# Inflammaging and Osteoarthritis

Francesca Motta<sup>1,2</sup> · Elisa Barone<sup>1,2</sup> · Antonio Sica<sup>1,3</sup> · Carlo Selmi<sup>1,2</sup>

Accepted: 18 April 2022 / Published online: 18 June 2022 © The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2022

## Abstract

Osteoarthritis is a highly prevalent disease particularly in subjects over 65 years of age worldwide. While in the past it was considered a mere consequence of cartilage degradation leading to anatomical and functional joint impairment, in recent decades, there has been a more dynamic view with the synovium, the cartilage, and the subchondral bone producing inflammatory mediators which ultimately lead to cartilage damage. Inflammaging is defined as a chronic, sterile, low-grade inflammation state driven by endogenous signals in the absence of infections, occurring with aging. This chronic status is linked to the production of reactive oxygen species and molecules involved in the development of age-related disease such as cancer, diabetes, and cardiovascular and neurodegenerative diseases. Inflammaging contributes to osteoarthritis development where both the innate and the adaptive immune response are involved. Elevated systemic and local inflammatory cytokines and senescent molecules promote cartilage degradation, and antigens derived from damaged joints further trigger inflammation through inflammasome activation. B and T lymphocyte populations also change with inflammaging and OA, with reduced regulatory functions, thus implicating self-reactivity as an additional mechanism of joint damage. The discovery of the underlying pathogenic pathways may help to identify potential therapeutic targets for the management or the prevention of osteoarthritis. We will provide a comprehensive evaluation of the current literature on the role of inflammaging in osteoarthritis and discuss the emerging therapeutic strategies.

Keywords Degenerative joint pain · Senior · Cartilage failure · Frailty

# Introduction

Osteoarthritis (OA) is a slowly progressing joint disease that predominantly affects the elderly population, affecting mobility, quality of life, and increasing mortality. Historically, it was thought to be a consequence of degenerative alterations leading to anatomical remodeling of the joints [1]. In the past decades, it has become increasingly clear that other factors are involved in the etiopathogenesis of OA and that the immune response to stressors plays a key role. In this

Carlo Selmi carlo.selmi@hunimed.eu

- <sup>1</sup> Division of Rheumatology and Clinical Immunology, IRCCS Humanitas Research Hospital, via Manzoni 56, 20089 Rozzano, Milan, Italy
- <sup>2</sup> Department of Biomedical Sciences, Humanitas University, via Rita Levi Montalcini, 20090 Pieve Emanuele, Milan, Italy
- <sup>3</sup> Department of Pharmaceutical Sciences, Università del Piemonte Orientale "Amedeo Avogadro", Largo Donegani 2, 28100 Novara, Italy

review, the role of inflammation in OA will be elucidated, in particular in light of new evidence related to the condition referred to as "inflammaging."

# The Burden of Osteoarthritis

OA is the most common form of joint disease. It involves the cartilage and other surrounding tissues, leading, in the advanced stages of the disease, to damage and loss of articular cartilage, remodeling of the subchondral bone with osteophyte formation, ligamentous laxity, and weakening of periarticular muscles [2]. OA predominantly affects the hands, hips, knees, feet, and cervical and lumbar spine while being less common in other joints. Pain is the typical symptom that leads people to present to healthcare providers and subsequently to receive a diagnosis of OA. Other primary symptoms include joint stiffness and limitation of movements. Progression is usually indolent and can eventually lead to complete joint impairment with pain and disability [3].



OA can be defined as a clinical or radiological entity. Clinical OA is defined by features in the history and in the medical examination, requiring invariably the presence of joint pain [3]. Some of the used standards for the diagnosis of clinical OA are the American College of Rheumatology (ACR) criteria which have been developed for hip [4], knee [5], and hand [6] OA.

Radiographic OA diagnosis and severity grade are often assessed in studies using the Kellgren and Lawrence score [3]. The overall grades of severity are determined from 0 to 4 and are related to the presumed sequential appearance of joint space loss, osteophytes, sclerosis, and cysts [7]. Magnetic resonance imaging (MRI) detects the disease earlier and provides evidence of matrix changes in cartilage, synovitis, bone marrow lesions, and degenerative changes in softtissue structures beyond the cartilage, including ligaments and the knee menisci [8, 9].

# Epidemiology

The number of people affected by OA is constantly rising worldwide, and OA is currently one of the major causes of disability in older adults [10]. As it is primarily related to aging, the prevalence of OA will increase and is expected to be the single greatest cause of disability in the general population by 2030 [2]. The prevalence of OA varies according to the definition of OA, the specific joint considered, and the features of the population studied. Incidence rates of symptomatic hand, hip, and knee OA increase rapidly around 50 years of age reaching a plateau after 70 years of age [11].

The prevalence of hand OA at imaging varies greatly and has been reported to range from 27 to over 80% [12]. Data from the Framingham Cohort demonstrated a prevalence of 13.2% in men and 26.2% in women aged 70 or more years with at least one hand joint with symptomatic OA [13]. Knee involvement occurs less frequently than hand OA, although both are more common in women. Among participants older than 45 years in the Framingham Study, the prevalence of radiographic knee OA was 19.2%, and in those over 80 years, it rose to 43.7%. Hip OA is the most unfrequently kind [3]. In addition, OA leads to a substantial amount of healthcare costs and resource use. In the USA, the estimated total annual average direct per-patient cost varied from \$1,442 to \$21,335 (adjusted to 2015 US\$ equivalent) [14]. The indirect costs on working activity have also to be taken into account. In a Swedish population-based study, the risk for sick leave or disability was more frequent among workers affected by knee OA [15].

OA is one of the major causes of disability around the globe [16] that invalidates an elevated number of everyday living activities [17], and years lived with disability were

exceptionally high among elderly people with OA [18]. Furthermore, OA may also negatively impact mental health, with higher probability to manifest depressive symptoms in patients with lower limb OA than those without the disease [19]. OA is also associated with greater odds of suicidal ideation [20]. A strong relationship between OA and perceived memory loss due to sleep and mood impairment was demonstrated [21]. There is also evidence that OA is a risk factor for cardiovascular disease (CVD) development, with a significant increase of the risk of myocardial infarction in patients having OA or other types of arthritis [22-24]. In the Chingford Cohort study, an increased risk of CVD-specific and all-cause mortality among women with symptomatic knee OA compared with an healthy group was found, while notably no relationship was found between hand OA and mortality risk [24, 25]. OA may impair the ability of people with cardio-metabolic conditions to exercise and lose weight, which is the core of the management of these conditions [26]. Among people with symptomatic hip and/or knee OA and concomitant diabetes, difficulty in walking predicts higher risk of serious complications related to diabetes [27] and an increased risk for diabetes in those who did not have diabetes at baseline [28]. Potential explanations include the influence of OA-related low-grade systemic inflammation on insulin resistance, the use of non-steroidal anti-inflammatory drugs (NSAIDs) for OA pain and weight gain for sedentary behavior [26, 29-31].

