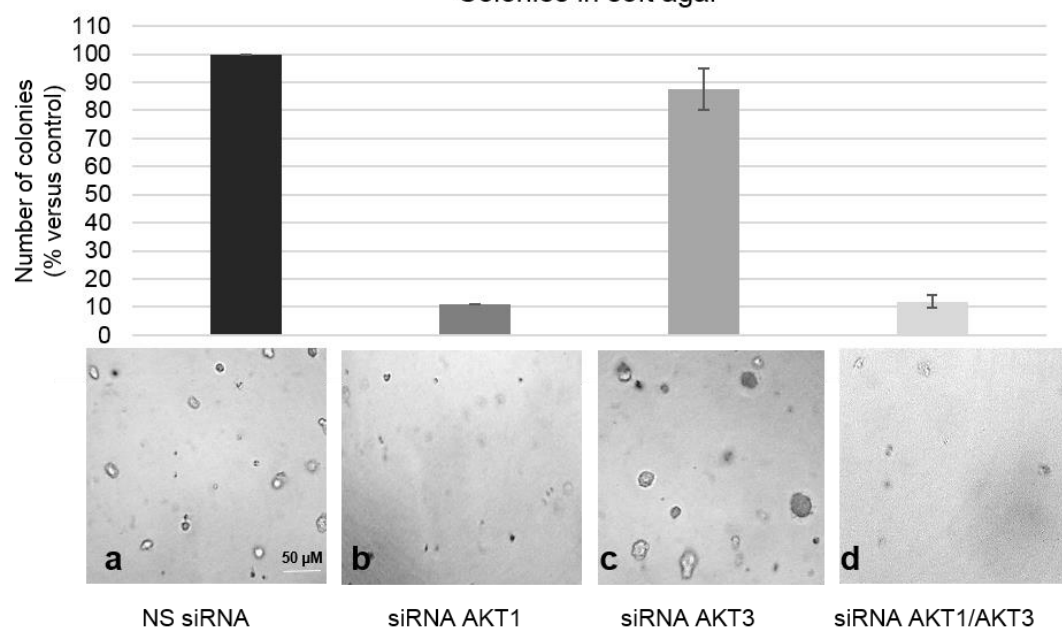


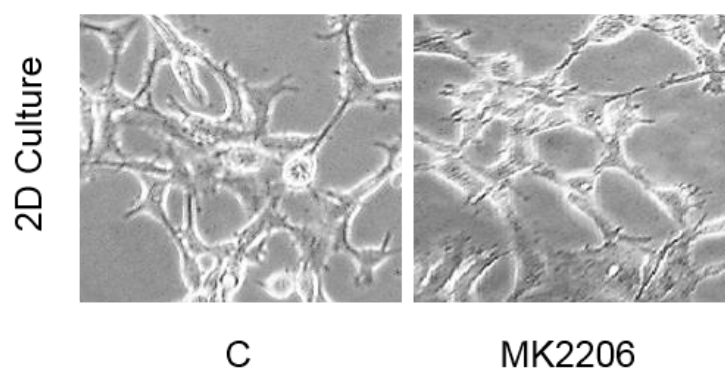
Figure 3. AKT1 silencing mediates MPM cell shape changes and affects clonogenic growth

A. Phase-contrast images of MSTO-211H cells after 72 hours of transfection with non-specific siRNA (a), AKT1 (b), AKT3 (c) or both AKT1 and -3 (d) grown as monolayer. **B.** Representative quantitative RT-PCR that documents CDH1 and MTA expression in cells transfected with non-specific siRNA, AKT1, AKT3 or both AKT1 and -3. **C.** Representative phase-contrast images of MSTO-211H cells transfected with non-specific siRNA (a), AKT1 (b), AKT3 (c) or both AKT1 and -3 (d) grown for 1 week on Matrigel. **D.** Phase-contrast images of MSTO-211H cells transfected with non-specific siRNA (a), AKT1 (b), AKT3 (c) or both AKT1 and -3 (d) grown for 1 week in soft agar. Total soft agar colony counts for silenced and non-silenced MSTO-211H cells were done in three independent experiments microscopically visualizing individual colonies (clusters of 15 or more cells) in 10 random microscopic fields. Columns represent the fold increase of the mean number of colonies in 10 fields; bars, SD; *p,0.05. Representative of three separate experiments.

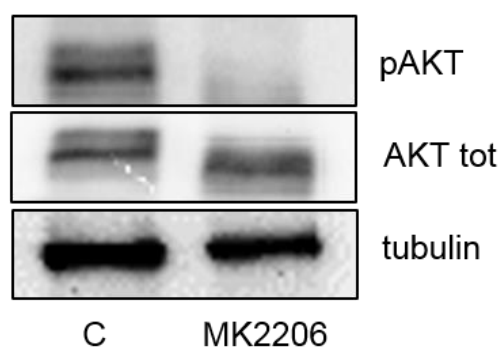
AKT1 kinase inhibition or protein loss exerts different effects in MPM cells

To discriminate if the observed phenotype transition was due to the loss of AKT1 protein or to the loss of its activity, we treated MSTO-211H cells with MK2206. As show in Figure 4A the morphology in monolayer culture did not change in response to treatment with MK2206, indicating that the observed induction of the less invasive epithelial phenotype depends on the physical presence of AKT1 rather than by its activation status. Figure 4B shows a representative Western blot analysis that confirms the inhibition of AKT phosphorylation upon MK2206 treatment. RT-PCR analysis shown Figure 4C confirms that neither CDH1 nor AKT1 expression changes in MK2206 treated cells. To reinforce our data, we over-expressed AKT1 by transient transfection in the epithelioid MPM derived REN cells. Western blot shown in Figure 4E confirms the induction in AKT1 expression and activation in transfected cells. As shown in Figure 4D transfected cells acquired a more spindle phenotype and in accordance, as shown in Figure 4F, the expression of the CDH1 gene was reduced.

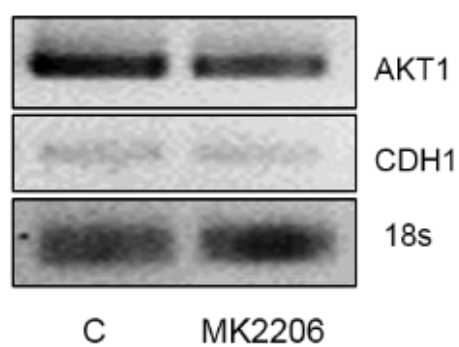
A



B



C



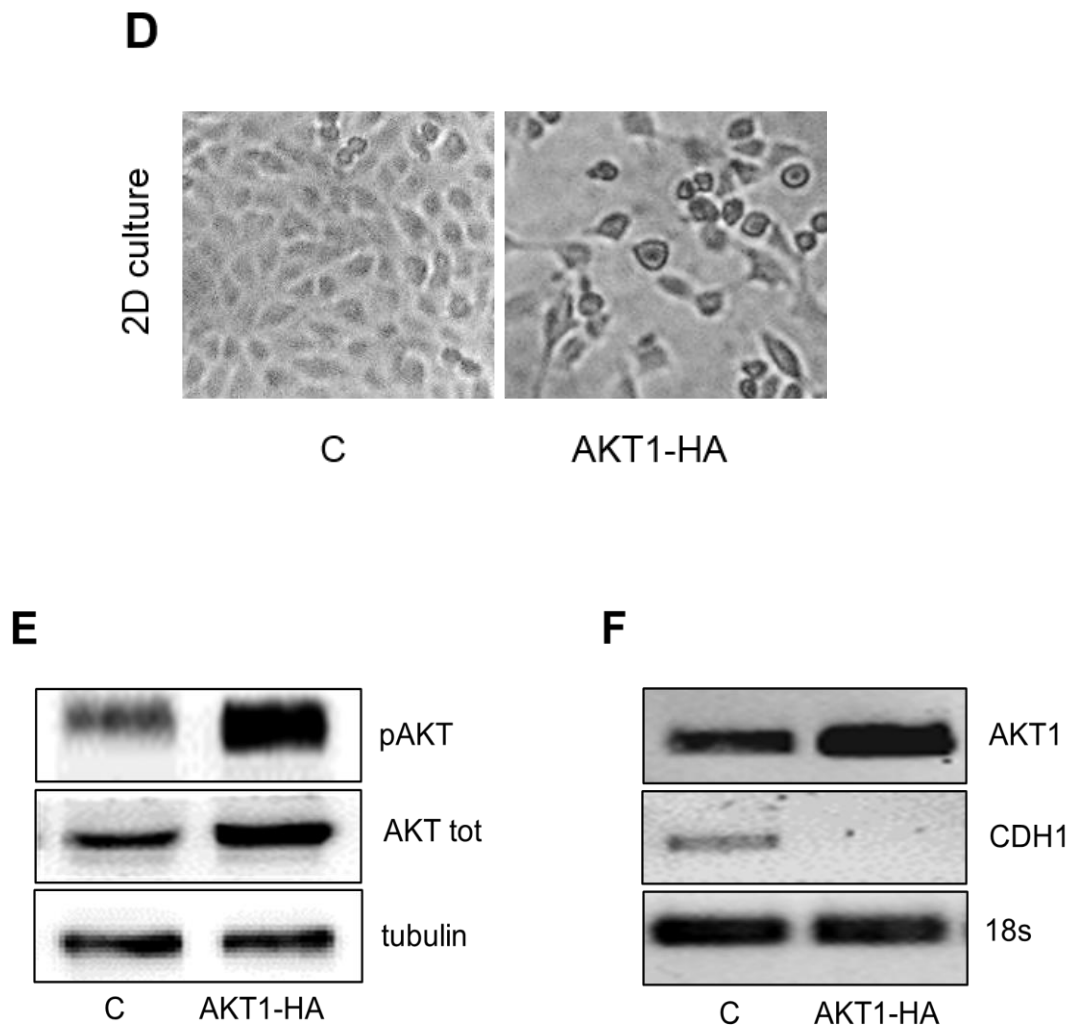


Figure 4. Different effects exerted by AKT1 kinase inhibition or protein loss in MPM cells

A. Phase-contrast images of MSTO-211H cells treated with MK2206 grown as monolayer. **B.** Representative Western blot analysis that documents AKT expression and phosphorylation in control and MK2206 treated cells. **C.** Representative quantitative RT-PCR that documents AKT1 and CDH1 expression in control and MK2206 treated cells. **D.** Phase-contrast images of REN cells transfected with an AKT1-HA expression vector grown as monolayer. **E.** Representative Western blot analysis that documents AKT expression and phosphorylation in control and AKT1 transfected cells, tubulin was used as loading control. **F.** Representative quantitative RT-PCR that documents AKT1 and CDH1 expression compared to 18S, in control and AKT1 transfected cells.

DISCUSSION

AKTs are activated in human cancer and play a critical role in tumor pathogenesis, via effects on metabolism, survival, and proliferation [22]. The role of AKTs in cell migration and metastases is less clear because of conflicting studies suggesting either positive or negative regulatory roles. In studies in which roles for AKT1 and AKT2 in cell motility have been reported, distinct and, in some cases, opposing functions for the two isoforms are observed. Differences seem to depend mainly on the cell type being studied. For example, in fibroblasts and endothelial cell, AKT1 has been found to promote invasion while AKT2 attenuates it [22, 23]. In PC-3 cells, AKT2 ablation stimulates and AKT1 ablation inhibits cell migration. Conversely, previous studies have found that AKT2 stimulates the motility of breast and ovarian cancer cells, AKT1 actually inhibits the motility of these cells [25]. In the present study, we show that AKT1 and -3 are expressed and active in MPM cells. Treatment of MPM cells with the pan-AKTs inhibitor MK2206 alone caused a significant reduction in cell viability. As a role of AKT inhibition in sensitizing cells to cisplatin treatment has been described, we tested the combined treatment with MK2206 and cisplatin in MSTO-211H cells. We observed that the concomitant treatment with MK2206 resulted in enhanced cisplatin induced apoptotic cell death. Therefore, we investigated the isoform-specific functions of AKT1 and -3 in MSTO-211H cells by the use of specific siRNA. Knockdown of AKT1 or-3 reduced cell viability and potentiated apoptotic cell death when cisplatin treatment was added. Further investigations are needed to elucidate the mechanisms by which the AKT isoforms promote cell survival in MSTO-211H cells, but it seems that even though both AKT1 and -3 play essential roles in this cell type survival, they likely act through different mechanisms.

Importantly, we observed that AKT1 down-regulation caused in these cells a reversion versus a more epithelial phenotype when cultured in monolayer, a reduction in spreading on Matrigel and a near complete inhibition of the growth in 3D cultures in soft agar. In contrast, AKT3 down-regulation didn't influence these processes. Our studies highlight the isoform-specific nature of these effects and establish a role for endogenous AKT1 protein in affecting cell invasion.

The specific mechanisms responsible for the distinct roles of AKT1 and -3 are not known; however, there are few explanations extrapolated from previously published studies. Differential subcellular localization or binding partners may determine isoform-specific functions. The balance between AKT isoforms activation downstream of growth factor receptors may influence the invasive or metastatic potential of tumors or tumor cell lines. The consequences of isoform-specific inhibition need to be carefully evaluated in different cellular contexts.

Efforts are underway to develop pan- or isoform-specific AKT inhibitors as cancer therapeutics [26-28]. By the use of a specific inhibitor of AKTs activation, we demonstrated that it is the physical presence and not the activation status of AKT1 that influence all of the effects observed in culture. This could be in part explained by the fact that the drug-inhibited protein may lack a certain activity but may still interact with some binding partners or assemble into macromolecular complexes.