# **Risk Factors for OA**

There are several risk factors for OA. First, there are strong genetic bases causing the susceptibility to OA. Genetics accounts for 40–65% of OA risk [32], with a non-Mendelian manner inheritance, as OA is a multifactorial disease. Genetics alone is unlikely to identify individuals who will develop disease, but it might reveal new biological insights into disease pathogenesis for individual joints [33]. The genetic bases are stronger for hand and hip OA compared to knee OA [32, 34, 35]. The Arthritis Research UK Osteoarthritis Genetics (arcOGEN) Consortium has identified 11 loci associated with OA [36]. Genome-wide association studies (GWAS) have identified 90 independent susceptibility loci for OA [37]. In addition, single-nucleotide polymorphisms have been associated with several known risk factors, including hip shape, body mass index, and bone mineral density [38]. Several inflammation-related genes are involved in OA susceptibility (summarized in Table 1) reinforcing the concept that OA has an inflammatory component [39, 40]. Nonetheless, the OA loci discovered explain only a small fraction of OA heritability estimated by epidemiological studies.

Genes	Genetic loci	Normal function	Dysregulation in OA
SMAD3	15q22.33	It is an intracellular mediator of TGF- $\beta$ signaling which is involved in regulating chondrocyte proliferation, differentiation, and matrix mineralization	The shift from ALK5-SMAD2/3 dependent to AL1- SMAD1/5/8 dependent signaling leads to loss of chondrocyte homeostasis and to development of OA
RUNX2	6P21.1	It is essential in endochondral ossification but has to be suppressed to allow formation and maintenance of permanent cartilage	Its alteration leads to chondrocyte hypertrophy, cartilage destruction, and osteophyte development
PTHLH	12p11.22	It encodes the parathyroid hormone-related protein (PTHrP), and it is involved in chondrocyte differentiation rate control and in cartilage homeostasis in postnatal life	Its alteration leads to the reduction of PTHrP/PTH signaling
GDF5	20q11.22	It signals through SMAD1/5/8 pathway, enhancing mesenchymal stem cell differentiation to the chondrogenic lineage and promoting chondrocyte hypertrophy during endochondral ossification. It has important function in postnatal joint homeostasis and repair	It is overexpressed in osteophytic cartilage
CHST11	12q23.3	It encodes for carbohydrate sulfotransferase 1, a Golgi membrane enzyme that catalyzes the transfer of sulfate groups to chondroitin, a disaccharide covalently linked to the aggrecan core protein, forming the highly abundant extracellular matrix glycoprotein aggrecan	Gene knockout in mice led to fibrillation of the cartilage, similar to that seen in OA cartilage

Table 1 The major inflammation-related genes involved in osteoarthritis [40]

New evidences show that part of the risk may be due to epigenetic modifications of genomic DNA [41–44], illustrated in Table 2. This is of particularly interest since aging-related DNA methylation involves different genome portions according to gender: less than 5% of DNA methylation changes are common between males and females [45].

Age is the strongest risk factor for OA [12], since it leads to reduction in regenerative capacity and accumulation of risk factors. Compared to other races, African-Americans are also more likely to develop symptomatic knee and hip OA, with ethnic differences in radiographic OA features [11, 46]. Another risk factor is sex, since OA is more common in women than in men, even if the underlying mechanism remains unclear [47]. Congenital anatomical factors (hip dysplasia [33], femoroacetabular impingement [48], tibial and femoral bone abnormal morphology [49], varus or valgus knee alignment [50, 51], and leg length inequality [52] ) and functional factors like a poor quadriceps function [53], sporting activity [54], and a high-intensity activity [55] may all promote the development of OA. Despite these strong associations, individuals with abnormal joint biomechanics may never develop OA, as susceptibility is partly determined by other factors [48]. Specific bone properties could confer some susceptibility, as high bone mineral density seems to increase the risk of incident OA but not disease progression [56]. Injuries can cause bone or cartilage damage that makes the

Table 2 Proposed epigenetic modifications in OA [44]

Gene	Epigenetic alteration	Effects
P65 activation	DNA methylation	Potentiation of inflammatory response
Col10a1 expression	Hypomethylation	Increased expression during chondrocyte hypertrophy and maturation
MMP2, MMP9, MMP13, ADAMTS4	Hypomethylation	ECM degradation
COL9A1	Hypermethylation	Attenuates SOX9 (a transcription factor essential for differentiation of precursor cells into chondrocytes) binding to the COL9A1 promoter
IL-8	Hypomethylation	Increased expression
IL-1alfa and TNF	Hypomethylation	Increased MMP13 expression
COL2A1	Histone deacetylation	Gene expression suppression
MMP	Histone deacetylation	Induction of gene expressions
SIRT1	Histone deacetylation	Reduced expression in OA (this gene has a chondroprotective effect)
miR-146a	Decreased expression in human last stage OA cartilage while their levels raise in early stage OA	It is a regulator of inflammatory mediators: it directly targets Irak1 (IL-1 receptor-associated kinase 1) and Traf6 (TNF receptor-associated factor 6) which are regulators of NF-kB
miR-140	Loss of miR-140	Accelerated OA following aging or surgical destabilization

joint more susceptible to further insults and damage to ligaments or meniscus, adversely affecting joint biomechanics [57].

Obesity increases the risk of knee OA more than threefold and accelerates disease progression by loading the weightbearing joints and increasing joint susceptibility through the action of inflammatory adipokines [58, 59]. An independent association between weight gain and hip OA diagnosis has been shown [60]. Conversely, weight loss has been consistently associated with improvement of arthritis symptoms and slower knee cartilage degeneration [61, 62]. It has also been demonstrated that hyperlipidemia is an independent risk factor for new onset of hand OA [63], and higher levels of high-density lipoprotein cholesterol are protective against the incidence of radiographic hand OA [64]. In parallel, use of an antilipemic agent like ezetimibe is associated with fewer structural and better knee pain changes [65]. An association between higher systolic blood pressure and increased incidence of radiographic knee OA has been reported [66], while no data support an association between diabetes mellitus and hand/knee OA [67-69]. As vitamin D plays a major role in cartilage and bone metabolism, it has been hypothesized that low levels may increase OA risk. The VIDEO study, a randomized double-blind trial comparing a group of patients receiving placebo with a group of patients receiving vitamin D, showed that after 2 years of treatment, synovitis (as measured by MRI) remained stable among patients taking vitamin D, while it increased in the placebo group [70].

# New Insights into OA Pathogenesis and the Role of Inflammation

OA has historically been considered a "wear and tear" disease, where weight overload, anatomical alterations, or other mechanical triggers induced loss of cartilage, particularly in genetically susceptible individuals, with subsequent bone reparative attempt. Chondrocytes were the only cell type considered in OA etiopathogenesis, and as cartilage is not vascularized, local or systemic inflammation was not taken into account. In the last decades, new evidence has led to the insight that other tissues are involved in OA development, as synovium and subchondral bone, both able to produce inflammatory mediators and to induce cartilage degradation (Fig. 1 illustrates the main actors of OA pathogenesis) [71]. Cartilage destruction and loss of chondrocytes are the peculiar starting features of OA, and cartilage plays an active role in disease pathogenesis. In fact, the function of chondrocytes is crucial in maintaining the production and the homeostasis of the extracellular matrix (ECM), and although it is not clear which the first event is, chondrocytes death and matrix loss may form a vicious cycle, with the onset of one inducing the other [72]. In addition, the ECM can promote the development of OA, in particular the region immediately surrounding and enclosing the chondrocytes, named "chondron." It can act as a transducer of biochemical and biomechanical signals for the chondrocyte, regulating its metabolic activity in response to environmental signals. Chondron alterations could serve as a driver of OA [73].

The subchondral bone is also an active player in the development of OA. Imaging studies have demonstrated that bone changes can be observed even before cartilage lesions, and an association between subchondral bone mineral density and osteoarthritis has been shown [74]. Cartilage and subchondral bone are linked by regulatory pathways, which can be disrupted in OA. Among them, the Wnt and the OPG-RANKL-RANK systems contribute to the regulation of bone remodeling and are impaired in OA. The resulting imbalance between subchondral bone osteoblasts and osteoclasts leads to altered bone deposition and late formation of osteophytes [75, 76].

The synovium, constituting the cellular lining of the joint and producing the synovial fluid, is another tissue implicated in OA that becomes inflamed and hypertrophic. Despite its degree of inflammation does not reach that of rheumatoid arthritis, synovitis is present in all stages of OA, is related to pain, is a marker of severity, and can predict structural progression [77]. Imaging allows to detect synovial alterations in patients with OA. A large meta-analysis described that people with knee OA or knee pain had high prevalence of ultrasound-detected effusion, synovial hypertrophy, and power Doppler signals [78]. At MRI, synovitis correlated with radiographic OA severity in patients with all degree of knee OA [79]. Moreover, synovitis detected at MRI was a risk factor for OA even when adjusting for confounding pathologies [80].

#### Immune Mechanisms

Histologically, the synovium may show significant alterations, even before cartilage degeneration. Synovial lining hyperplasia, sublining fibrosis, and stromal vascularization are the typically patterns found. Early events, as release of debris after cartilage damage, induce the production of pro-inflammatory cytokines by synoviocytes, which leads to recruitment of leukocytes from the vascular compartment. The balance between anabolic and catabolic cytokines, which allow a proper turnover of cartilage, is impaired in OA. The cytokines upregulated in OA include interleukin-1 beta (IL-1 $\beta$ ), tumor necrosis factor (TNF)- $\alpha$ , IL-6, IL-15, IL-17, and IL-18. IL-1 $\beta$  and TNF- $\alpha$  are the most deeply studied and are elevated in OA synovial fluid, synovial membrane, cartilage, and subchondral bone and have an effect in inducing further inflammation and cartilage degradation.



Cytokines (IL-1β, IL-6, IL-7, IL-33, IL-15, IL-17, IL-18, TNFα), chemokines (CXCL13, CCL8, CCL5), adipokines (leptin, visfatin, adiponectin, resistin), alarmins, MMP, AGEs, ROS

**Fig. 1** Schematic representation of a knee (on the left) and of the main actors and inflammaging-related pathways involved in OA pathogenesis described in the text (on the right). Senescent joint tissues release pro-inflammatory mediators; among them, chondrocytes release IL-7, which induces cartilage degradation and the release of debris and extracellular matrix fragments. They induce inflammasome activation in chondrocytes and macrophage-like synoviocytes, with further release of cytokine and chemokines in the synovial fluid. In addition, biomechanical stress or catabolic activity further stimulates synovial cells. This process is enhanced by adipokines

The predominant cells observed in the OA synovium are T lymphocytes and macrophages, responsible, with activated synoviocytes, for further cytokine production and for the increased angiogenesis, leading to a vicious cycle which induces secretion of metalloproteinase and proteolytic enzymes, with perpetuation of cartilage degradation [81–84]. Of note, synovial fibroblasts undergo a transition process to activated and pro-inflammatory synoviocytes that is similar for OA and rheumatoid arthritis, although it is much more intense in the latter [85].

The local articular inflammation can be mirrored by plasmatic increase of inflammatory markers. Increased IL-6 and C-reactive protein levels have been detected in blood of individuals with OA compared with healthy controls and may predict the progression of the disease [86–89].

Adipokines are also involved in the pathogenesis of OA. In fact, despite obesity is a well-recognized risk factor for OA in relation to weight overload, adipose tissue and in particular infiltrated macrophages produce and secrete adipokines, proteins involved in modulation of inflammation, released by fat pads, in a vicious circle. These pro-inflammatory mediators induce synovial hypertrophy, neoangiogenesis, and infiltration by lymphocytes, in particular CD4+T cells, which can induce self-reactivity. The subchondral bone activity is dysregulated by the inflammatory environment, with impaired RANK-RANKL signaling and ectopic bone deposition and formation of osteophytes. Legend: RANK, receptor activator of nuclear factor kappa-B; RANKL, RANK ligand; IL, interleukin; CXCL, C-X-C motif chemokine ligand; CCL, C–C motif ligand; MMP, metalloproteinase; AGEs, advanced glycation end products; ROS, reactive oxygen species