In conclusion, we have demonstrated that AKT1 and -3 comparably contribute to cell survival but have different impact on cell invasion and consideration of this may aid in the development of targeted strategies for specific AKT isoforms inhibition in MPM therapy.

Acknowledgements

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REFERENCES

1. Prazakova S, Thomas PS, Sandrini A, Yates DH. Asbestos and the lung in the 21st century: an update. *Clin Respir J* 2014; 8(1):1-10.
2. Stayner L, Welch LS, Lemen R. The worldwide pandemic of asbestos-related diseases. *Annu Rev Public Health* 2013;34:205-16.
3. Scherpereel A, Astoul P, Baas P, Berghmans T, Clayson H, de Vuyst P, et al. Guidelines of the European Respiratory Society and the European Society of Thoracic Surgeons for the management of malignant pleural mesothelioma. European Respiratory Society/European Society of Thoracic Surgeons Task Force. *Eur Respir J* 2010; 35:479-95.
4. Mossman BT, Shukla A, Heintz NH, Verschraegen CF, Thomas A, Hassan R. New insights into understanding the mechanisms, pathogenesis, and management of malignant mesotheliomas. *Am J Pathol* 2013; 182(4):1065-77.
5. Vogelzang NJ, Rusthoven JJ, Symanowski J, Denham C, Kaukel E, Ruffie P, et al. Phase III study of pemetrexed in combination with m versus cisplatin alone in patients with malignant pleural mesothelioma. *J Clin Oncol* 2003; 21:2636-44.
6. Haas AR, Stermann DH. Malignant pleural mesothelioma: update on treatment options with a focus on novel therapies. *Clin Chest Med* 2013; 34(1):99-111.
7. Pinton G, Manente AG, Taviani D, Moro L, Mutti L. Therapies currently in Phase II trials for malignant pleural mesothelioma. *Expert Opin Investig Drugs* 2013; 22(10):1255-63.
8. Mikami I, Zhang F, Hirata T, Okamoto J, Koizumi K, Shimizu K, Jablons D, He B. Inhibition of activated phosphatidylinositol 3-kinase/AKT pathway in malignant pleural mesothelioma leads to G1 cell cycle arrest. *Oncol Rep* 2010; 24(6):1677-81.
9. Carbone M, Yang H. Molecular pathways: targeting mechanisms of asbestos and erionite carcinogenesis in mesothelioma. *Clin Cancer Res* 2012; 18(3):598-604.

10. Altomare DA, You H, Xiao GH, Ramos-Nino ME, Skele KL, De Rienzo A, et al. Human and mouse mesotheliomas exhibit elevated AKT/PKB activity, which can be targeted pharmacologically to inhibit tumor cell growth. *Oncogene* 2005; 24(40):6080-9.
11. Woodgett JR. Recent advances in the protein kinase B signaling pathway. *Curr Opin Cell Biol* 2005; 17:150-7.
12. Dummler B, Tschopp O, Hynx D, Yang ZZ, Dirnhofer S, Hemmings BA. Life with a single isoform of Akt: mice lacking Akt2 and Akt3 are viable but display impaired glucose homeostasis and growth deficiencies. *Mol Cell Biol* 2006; 26(21):8042-51.
13. Cheung M, Testa JR. Diverse mechanisms of AKT pathway activation in human malignancy. *Curr Cancer Drug Targets* 2013; 13(3):234-44. Review.
14. Altomare DA, Testa JR. Perturbation of the AKT signaling pathway in human cancer. *Oncogene* 2005; 24:7455-64.
15. Bellacosa A, Kumar CC, Di Cristofano A, Testa JR. Activation of AKT kinases in cancer: implication for therapeutic targeting. *Adv Cancer Res* 2005; 94:29-86.
16. Pinton G, Manente AG, Angeli G, Mutti L, Moro L. Perifosine as a potential novel anti-cancer agent inhibits EGFR/MET-AKT axis in malignant pleural mesothelioma. *PLoS One* 2012; 7:e36856.
17. Orecchia S, Schillaci F, Salvio M, Libener R, Betta PG. Aberrant E-Cadherin and γ -catenin expression in malignant mesothelioma and its diagnostic and biological relevance. *Lung Cancer* 2004; 45:S37–S43.
18. Reale FR, Griffin TW, Compton JM, Graham S, Townes PL, Bogden A. Characterization of a human malignant mesothelioma cell line (H-MESO-1): a biphasic solid and ascetic tumor model. *Cancer Res* 1987; 47(12):3199-205.
19. Pass HI, Stevens EJ, Oie H, Tsokos MG, Abati AD, Fetsch PA, et al. Characteristics of nine newly derived mesothelioma cell lines. *Ann Thorac Surg* 1995; 59(4):835-44.

20. Lewis JD, Payton LA, Whitford JG, Byrne JA, Smith DI, et al. (2007) Induction of Tumorigenesis and Metastasis by the Murine Orthologue of Tumor Protein. *Mol Cancer Res* 5: 133-44.
21. Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidiniumthiocyanate-phenol-chloroform extraction. *Anal Biochem* 1987; 162:156-9.
22. Manning BD and Cantley LC. AKT/PKB signaling: navigation downstream. *Cell* 2007; 129:1261-74.
23. Zhou GL, Tucker DF, Bae SS, Bhatheja K, Birnbaum MJ, Field J. Opposing roles for Akt1 and Akt2 in Rac/Pak signaling and cell migration. *J Biol Chem* 2006; 281:36443-53.
24. Irie HY, Pearline RV, Grueneberg D, Hsia M, Ravichandran P, Kothari N, et al. Distinct roles of Akt1 and Akt2 in regulating cell migration and epithelial-mesenchymal transition. *J Cell Biol* 2005; 171:1023-34.
25. Arboleda MJ, Lyons JF, Kabbinar FF et al. Overexpression of AKT2 / protein kinase B beta leads to up-regulation of beta1 integrins, increased invasion, and metastasis of human breast and ovarian cancer cells. *Cancer Res* 2003; 63:196-206.
26. Kumar CC and Madison V. AKT crystal structure and AKT-specific inhibitors. *Oncogene* 2005; 24:7493-501.
27. Yap TA, Yan L, Patnaik A, Fearen I, Olmos D, Papadopoulos K, Baird RD, Delgado L, Taylor A, Lupinacci L, Riisnaes R, Pope LL, Heaton SP, Thomas G, Garrett MD, Sullivan DM, de Bono JS, Tolcher AW. First-in-man clinical trial of the oral pan-AKT inhibitor MK-2206 in patients with advanced solid tumors. *J Clin Oncol* 2011; 29(35):4688-95.
28. Molife LR, Yan L, Vitfell-Rasmussen J, Zernhelt AM, Sullivan DM, Cassier PA, Chen E, Biondo A, Tetteh E, Siu LL, Patnaik A, Papadopoulos KP, de Bono JS, Tolcher AW, Minton S. Phase 1 trial of the oral AKT inhibitor MK-2206 plus carboplatin/paclitaxel, docetaxel, or erlotinib in patients with advanced solid tumors. *J Hematol Oncol* 2014; 7(1):1.