angiogenesis, and glucose and lipid metabolism. Evidence suggests that chronically elevated local or systemic concentrations of adipokines have an important role in obesityassociated metabolic complications [90]. Elevated levels of adipokines have been detected in the infrapatellar fat pad, synovial fluid, and serum of patients with OA, with different levels detected in relation to the specific adipokine, the biological sample analyzed, and the clinical features of the patients [83, 91–93]. Studies on non-weight-bearing joints have been performed to distinguish the metabolic from the mechanical effect of fat excess and the relationship between OA and adipokines. They demonstrated that obesity is significantly associated with the development of clinical and radiological hand OA and that adipokine levels correlated with hand OA severity [94]. Distinct adipokines have been found to modulate bone remodeling and inflammatory responses, and their effects can change in relation to the stage of the disease. Leptin seems involved in inflammation, destruction of cartilage, and osteogenesis, as its serum levels have been correlated with presence of synovitis, cartilage defects, bone marrow lesions, and osteophytes. Adiponectin can enhance the matrix metalloproteinase (MMP)-mediated destruction of cartilage and bone, the osteogenic differentiation, and the capacity of osteoblasts to mineralize bone. It can also induce pro-inflammatory cytokines and can reduce osteoclastogenesis and osteoclast-mediated resorption activity. Resistin and visfatin seem involved in the production of pro-inflammatory cytokines and degradative enzymes [95, 96]. Interestingly, visfatin is related to the NAMPT/SIRT1 pathway, involved in metabolic redox reactions and in the modulation of inflammation. Nicotinamide phosphoribosyltransferase (NAMPT) is an intracellular key enzyme involved in nicotinamide adenine dinucleotide (NAD) salvage pathway. NAD, besides being an essential coenzyme involved in cellular redox reactions, also promotes the activation of sirtuin-1 (SIRT1), a histone deacetylase [97]. SIRT1-mediated histone deacetylation regulates gene expression, proteostasis, cell differentiation, and inflammation, also by inducing the release of myeloid suppressor-derived cells from the bone marrow [98, 99]. In cartilage, SIRT1 promotes the expression of protective genes, such as aggrecan and type II collagen, and inhibits chondrocyte apoptosis [100]. With aging, the expression levels of SIRT1 are reduced [101], and this might be deleterious both systemically and locally. Visfatin is the extracellular deacetylated form of NAMPT and plays a role in enhancing inflammatory pathways and in osteogenesis [95, 97]. Visfatin also enhances vascular endothelial growth factor (VEGF) expression in synovial fibroblasts, inducing neovascularization and OA progression [102]. The dual function of the intracellular and extracellular forms of NAMPT/visfatin, one with a protective role and the other with pro-inflammatory activity, has yet to be fully elucidated.

Another mechanism involves advanced glycation end products (AGEs), formed by a non-enzymatic process in aging tissues, often induced by environmental factors such as cigarette smoke. This process leads to the irreversible glycation of proteins and lipids, which undergo structural changes and the formation of cross-linkages. The modifications result in change of enzymatic functions, in accumulation and in damage of tissues. AGEs can accumulate in the cartilage and trigger chondrocyte activation by binding to their receptors (RAGEs). RAGEs are transmembrane receptors belonging to the immunoglobulin gene superfamily and can bind several AGEs and other several DAMPs as HMGB1 and amyloid- $\beta$ and induce inflammation by activation of the NF- $\kappa$ B and MAPK pathways [71, 103–105].

Toll-like receptors (TLRs) are involved in OA pathogenesis being expressed by immune cells, chondrocytes, osteoblasts, and synoviocytes. Their expression is upregulated in the OA synovium, and high-soluble TLR levels have been found in OA synovial fluid, as well as several damage-associated molecular patterns (DAMPs) able to induce inflammation [105].

#### Inflammaging in Clinics

Inflammaging is defined as a chronic, sterile, low-grade inflammation state, primarily driven by endogenous signals in absence of infection and involving self-reactivity, occurring during aging, leading to increased levels of pro-inflammatory mediators compared to younger individuals [106, 107]. Inflammaging has been proposed as a relevant risk factor for morbidity and mortality in the elderly population, as many age-related diseases share an inflammatory pathogenesis [108]. In the large scale, there is an increasing incidence worldwide of chronic age-related diseases, including obesity, metabolic syndrome, type 2 diabetes mellitus, and CVD, reflecting the growing aging population. Statistics show a raise of new cancer cases, attributed to increased risk factors as well as to the growth of the aging population [109]. There is a large overlap between the mechanisms driving aging and age-related diseases: common "pillars" are adaptation to stress, epigenetics, inflammation, macromolecular damage, metabolism, proteostasis, stem cells, and regeneration, which operate in an interconnected way, influencing and modulating each other to the constitution of an integrated network [110, 111]. Of major note, these pillars converge on chronic inflammation, and an impairment of any one fuels inflammation, which subsequently affects all the other pillars. Inflammation leads to increased levels of reactive oxygen species (ROS) inducing oxidative stress, which is a key component in the pathogenesis of chronic inflammation. Aging cells have increased levels of oxidant-damaged DNA [112], and oxidative stress can lead to the activation of systemic proinflammatory pathways [113], contributing to the pathogenesis of many age-related diseases. This has serious implication in tumorigenesis [114], development of diabetes [115], CVD [116], cognitive dysfunction and neurodegenerative diseases such as Alzheimer's and Parkinson's disease [117, 118], chronic obstructive pulmonary disease [119], and rheumatic diseases such as rheumatoid arthritis [120, 121].

One major characteristic of inflammaging is the chronic activation of the immune system [122]. Several cellular and molecular mechanisms are involved in inflammaging, including cellular senescence, mitochondrial dysfunction, defective autophagy, activation of the inflammasome, dysregulation of the ubiquitin-proteasome system, activation of the DNA damage response, and dysbiosis [123]. Senescent immune cells can acquire a pro-inflammatory phenotype since misplaced nucleic acids can accumulate and trigger an immune response, self-reactive T-cells can be released, while infections or gut dysbiosis can promote and modulate the inflammatory state. During life, chronic diseases, physical activity, stressors, infections, and nutrition may play a role in this process, contributing to the development of different phenotypes. Essentially, inflammaging can be considered the consequence of a dysregulated immune response [124].

#### **Causes of Inflammaging**

Inflammaging was originally attributed to somatic cell senescenceassociated secretory phenotype (SASP) and chronic innate immune activation [125-128]. In recent years, contribution of aged adaptive immune components, particularly self-reactive T lymphocytes, has been proposed as a probable contributor to the age-related development of subclinical autoimmune predisposition [124, 129]. In fact, in elderly, thymic involution reduces the capacity to maintain central tolerance, allowing escaping of selfreactive T cells, which participate in the process of inflammaging [130]. Other major contributors are persistent viral [131] and bacterial [132] infections, cell debris, misplaced self-molecules, and misfolded and oxidized proteins [108, 133]. Persistent infections by viruses such as cytomegalovirus may lead to chronic immune activation through persistent production of low levels of pro-inflammatory cytokines, such as IFNy and TNFa. In fact, the virus is found in a latent state in myeloid progenitor cells and monocytes, but also in epithelial cells of salivary glands and in the kidney, leading to chronic cytokine release; in addition, due to a severe impairment of the T cell system, highly differentiated exhausted CD28- CD4+ and CD8+T cells accumulate [129]. Furthermore, the alternation between reactivation and latency can alter lipid and glucose metabolism; in fact, cytomegalovirus infection restructures lipid rafts and has a deleterious effect on pancreatic β-cells, and the infection-related pro-inflammatory state may lead to type 2 diabetes [134–138].

Another factor linked to the activation of the immune response is nutrition, as food and water are contaminated by microbial stimuli. There is evidence that innate immunity is stimulated when food is ingested, starting "postprandial inflammation" that is part of the response to meals, as inflammatory markers increase after ingestion of food through several molecular mechanisms [139, 140]. The gut microbiota represents the boundary between diet, the host metabolism, and the innate immune response [141] and may also undergo profound remodeling with aging [142–145]. Its composition depends on several factors such as individual-based (age, gender, genetics, lifestyle, method of childbirth, and whether one was breastfed or formula fed), population-based (ethnicity, cultural habits, nutrition, population genetic structure, and ancestry), and environmentbased (climate, use of antibiotics, and lifelong immunological stimuli) factors, constituting a sort of biography of each person [146]. The adaptive and plastic nature of the gut microbiota allows it to adjust the host's immune and metabolic pathways in response to dietary habits and energy requirements having a profound effect on health and disease. The age-related remodeling of gut microbiota might contribute to systemic inflammaging, which can directly or indirectly affect its composition, in a self-sustaining loop [147, 148]. Changes in the gut microbiota profile in centenarians by an enrichment of Proteobacteria and a decrease in butyrate-producing bacteria are correlated with a systemic increase in levels of the pro-inflammatory cytokines IL-6 and IL-8 [149].

On the other hand, a low-calorie diet activates pathways stimulating autophagy, stress defense mechanisms, and survival pathways acting like a mild stressor that promotes hormetic responses [149, 150], leading to a general reduction of inflammation in the body with many effects on aging mechanisms including insulin resistance, adult neurogenesis and neuronal plasticity, autophagy, and mitochondrial biogenesis [151, 152]. Reduction of specific dietary proteins and amino acids without the restriction of calories currently seems to be the most effective dietary intervention for promoting longevity [149, 153].

## **Role of Inflammaging in OA**

Considered that OA is an age-related disease and that inflammation is strictly involved in its pathogenesis, inflammaging may play a relevant role in the development and in the evolution of the disease. Evidence suggests that OA is associated with low-grade systemic and local inflammation, and both the innate and the adaptive immune response are involved.

With aging, joints can release pro-inflammatory mediators; in mouse models, a different genetic expression in joint tissues has been described with aging, with an upregulation of pro-inflammatory cytokines and chemokines, in particular of IL-33, CXCL13, CCL8, and CCL5 [154]. Human chondrocyte cultures derived from elderly release higher levels of IL-7 compared to cultures from young subjects. Moreover, chondrocytes from patients with OA release more IL-7 than age-matched subject without OA, and IL-7 leads to increase of metalloproteinases and proteoglycan release from cartilage [155]. The term "chondrosenescence" has been coined to include the age-related alterations of chondrocytes, due to systemic or local stimuli, as increased levels of ROS and pro-inflammatory cytokines, fluctuating oxygen and nutrient supply due to vascular impairment, reduced cartilage water content and elasticity, and increased advanced glycation end products [156].

This process can trigger the inflammasome, via the pathogenassociated molecular patterns (PAMPs) or damage-associated molecular patterns (DAMPs). In OA, the principal DAMPs include hydroxyapatite, basic calcium phosphate, calcium pyrophosphate dihydrate, or uric acid crystals. These are capable to activate intracellular NLRP3, a major component of the inflammasome, highly expressed in macrophages, chondrocytes, synoviocytes, and osteoblasts [157]. Subjects with innate genetic-related reduced capacity to produce IL-1 $\beta$  and IL-6 are protected from OA in old age, while no impact was found for TNF $\alpha$  and IL-10 [158]. IL-1 $\beta$ , produced by the inflammasome, is involved in the link between inflammaging and OA, as IL-1 $\beta$ expression is increased in OA [159] and synovial fibroblasts stimulated with IL-1 $\beta$  induce OA-like modifications in the joint

[160, 161]. IL-6 levels are associated with the risk of OA progression [88], and in chondrocytes, IL-6 appears to interact with basic calcium phosphate to promote cartilage degradation and OA [162]. An association between pro-inflammatory cytokines and articular functionality or level of pain has also been demonstrated [156]. The age-related increase in fat mass could contribute to the increased levels of adipokines and cytokines involved in OA pathogenesis. This can occur both systemically, given the increment of the visceral fat, and locally, due to the increase in fat pad volume into the knee joints [163]. Evidence also shows that senescent cells accumulate in degenerated tissues and induce the expression of senescent markers. Cartilage and synovium are the sites where senescent cells localize in experimental OA and secrete matrix degrading proteins and cytokines, thus inducing chronic inflammation. The molecules released exacerbate the disease and seem to affect the subchondral bone, with dysregulation of bone remodeling and osteophyte formation [157].

Autophagy, the homeostatic mechanism of removal of macromolecules or dysfunctional organelles as inflammasomes through lysosome degradation, is reduced in OA [156, 164], and animal models have shown that autophagy activation can prevent OA damage [165]. The adaptive immune response may connect inflammaging and OA. An analysis of blood lymphocyte composition in OA patients and healthy controls showed that T and B cell function was compromised, with CD4, T regulatory, and B-cell decreased in OA, while CD8 increased in OA compared to controls, possibly reflecting inflammation and autoreactivity [166]. In other studies, T and B cell profiles in OA patients were proved altered compared to healthy controls, although with different features among reports. Nevertheless, they all suggest that the acquired immune system is impaired in OA and may play a relevant role in the pathogenesis of OA [167, 168]. Locally, an increased infiltrate of T lymphocytes has been shown in OA synovium compared to healthy synovium [169], accounting for up to 25% of inflammatory cells [170]. Moreover, an increased CD4+/CD8+ratio has been reported in synovium from OA patients compared to controls [171], and the number of synovial CD4 + lymphocytes has been associated with the pain visual analog scale [83]. Other lymphocytes, including B and NK, and plasma cells have been detected in synovial tissue from OA patients, although their precise role still has to be elucidated [172].

The alterations due to systemic and local factors lead to an inflammatory environment, with increased cytokines, ROS, adipokines, proteinases, and self-reactive lymphocytes; in joints, this ultimately results in chondrosenescence and degenerative modifications, which further enhance inflammation and autoimmunity, in a vicious circle. One hypothesis is based on the existence of a spectrum of OA phenotypes, with a portion of subject in whom inflammaging is the primary driver, as identified with imaging and serum biomarkers, and who may benefit from anti-inflammatory treatments [173]. In this

context, the concept of frailty can be introduced as its definition, and implementation in clinical practice can help in the management of patients with OA. Frailty is a clinical syndrome characterized by physiological changes across multiple systems and organs, resulting in a progressive loss of internal homeostasis, reduced physiological reserves, loss of function, reduced resilience, compromised ability to cope with everyday or acute stressors, and increased vulnerability to internal and external stressors [174, 175].