Chapter 4

HSC70 (Heat Shock Cognate Protein-70) and AKT interaction

INTRODUCTION

Chaperone proteins, acting in different cell compartments, play an important role in protein homeostasis, and are involved in different cellular processes.

The majority of the HSP70 proteins are mainly located in the cytosol and in the nucleus (HSPA1A/B, HSPA1L, HSPA2, HSPA6 and HSPA8), while two members remain restricted to mitochondria (HSPA9) or endoplasmic reticulum (HSPA5). HSP70 family members present high sequence identity (around 80% of their sequence) and similar structure (1).

Among HSP70 chaperones, one of the most abundantly and ubiquitously expressed across all tissues protein is HSPA8/HSC70. HSPA8 (also known as HSC70, HSC71, HSP71, or HSP73) is known as the constitutive member of the cytosolic HSP70 family and plays a central role in many cellular processes; it presents sequence identity of 85% to HSPA1A and represents up to 1% of the total cellular protein content in transformed cells (2). The main processes that have been related to HSPA8 (often in association with co-chaperones), are represented by clathrin-mediated endocytosis, protein folding and the ubiquitin-proteasome mediated degradation.

During the first process, binding of a ligand to a transmembrane receptor induces the formation of a cavity. Clathrin binds the nascent pit in order to induce the formation of regular endosomes; then clathrin is dissociated from endosomes through an HSPA8-dependent mechanism.

Moreover, an *in vivo* study on protein folding revealed that HSPA8 binds a wide variety of polypeptides of size larger than 20 kDa, representing 20% of

newly translated proteins: also, protein folding seems to occur in a protected environment formed by chaperones and co-chaperones (3). HSPA8 is composed of 2 domains: the N-terminal Nucleotide Binding Domain (NBD) and the C-terminal Substrate Binding Domain (SBD). The NBD is composed of 2 lobes I and II, divided in 2 subdomains Ia-Ib and II-Ib; the nucleotide-binding pocket is also composed of two subdomains, a 15-kDa β -sandwich binding the substrate and a 10-kDa R-helix that closes the binding site and increases the affinity for the peptide (4, 5). HSPA8 binds to small hydrophobic stretches of nascent or partially unfolded proteins in an ADP/ATP-dependent manner. HSPA8 firstly binds client proteins in a low affinity; after this binding and with the help of accessory proteins from the HSP40 family, it hydrolyzes ATP into ADP, adopting a high affinity; through the action of so-called nucleotide exchange factors (NEFs), it reverts to its ATP-state thus releasing its substrate peptide. The cycling between these two states (fast-exchange to low-exchange ATP) enables HSPA8 to exert its chaperone activities that rely on conformational changes of the protein upon nucleotide and clients binding.

HSPA8 exerts a key role in ubiquitin mediated protein degradation: the ubiquitin-proteasome degradation system consists in adding multi-ubiquitin chains to a protein substrate, as a signal for targeting and degradation by the proteasome proteolytic complex: in this system, the C-terminus of the HSPA8-interacting protein (CHIP) bridges HSPA8 to the proteasome by interacting with the chaperone and acts as an E3 ubiquitin ligase, coupling ubiquitin chains to the chaperone substrate (6).

Furthermore, the involvement of HSPA8 in protein import into cellular compartments has been broadly studied. The chaperone is able to shuttle between the nucleus and the cytoplasm in an ATP-dependent manner and this property allows HSPA8 to import different cytoplasmic proteins into the nucleus, while other studies correlate the role of HSPA8 in cellular transport

of organelles along microtubules. As shown by Nirdé et al., HSPA8 is also detected in the extracellular space: it may derive from dying cells, but it can also be actively released from viable cells, free or associated with exosomes; in this case, extracellular HSPA8 may inhibit tumor cell growth and affect cell viability (7).

In literature, it has been described the role of SIRT1 (Sirtuin1), a NAD dependent deacetylases, in the modulation of AKT activation and a role of PARP1 ((Poly(ADP ribose)polymerase 1) as a gatekeeper for SIRT1 activity by limiting NAD⁺ availability (8).

SIRT1 inhibition or silencing results in a more evident AKT acetylation and reduced phosphorylation (9). Previously our group has demonstrated that in basal conditions AKT was in part acetylated and in part phosphorylated, while on the contrary, upon PARP1 inhibition, AKT becomes highly phosphorylated and completely de-acetylated. Moreover, AKT acetylation, upon SIRT1 inhibition or silencing, results in the association of different acetylated proteins to AKT. We identified these interactors by MALDI-TOF analysis. In this work, among all the interactors, we decided to investigate HSC70. We better defined its relationship with AKT and explored its involvement in AKT activation and in EGF-signaling in MPM cells.

MATERIALS AND METHODS

Reagents and antibodies

EGF (Sigma-Aldrich) was dissolved, stored at -20° and used at final concentration of 10 ng/ml; EX527 (Selleckchem, Houston, TX) was dissolved in DMSO, stored at -20° and used at final concentration of 10 ug/ml. Nitrocellulose membranes and protein assay kits were from Bio-Rad (Hercules, CA). The EGFR, HSC70, AKT and phospho-AKT antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA). The

phospho-ERK antibody was purchased from Cell Signaling Technology (Beverly, MA). Anti-mouse and anti-rabbit IgG peroxidase conjugated antibodies were from Sigma-Aldrich (St Louis, MO).

Cell Culture, treatments and transfection

The MPM derived REN cell line, used as the principal experimental model in the current study, was kindly provided by Dr. S.M. Albelda (University of Pennsylvania, Philadelphia, PA); cells were cultured in RPMI 1640 medium supplemented with 10% foetal bovine serum (FBS), 100 units/ml penicillin, 100 units/ml streptomycin, and 2mM glutamine, at 37°C in a 5% CO₂-humidified atmosphere. For HSPA8 silencing in REN cells, we used specific siRNA by QIAGEN (Hilden, Germany) and Lipofectamine® transfection reagent (Invitrogen, as described by the manufacturer).

Cell proliferation assay

For cell proliferation assays, cells were seeded at a density of 10×10^4 in 51 mm petri dishes in complete growth medium and incubated overnight at 37°C in a humidified environment containing 5% CO₂. After 24 hours, cells were transfected with HSC70 specific or non-specific siRNA. Human recombinant EGF was added to medium supplemented with 2% of FBS after 4h of transfection and cells were allowed to grow for 24, 48 and 72 hours, ; at the end of treatment, viable cells were counted in a Burkert haemocytometer.