The pathogenesis of frailty is related to multiple factors. Polymorphisms in genes involved in inflammation, metabolism, endocrine system, and homeostasis have been proposed, although acquired and environmental factors appear to be predominant. Inflammaging plays a role in accelerated aging in individuals with chronic diseases and multimorbidity, mediates the development of functional limitations and disability, impairs the mechanisms of homeostasis and repair that occur in healthy and damaged tissue, and can therefore have detrimental effects and contribute to frailty. Chronic low-grade inflammation in inflammaging leads to sarcopenia, loss of strength, increased risk of falls, reduced physical activity, anemia, cardiovascular diseases, and poor nutrition, all features well described in frailty syndrome. Moreover, in the frail syndrome, low-grade inflammation persists after removal of the trigger, and the immune system is hyperresponsive to further stimuli; acute illness or exacerbation of chronic conditions may further worsen the clinical scenario. Inflammaging and frailty are thus closely related, and ultimately frailty has been regarded as an inflammaging-related disease [124, 176] or the clinical counterpart of inflammaging. OA appears to be closely related to inflammaging and frailty, with high prevalence of frailty in patients with OA since low-grade systemic inflammation, pain, impaired joint function, and disability due to OA contribute to reduced movement autonomy and decreased resilience typical of frailty [177]. Of clinical relevance, frailty predicts mortality in patients with OA [178], and this suggests that targeting inflammaging would affect not only the mechanism involved in the development and worsening of OA, but also the complex pathways involved in homeostasis and in inflammagingrelated diseases, thus potentially reducing vulnerability to stressors and increasing survival.

# Potential Therapeutic Targets and Biomarkers

Currently, there is no effective treatment to prevent or reverse OA short of prosthetic surgery, despite several molecules, even with anti-inflammatory properties, have been studied based on basic science.

Based on the unique OA characteristics, the use of intraarticular injection of platelet-rich plasma (PRP) has been proposed. PRP is an autologous serum containing high concentrations of platelets and growth factors, obtained by centrifugation of blood. Growth factors and cytokines are released after degranulation of platelet alpha-granules. In vitro experiments have demonstrated that they can increase the synthetic capacity of chondrocytes through upregulation of gene expression, proteoglycan production, and type II collagen deposition, thereby accelerating cartilage matrix synthesis, promoting cartilage healing, and reducing synovial inflammation. Many studies have shown great pain relief and improved patient-reported outcomes with PRP injection into the knee compared with other intraarticular injections, such as corticosteroid or hyaluronic acid, or oral medications. There is less evidence on the efficacy of PRP injection into the hip compared with hyaluronic acid. No differences in side effects have been found between PRP, hyaluronic acid, and placebo [179-186].

Drugs targeting IL-6, IL-1, and TNF- $\alpha$  have demonstrated no effect. On the other hand, antagonists of colony-stimulating factors (CSFs), molecules which increase the myeloid population during an inflammatory response, showed beneficial effects in preventing disease development and pain in experimental preliminary analyses [187, 188]. Blocking granulocyte macrophage CSF (GM-CSF) receptor prevents and reverses arthritic pain and is able to modulate the degree of osteoarthritic changes by reducing osteophyte formation in the joint of OA-induced murine experimental models. There is now ongoing a phase II trial in hand OA exploring potential of one of this molecule (GSK3196165) in the future for the prevention of OA damage and OA-related pain [187, 189].

As high levels of IL-4R and IL-10R have been described in the knee of patients with OA compared with healthy controls, drugs targeting these molecules with anti-inflammatory properties have been studied [190]. IL4-10FP in vitro can reduce release of inflammatory, catabolic, and pain-related mediators from human OA cartilage and synovial tissue, and it beneficially affects proteoglycan turnover. According to these data, IL4-10FP reduces pain in animal models [190]. A recent study by van Helvoort et al. demonstrated for the first time that in rats, repeated intra-articular injections of rIL4-10 FP, compared to saline solution injections, resulted in preventing/reducing joint damage in early OA stages, as well as relieving OA-induced pain [191].

More subtle and inflammaging-targeted treatments are now under investigation, and promising results are expected. In particular, recent research focuses on pattern recognition receptors (PRRs), which bind PAMPs and DAMPs and trigger inflammation. Among them, TLRs and RAGEs appear as promising targets. TLRs can be blocked by high-molecular weight hyaluronic acid, Pep-1 and a 12-mer peptide, which prevent the binding of DAMPs as short hyaluronic acid oligosaccharides. The activation of TLR antagonist pathways can be induced by prostaglandin 2, vasoactive intestinal peptide, adenosine 2A receptor, bone morphogenic protein 7, and peroxisome proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ). The latter is activated by rosiglitazone and pioglitazone, commonly used for type 2 diabetes, which are now under evaluation as they inhibit TLR stimulation in OA synoviocytes and chondrocytes and reduce inflammatory response in vitro [105, 192–194]. In animal experiments, pioglitazone arrests and reverses AGE-induced arthritis by reducing inflammation and cartilage damage by MAPKs and NF-kB signaling inhibition. All these evidences suggest the potential of this drug in early treatment of OA, especially among diabetic patients [193, 195–197].

RAGEs can also be targeted by its soluble receptor, sRAGE. In OA plasmatic levels of sRAGE are significantly lower compared to those in healthy controls [198]. sRAGE acts as a competitive inhibitor of RAGE, scavenges DAMP/PAMP ligands, and reduces inflammatory signals. It can modulate the chronic low-grade inflammation occurring in elderly and has been proposed as anti-inflammaging-related disease molecule [199].

Since many infectious agents are recognized by the same PRRs as DAMPs, directly blocking TLRs or RAGEs could impair the host immune defense against pathogens. Therefore, specifically addressing DAMPs as alarmins S100 and HMGB1 might be a safer approach. Alarmins are secreted by alternative pathways not involving the endoplasmic reticulum or the Golgi complex; therefore, inhibiting these pathways could block the release of multiple alarmins. Glycyrrhizin and ethyl pyruvate act on these mechanisms. In OA rat models, treatment with glycyrrhizin led to hyperalgesia and joint edema improvement, with suppression of serum levels of cartilage catabolic markers and decreased serum and intra-articular levels of IL-1β, IL-6, TNFα, and inducible nitric oxide synthase (iNOS) [200]. Furthermore, ethyl pyruvate blocks the NLRP3 inflammasome involved in the release of both IL-1 and alarmins [201].