Cell lysis, immunoprecipitation and Western Blot

REN cells were seeded at a density of 10×10^4 cells in 51 mm petri dishes in complete growth medium and incubated overnight at 37°C in a humidified environment containing 5% CO₂. After 24 hours, cells were transfected with HSC70specific siRNA. After 72 hours of transfection, cells treated with EGF

(10ng/ml) for 5, 10 and 15 minutes. Cells were then lysed and proteins were analyzed by Western Blot.

Cells were lysed in NP40-lysis buffer [50mM Tris-HCl (pH 8.0), 150mM NaCl, 1% NP40, 5mM EDTA, 10 mM NaF, 10mM $\text{Na}_4\text{P}_2\text{O}_7$, 0.4 mM Na_3VO_4 , 10 $\mu\text{g/ml}$ leupeptin, 4 $\mu\text{g/ml}$ pepstatin and 0.1 Unit/ml aprotinin]. Lysates were centrifuged at 13.000 x g for 10 minutes and the supernatants were collected and assayed for protein concentration using the Bradford protein assay reagent (Bio-Rad). Proteins were separated by gel electrophoresis on 8% polyacrylamide gels, transferred to nitrocellulose membranes and detected by immunoblotting using an enhanced chemiluminescence system (Amersham Pharmacia Biotech, Uppsala, Sweden). For immunoprecipitation experiments, 2 mg of proteins were incubated with the antibody for 1 hour at 4° C in the presence of 40 μl protein A-Sepharose beads (50%) per 1 ml. The beads were washed three times with 1 ml of PBS, 0.5% Triton X-100 and once with 1 ml of PBS, 0.5% Triton X-100, 0.1% SDS and the immunoprecipitates were eluted by boiling the beads in 2X Laemmli sample buffer for 5 minutes and loaded on SDS-PAGE. For interactors identification, gel was stained with blue Coomassie, the evidenced bands were cut and proteins analyzed through MALDI-TOF analysis.

RNA isolation and RT-PCR

Total RNA was extracted from REN cells transfected with HSC70 or nonspecific siRNAs with the Trizol Reagent (Life Technologies, Inc.), according to the manufacturer's protocol. To obtain cDNA, reverse-transcription reaction was performed starting from 5 mg of RNA extracted, using RevertAidTM Minus First Strand cDNA Synthesis Kit from Thermo Scientific (Burlington, Ontario), as manufacturer's instructions. Primers were synthesized by MWG. The primers for 18s rRNA(housekeeping gene)

were: 5'-AAACGGCTACCACATCCAAG-3' (sense) and 5'-CCTCCAATGGATCCTCGTTA-3' (antisense) and for CDH1 were: 5'-TGGGCTGGACCGAGAGAGTT-3' (sense) and 5'-ATCTCCAGCCAGTTGGCAGT-3' (antisense). RT-PCR reactions were performed using 2xPCR Fusion Flash Master Mix from Thermo Scientific with the following programs: (18s) denaturation at 94°C for 3 minutes, followed by 30 cycles at 94°C for 30 seconds, 57°C for 5 seconds and 72°C for 3 seconds, with a final extension at 72°C for 1 minute; (CDH1) denaturation at 94°C for 3 minutes, followed by 35 cycles at 94°C for 30 seconds, 55°C for 5 seconds and 72°C for 30 seconds, with a final extension at 72°C for 1 minute. The PCR products expected (respectively, 155 Bp and 612 Bp) were electrophoresed on a 1% agarose gel and visualized with ethidium bromide.

RESULTS

Sirt1 inhibition induces AKT acetylation and association with different proteins

The role of SIRT1 in the modulation of AKT activation has been clearly described. SIRT1 activates AKT by enhancing its binding to PIP₃ and thereby increasing its membrane localization and phosphorylation; moreover, two conserved lysine residues in the PH domain of AKT are subject to acetylation and their SIRT1-dependent deacetylation is necessary to allow AKT binding to PIP₃ (10). Recent data published by our group demonstrate the role of SIRT1 in the control of the balancing between acetylation and phosphorylation of AKT upon PARP1 inhibition, in MPM cells: in basal conditions AKT is in part acetylated and in part phosphorylated, while inhibition (or silencing) of SIRT1 resulted in a more evident AKT acetylation (Fig. 1A) (9). REN cells were treated with the

SIRT1 inhibitor EX527 at the concentration of 10 $\mu\text{g/ml}$ for 24 hours, AKT was immunoprecipitated and samples were loaded on SDS-PAGE and subsequently stained with blue Comassie. Bands representing different AKT interactors were excised from the gel and then identified through MALDI-TOF analysis. Among these proteins we focused further studies on the Heat shock cognate protein 70 (HSC70) (Fig. 1B).