Individual alarmins can also be blocked: antibodies against HMGB1 and S100A8/A9 have been studied, and recently identified epitopes of the S100A8/A9 complex capable of activating TLRs could further guide the production of specific antibodies. Natural or synthetic small molecules can target alarmins, and among them, the S100A9 inhibitor paquinimod proved effective to reduce synovial inflammation, Wnt signaling, osteophyte formation, and cartilage damage in a preclinical model of OA [105, 202–209]. The potential therapeutic targets in OA, considered an inflammaging-related disease, are summarized in Table 3.

One more proposed therapeutic approach is based on the removal of senescent cells to prevent the secretion of proinflammatory factors produced by an impaired regulation of oxidative processes. Studies in vitro on chondrocytes from OA patients undergoing total knee replacement showed that removal of senescent cells reduced the expression of pro-inflammatory markers and improved the expression 
 Table 3
 Potential therapeutic

 targets in inflammaging-related
 mechanisms of OA

Potential therapeutic molecule	Target	References
-Anti-colony-stimulating factors	Colony-stimulating factors	[187, 188]
-Fusion protein of IL-4 and IL-10 (IL4-10 FP)	IL-4 and IL-10	[190, 191]
-High-molecular weight hyaluronic acid -Pep-1 -12-mer peptide	TLR blockers	[105, 192, 194]
-Pioglitazone -Rosiglitazone	Activation of TLR antagonist pathways via PPARγ	[105, 193]
-Soluble RAGE	RAGEs	[199]
-Glycyrrhizin -Ethyl pyruvate	Alarmins, i.e., S100 and HMGB1	[202, 204]
-Ethyl pyruvate	NLRP3 inflammasome	[203]
-HMGB1 and S100A8/A9 blockers	HMGB1 and S100	[205, 209]
-Paquinimod	S100A9 inhibition	[206]

*IL* interleukin, *TLR* Toll-like receptor, *PPAR* $\gamma$  peroxisome proliferator-activated receptor  $\gamma$ , *RAGE* receptor for advanced glycation end products, *HMGB1* high mobility group box 1, *NLRP3* NLR family pyrin domain containing 3

of extracellular matrix proteins. In a mouse model of OA, removal of senescent cells accumulated in the cartilage and in the synovium attenuated the development of OA. In vitro studies on chondrocytes from patients with OA undergoing total knee replacement showed that removal of senescent cells reduced the expression of pro-inflammatory markers and improved the expression of extracellular matrix proteins. Senolytics as ABT-263 can inhibit the anti-apoptotic proteins belonging to the BCL2 family members, and after intra-articular injection, they can target senescent cells and prevent cartilage damage [157, 210]. The development of drugs to modify structural deterioration of the joint (disease-modifying OA drugs, DMOADs) requires measurable and universally applicable and accepted endpoints beyond traditional patient-reported outcomes [211].

Collagen is the main component of articular cartilage, and type II is the most represented. Many biomarkers are associated to type II collagen turnover, such as urinary C-telopeptide fragments of type II collagen (u-CTXII), serum (s) N-propeptide of collagen IIA (PIAANP), and serum N-propeptide of collagen IIB (PIIBNP or PRO-C2). The u-CTX II is a product of type II collagen degradation generated by MMPs, and high baseline levels of u-CTXII predict increased risk to undergo total joint replacement of hip or knee [211, 212]. Conversely, PIAANP and PIIBNP are two splice variants of N-terminal propeptide of type II collagen reflecting cartilage formation [213]. High serum PIIANP predicts reduced progression of clinically relevant knee OA over 4 years [212], and PIIANP and PIIBNP levels are lower in patients with established OA compared to controls [213, 214].

At the early OA stages, first changes appear in the subchondral bone rather than in cartilage, with horizontal trabeculae thickening followed by vertical trabeculae thinning in a second phase [212]. Bone biomarker abnormalities, as of urinary N-terminal telopeptide of type I collagen (uNTX-I), u-CTX-I, and serum CTX-I, are able to detect disease at early stages [215–217]. In the contest of inflammation, some synovial inflammatory fluid biomarkers specifically connected with macrophage and neutrophil activation have been demonstrated to be associated with synovitis and radiographic and symptom severity, i.e., metalloproteinase-3 (MMP3), soluble vascular cell adhesion molecule-1 (sVCAM1), soluble intercellular cell adhesion molecule-1 (sICAM1), VEGF, tissue inhibitor of metalloproteinases 1 (TIMP1), and monocyte chemoattractant protein 1 (MCP1) [218, 219]. As mentioned, reduced serum iNOS, MMPs, and TNF $\alpha$  levels have been described during experimental treatment with pioglitazone and glycyrrhizin [196, 200] and can be evaluated in future studies as biomarkers in OA patients.

S100A8/A9 are the best studied alarmins in the contest of OA, with high serum and synovial levels in the course of the disease, strongly related to synovitis and inflammation [220, 221]. Furthermore, their levels increase with aging, the principal risk factor for disease development [222], and they have been described as markers in many inflammatory arthritis [223–225]. The evaluation of their serum levels could be useful for monitoring specific target therapies, such as paquinimod treatment.

#### **Conclusions and Future Perspectives**

Despite being a condition affecting nearly a third of the world population and a growing amount of basic evidence on the mechanisms of disease, our therapeutic approach to OA is largely unsatisfactory. Recent research pointed out that several pathways and molecules in cartilage, subchondral bone, adipose tissue, and synovium are involved and that inflammaging plays a major role. A hypothesis is that in patients with OA, mechanisms underlying the disease may differ. Therefore, beyond screening patients at risk to develop the disease by dosing bone biomarkers as serum or urinary CTX, research should focus on identifying and validating biomarkers to detect at early stages patients more at risk of developing severe OA, in order to stratify the population. To this regard, intriguing could be to study serum levels of alarmins and their modifications according to cartilage structural damage. Furthermore, their evaluation could be helpful to monitor the disease in future clinical trials, particularly if S100A8/A9 blockers such as paquinimod will be implemented in clinical practice. Similar strategies could be used to investigate the efficacy of molecules that currently appear promising in preventing or reversing OA early damage, such as IL4-10 FP and GSK3196165. Furthermore, patients affected by diabetes starting treatment with pioglitazone and rosiglitazone could be studied; understanding the impact of these drugs on joint damage could be relevant to define their role in the treatment of OA.

A deeper understanding of the key inflammatory pathways leading to OA could improve not only the treatment, but more importantly allow to early address target molecules and therefore prevent the disease, paving the way to a true personalized approach and reducing its high morbidity and mortality.