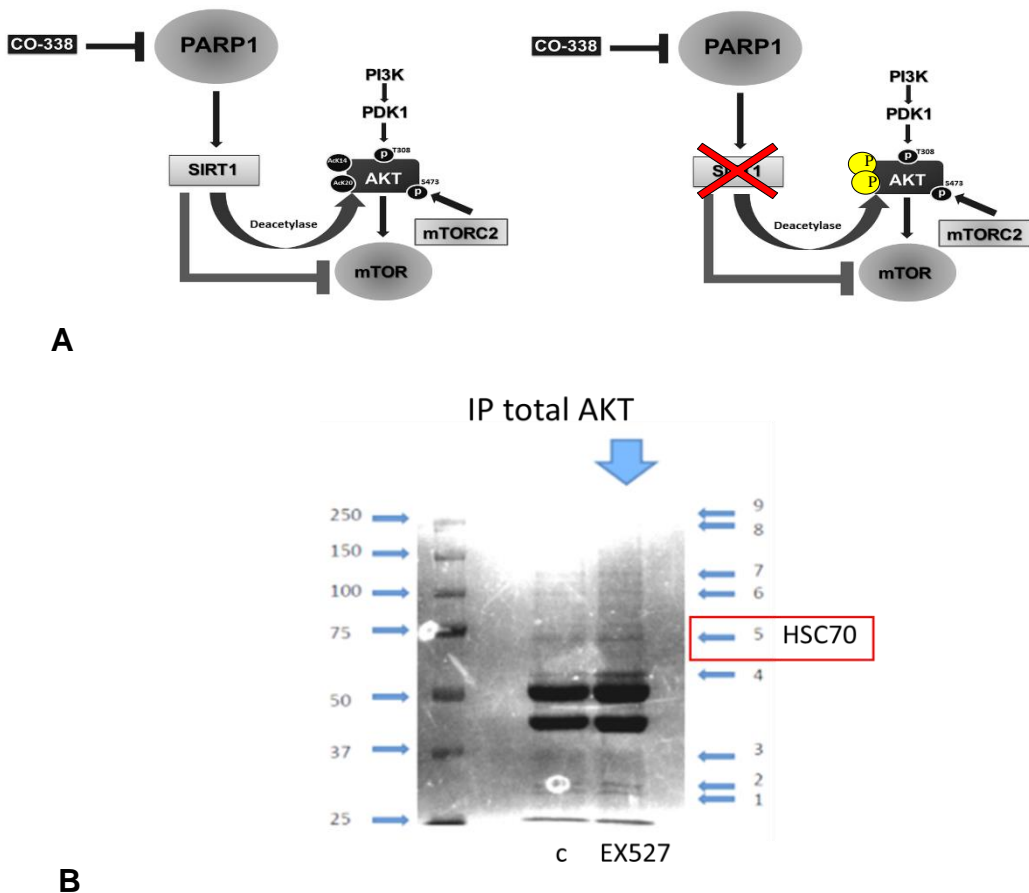
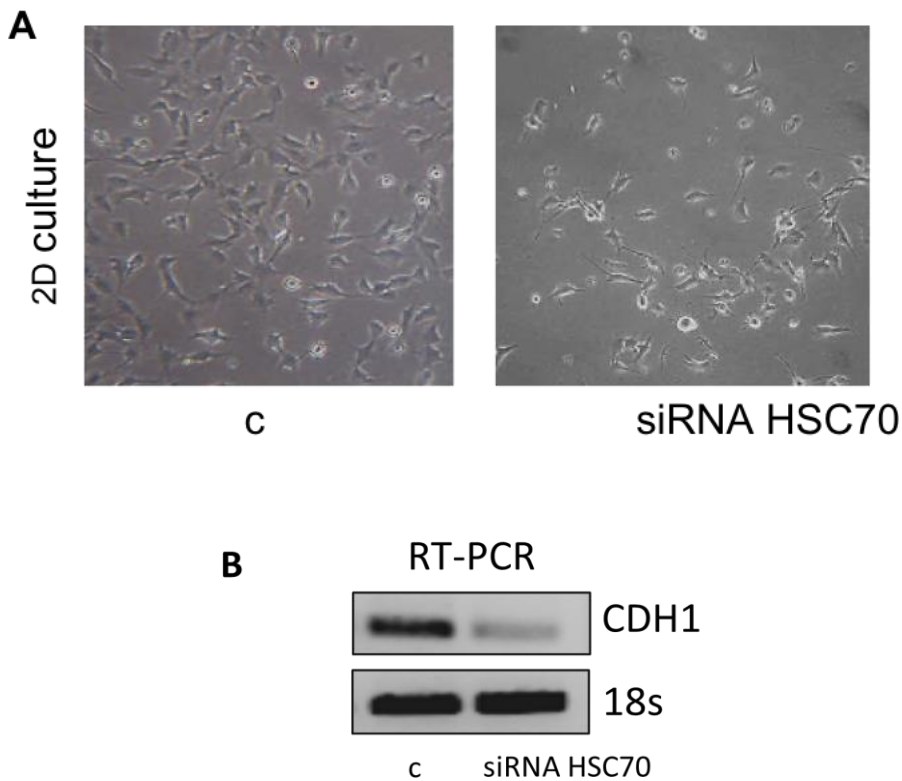


Fig. 1 - Upon SIRT1 inhibition AKT is more acetylated and associates other acetylated proteins.

A. A brief scheme of balancing between acetylation and phosphorylation of AKT (Pinton et al, 2013). **B.** REN cells were treated with 10uM EX527 for 24h. Immunoprecipitation of total AKT was performed and different bands corresponding to interactors were highlighted with Comassie staining of SDS-PAGE and finally identified through MALDI-TOF analysis.

HSC70 knock-down resulted in morphological changes and decreased cell viability

HSC70 is considered an essential housekeeping gene, and HSC70-knockout mice are embryonically lethal as a result of the pivotal role of HSC70 in cell survival. In literature it has been reported that transfection with HSC70 siRNA results in massive cell death. As a consequence of HSC70 inhibition in REN cells, we observed a significant change in cell morphology from an epithelial, to a spindle shape (Fig. 2A). Morphological changes were associated with a decrease in E-Cadherin expression as documented by the RT-PCR expression analysis of CDH1 gene, shown in (Fig. 2B). Moreover, according to previous studies, we observed an important decrease in cell number upon HSC70 silencing (Fig. 2C).



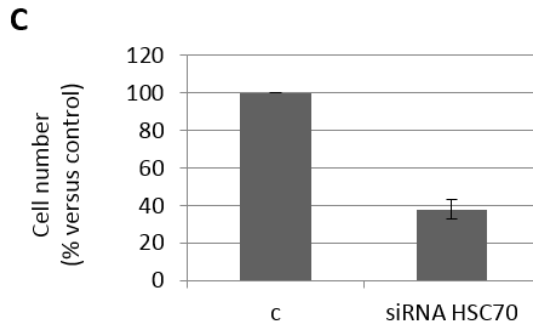


Fig. 2 - Morphological changes and viability of mesothelioma cells after HSC70 silencing. **A.** Phase-contrast images of REN cells after 72h from transfection with non-specific (c) or HSC70 siRNA. Silenced cells result reduced in number and display a more spindle phenotype. **B.** RT-PCR documents the expression of CDH1 in control and HSC70 silenced cells. 18S rRNA was used as housekeeping gene. **C.** Bar graphs documents cells viability reduction in HSC70 silenced cells. Bars represent means \pm SD of three independent measurements.

HSC70 knock-down affects AKT activation in REN Cells

We examined the effects of HSC70 silencing on basal AKT phosphorylation. Western Blot analysis, shown in Fig. 3A, documents that AKT phosphorylation was abolished in HSC70-silenced cells; while the total amount of AKT resulted increased, and in particular, AKT1 is the main isoform modulated in Hsc70 silenced cells (Fig. 4).

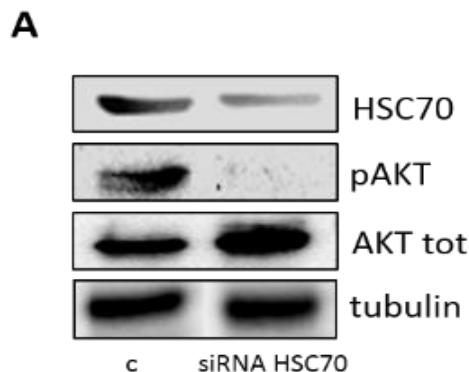


Fig. 3 – HSC70 silencing reduces AKT phosphorylation.

A. Representative Western blot analysis that documents the inhibition of AKT phosphorylation and the total amount of AKT protein in wild type and HSC70 silenced cells.

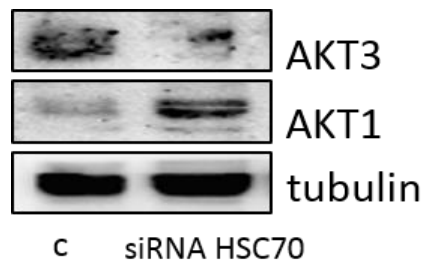
A

Fig. 4 – Loss of HSC70 resulted in AKT1 accumulation.

A. Representative Western blot analysis of AKT1 and AKT3 isoform expression in wild type and HSC70 silenced cells.

HSC70 silencing reduces EGFR phosphorylation and expression and affects EGF-induced proliferation in REN cells

REN cells express high levels of EGFR, so we explored the effects of HSC70 silencing on its activation and signaling. After 72 hours of transfection, cells were starved 1 hour and treated with 10ng/ml EGF for 5, 10 and 15 minutes. Western blot analysis of lysates from REN cells revealed that the level of phosphorylation reached a maximum after 10 minutes of treatment; however, this was significantly inhibited in cells transfected with siRNA–HSC70 (data not shown). In particular, at 10 minutes of EGFR treatment, we observed that EGFR phosphorylation diminished such as the total expression of the receptor, suggesting a direct involvement of HSC70 in EGFR signaling pathway and recycle (Fig. 5A). AKT phosphorylation was compromised even in the presence of EGF, while ERK 1/2 phosphorylation was not affected (Fig. 5A). After assessing the relationship between HSC70 and EGFR activation, we explored if HSC70 could play a role in EGF-dependent cell growth. Transfected cells were treated with EGF (10ng/ml) in medium supplemented with 2% FBS for 72hours. As previously described, HSC70 silencing significantly

compromised REN cells viability both in basal and EGF stimulated conditions (Fig.5B).

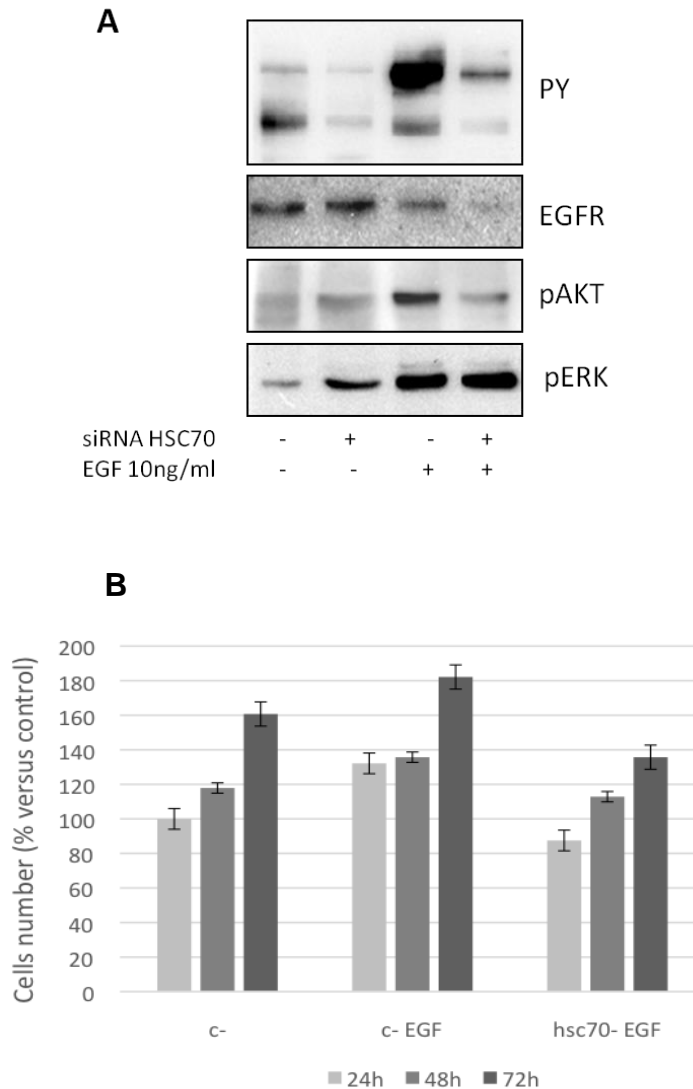


Fig. 5 – HSC70 silencing affects EGFR signaling.

A. Representative Western blot analysis that documents phosphorylation and expression of EGFR in wild type and HSC70 silenced cells, treated or not with 10 ng EGF for 24 hours. **B.** Cell viability of wild type and HSC70 silenced cells treated 24, 48 and 72 hours with 10 ng EGF. Bars represent means \pm SD of three independent measurements.

DISCUSSION

In the present study, we examined the role of HSC70 in REN cells in basal conditions and in the response to EGF treatment. HSC70 is a chaperone protein constitutively expressed in cells in normal conditions and is involved in protein folding; if cells are exposed to stress, the expression of HSC70 changes and often increases, to prevent mis-folding and de-regulation of proteins. As reported in many papers, tumour cells express high levels of HSC70. Silencing of HSC70 in REN cells, an epithelioid mesothelioma derived cell line, caused a strong decrease in cell viability that was reduced up to less than 50 % compared to the untreated cells.

Moreover, morphological changes observed in silenced cells (cells assumed a spindle shape) suggest the importance of this chaperone in the modulation of proteins involved in the organization of the cytoskeleton. In summary, our results show that a transient HSC70 knockdown decreased AKT phosphorylation; on the contrary, the total amount of protein was increased indicating that AKT was accumulated in an inactive state, and that HSC70 silencing prevented its phosphorylation, but probably acted on its stability. Furthermore, we demonstrated that AKT isoforms has different relationship with this chaperone protein: in fact in HSC70 silenced cells, AKT1 was increased while AKT3 was markedly reduced.

We evidenced in silenced cells a reduction of E-Cadherin mRNA and these data are consistent with the morphological changes and the observed increase of AKT1. Finally, EGF did not contrast the effects of HSC70 silencing, in terms of AKT activation, because also EGFR expression and signaling were affected.

Taken together, these data suggest that HSC70 acts modulating AKT stability and activity; AKT contributes to cell survival and AKT blockade by HSC70 knock-down may trigger cell death; the absence (or the alteration)

of HSC70 led to an increase and accumulation of AKT, in particular of AKT1 isoform.

In conclusion, this study suggests that HSC70 is strictly involved in AKT stability, and better understanding this inter-relationship, may be useful to provide new insights for therapeutic strategies.

References:

1. Daugaard M, Rohde M, Jaattela M 2007. The heat shock protein 70 family: Highly homologous proteins with overlapping and distinct functions. *FEBS Letters* 581:3702-10
2. Stricher F, Macri C, Ruff M, Muller S 2013. HSPA8/HSC70 chaperone protein. Structure, function, and chemical targeting. *Autophagy* 9(12):1937-54
3. Ungewickell E 1985. The 70-kd mammalian heat shock proteins are structurally and functionally related to the uncoating protein that releases clathrin triskelia from coated vesicles. *EMBO J* 4(13):3385-91
4. Flaherty KM, DeLuca-Flaherty C, McKay DB 1990. Three-dimensional structure of the ATPase fragment of a 70K heat-shock cognate protein. *Nature* 346:623-28
5. Kityk R, Kopp J, Sinning I, Mayer MP 2012. Structure and dynamics of the ATP-bound open conformation of Hsp70 chaperones. *Mol Cell* 48:863-74
6. Murata S, Minami Y, Minami M et al 2001. CHIP is a chaperone-dependent E3 ligase that ubiquitylates unfolded protein. *EMBO Rep* 2 :1133-38
7. Nirdé P, Derocq D, Maynadier M et al 2010. Heat shock cognate 70 protein secretion as a new growth arrest signal for cancer cells. *Oncogene* 29:117-27
8. Rouleau M, Patel A, Hendzel MJ et al 2010. PARP inhibition: PARP1 and beyond. *Nat Rev Cancer* 10: 293-301

9. Pinton G, Manente AG, Murer B et al 2013. PARP1 inhibition affects pleural mesothelioma cell viability and uncouples AKT/mTOR axis via SIRT1. *J Cell Mol Med* 17(2):233-41
10. Sundaresan NR, Pillai VB, Wolfgeher D et al 2011. The deacetylase SIRT1 promotes membrane localization and activation of Akt and PDK1 during tumorigenesis and cardiac hypertrophy. *Sci Signal* 4(182):ra46

Chapter 5

Conclusions

Malignant Mesothelioma (MM) is an asbestos-related highly aggressive cancer arising from the mesothelium that lines pleura, peritoneum or pericardium. It is characterized by a long latency period, poor prognosis and high chemo-resistance and its incidence, although the use of asbestos has been banned in many countries, has been predicted to increase dramatically worldwide during the next two decades. The PI3K/AKT signaling pathway is one of the major pathways involved in carcinogenesis and tumor progression; it is aberrantly active in several human cancer and it has been described to play an important role in human malignant pleural mesothelioma (MPM).

We firstly demonstrated that MPM cell lines express only two AKT isoforms, AKT1 and AKT3 and, differently from lung adenocarcinoma cells, they lack of AKT2; the levels of the two expressed isoforms resulted increased if compared with normal mesothelial cells and was related to tumor histotype.

Based on these data, in our first work we better characterized AKT isoforms expression and function in MPM cells.

AKT1 and -3 resulted increased in MPM derived cell lines and in particular the highest expression of both isoforms was observed in the MSTO-211H cell line established from a tumor with biphasic histotype. The same distribution was observed *in vivo* in freshly prepared surgical specimens.

To confirm the importance of the AKT pathway in this tumor, we investigated the effects of the MK2206 inhibitor on our cell model and evidenced both a significant reduction in AKTs phosphorylation and in cell viability. As the importance of AKT in mediating cisplatin resistance has been described, we tested the effects of the combined treatment with

cisplatin and MK2206. AKT inhibition resulted in enhanced cisplatin sensitivity with increased apoptotic cell death, as shown by PARP-1 cleavage.

Moreover, in order to define the specific role of each AKT isoform we evaluated the effect of specific silencing of AKT1 and -3. The specific knock-down of AKT1 or -3 had a strong effect on cell growth in monolayer, more evident for AKT3, while in terms of cisplatin sensitivity, both silencing of AKT1 or -3 reduced cell viability and increased apoptotic cell death, suggesting the essential, but different roles of these two isoforms.

The role of different AKTs in cell migration and metastases has been described in distinct tumors, but it remains to be better clarified, as different and conflicting studies suggest either positive or negative regulatory roles of each AKT isoform. As previously described, MSTO-211H cells are derived from a biphasic histotype, show a spindle morphology and express high level of both AKT1 and -3.

AKT1 silencing resulted in a reversion versus a more epithelial phenotype when MSTO-211H cells were cultured in monolayer, while upon AKT3 down-regulation the spindle morphology was stressed.

According to the evidenced morphological changes, AKT1 silenced cells showed an increase in E-Cadherin expression. Double silencing experiments underlined that this change was maintained both in the presence or in the absence of AKT3 expression, indicating that it strictly depends from AKT1 expression.

As described in different works, E-Cadherin is considered a marker of epithelial-mesenchymal transition and its increase or decrease corresponds to cell capability of invasion/migration. We explored if the silencing of AKT1 could affect cell adhesion and 3D cultures: AKT1-silenced cells resulted in a reduction of cell spreading on Matrigel and in a complete inhibition of colony growth in soft agar. Conversely, AKT3 knock-

down did not influence these processes: we didn't observe significant variations in the number of colonies which appeared even larger if compared to the controls. Taken together, all these results led us to establish a primary role for the endogenous AKT1 protein in affecting anchorage independent cell growth and underline the isoform-specific nature of this effect.

Again, we wanted to discriminate if the observed phenotype transition was a consequence of AKT1 loss-of-protein or was dependent by the loss of its activity: AKT inhibition with MK2206 did not cause the morphological changes observed in silenced cells. Conversely when AKT1 was overexpressed in epithelioid REN cells induced phenotypic changes towards more spindle-shaped cells.

Taken together all these data demonstrate that AKT1 and -3 contribute to cell survival, evidence distinct roles on MPM cell adhesion and highlighting that the observed effects mainly depends on the physical presence (and not on the activation status) of AKT1.

Drug-inhibited AKT1, in fact, lack of a specific kinase activity but could still retain interactions with other binding partners or assemble into macromolecular complexes.

As previously described in our laboratory, in MPM cells, AKT is in part phosphorylated and in part acetylated in basal conditions, while upon Sirt1 inhibition it becomes more acetylated and associates with different proteins, as shown by immunoprecipitation experiments.

Based on this evidence, the last aim of my studies was to explore the roles of these associated proteins and among those identified by MALDI-TOF analysis, we focused on HSC70, a chaperone protein constitutively expressed across tissues and cell types.

HSC70 is considered an essential *housekeeping* gene and in literature it has been reported that its silencing results in massive cell death.

We reported that silencing of HSC70 leads to a strong reduction in cell viability and in visible morphological changes, associated with a decrease in E-Cadherin expression. We further investigated the relationship between HSC70 and AKTs and the effects of its knock-down on AKTs activation.

Our data indicated that AKT1 phosphorylation was affected by silencing of HSC70, while the total protein amount resulted increased and interestingly our results demonstrated that EGF stimulation was not able to rescue AKT activation and that EGFR phosphorylation itself diminished. HSC70 silencing significantly compromised viability of MPM cells both in basal and EGF stimulated conditions.

In conclusion, these data suggest that HSC70 acts modulating AKT stability and activity; better understanding of this inter-relationship, may be useful to further explain the role of this protein in MPM and also provide new insights for novel therapeutic strategies.

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