**Author Contribution** Both FM and CS conceived the idea for the paper. FM and EB performed the literature search and drafted the manuscript. CS and AS provided helpful suggestions in the preparation of this manuscript, and all the authors reviewed and approved the final version of the paper.

**Funding** FM, AS, and CS were funded by the Italian Ministry of Health (Ricerca Corrente and grant RF-2016–02364842).

Data Availability Not applicable.

Code Availability Not applicable.

#### Declarations

**Ethics Approval** This article does not contain any studies with human participants or animals performed by any of the authors.

Consent to Participate Not applicable.

Consent for Publication Not applicable.

Conflict of Interest The authors declare no competing interests.

# References

1. Bland JH, Cooper SM (1984) Osteoarthritis: a review of the cell biology involved and evidence for reversibility. Management

rationally related to known genesis and pathophysiology. Semin Arthritis Rheum 14:106–133

- Hutton CW (1989) Osteoarthritis: the cause not result of joint failure? Ann Rheum Dis 48:958–961. https://doi.org/10.1136/ ard.48.11.958
- Litwic A, Edwards MH, Dennison EM, Cooper C (2013) Epidemiology and burden of osteoarthritis. Br Med Bull 105:185–199. https://doi.org/10.1093/bmb/lds038
- Altman R, Alarcón G, Appelrouth D et al (1991) The American college of rheumatology criteria for the classification and reporting of osteoarthritis of the hip. Arthritis Rheum 34:505–514. https://doi.org/10.1002/art.1780340502
- 5. Altman R, Asch E, Bloch D et al (1986) Development of criteria for the classification and reporting of osteoarthritis: classification of osteoarthritis of the knee. Arthritis Rheum 29:1039–1049. https://doi.org/10.1002/art.1780290816
- Altman R, Alarcon G, Appelrouth D et al (1990) The American College of Rheumatology criteria for the classification and reporting of osteoarthritis of the hand. Arthritis Rheum 33:1601– 1610. https://doi.org/10.1002/art.1780331101
- Kellgren JH, Lawrence JS (1957) Radiological assessment of osteoarthrosis. Ann Rheum Dis 16:494–502. https://doi.org/10. 1136/ard.16.4.494
- Loeser RF, Goldring SR, Scanzello CR, Goldring MB (2012) Osteoarthritis: a disease of the joint as an organ. Arthritis Rheum 64:1697–1707
- Sharma L, Chmiel JS, Almagor O et al (2014) Significance of preradiographic magnetic resonance imaging lesions in persons at increased risk of knee osteoarthritis. Arthritis Rheumatol 66:1811–1819. https://doi.org/10.1002/art.38611
- Hunter DJ, March L, Chew M (2020) Osteoarthritis in 2020 and beyond: a Lancet Commission. Lancet 396:1711–1712
- Zhang Y, Jordan JM (2010) Epidemiology of osteoarthritis. Clin Geriatr Med 26:355–369
- Lawrence RC, Felson DT, Helmick CG et al (2008) Estimates of the prevalence of arthritis and other rheumatic conditions in the United States. Part II Arthritis Rheum 58:26–35. https://doi.org/ 10.1002/art.23176
- Zhang Y, Niu J, Kelly-Hayes M et al (2002) Prevalence of symptomatic hand osteoarthritis and its impact on functional status among the elderly: the Framingham study. Am J Epidemiol 156:1021–1027. https://doi.org/10.1093/aje/kwf141
- Xie F, Kovic B, Jin X et al (2016) Economic and humanistic burden of osteoarthritis: A systematic review of large sample studies. Pharmacoeconomics 34:1087–1100
- Hubertsson J, Turkiewicz A, Petersson IF, Englund M (2017) Understanding occupation, sick leave, and disability pension due to knee and hip osteoarthritis from a sex perspective. Arthritis Care Res 69:226–233. https://doi.org/10.1002/acr.22909
- Cross M, Smith E, Hoy D et al (2014) The global burden of hip and knee osteoarthritis: estimates from the global burden of disease 2010 study. Ann Rheum Dis 73:1323–1330. https://doi. org/10.1136/annrheumdis-2013-204763
- Haan MN, Lee A, Odden MC et al (2016) Gender differences in the combined effects of cardiovascular disease and osteoarthritis on progression to functional impairment in older Mexican Americans. J Gerontol - Ser A Biol Sci Med Sci 71:1089–1095. https://doi.org/10.1093/gerona/glw014
- Park JI, Jung HH (2017) Estimation of years lived with disability due to noncommunicable diseases and injuries using a populationrepresentative survey. PLoS ONE 12. https://doi.org/10.1371/ journal.pone.0172001
- Veronese N, Stubbs B, Solmi M et al (2017) Association between lower limb osteoarthritis and incidence of depressive symptoms: data from the osteoarthritis initiative. Age Ageing 46:470–476. https://doi.org/10.1093/ageing/afw216

- Kye SY, Park K (2017) Suicidal ideation and suicidal attempts among adults with chronic diseases: a cross-sectional study. Compr Psychiatry 73:160–167. https://doi.org/10.1016/j.comppsych.2016. 12.001
- Innes KE, Sambamoorthi U (2018) The association of perceived memory loss with osteoarthritis and related joint pain in a large Appalachian population. Pain Med (United States) 19:1340– 1356. https://doi.org/10.1093/pm/pnx107
- Schieir O, Tosevski C, Glazier RH et al (2017) Incident myocardial infarction associated with major types of arthritis in the general population: a systematic review and meta-analysis. Ann Rheum Dis 76:1396–1404. https://doi.org/10.1136/annrheumdis-2016-210275
- Chung WS, Lin HH, Ho FM et al (2016) Risks of acute coronary syndrome in patients with osteoarthritis: a nationwide populationbased cohort study. Clin Rheumatol 35:2807–2813. https://doi. org/10.1007/s10067-016-3391-x
- Courties A, Sellam J, Maheu E et al (2017) Coronary heart disease is associated with a worse clinical outcome of hand osteoarthritis: a cross-sectional and longitudinal study. RMD Open 3. https://doi.org/10.1136/rmdopen-2016-000344
- 25. Gao SG, Zeng C, Xiong YL et al (2016) Is painful knee an independent predictor of mortality in middle-aged women? Ann Rheum Dis 75:e22
- Piva SR, Susko AM, Khoja SS et al (2015) Links between osteoarthritis and diabetes: implications for management from a physical activity perspective. Clin Geriatr Med 31:67–87
- Hawker GA, Croxford R, Bierman AS et al (2017) Osteoarthritisrelated difficulty walking and risk for diabetes complications. Osteoarthr Cartil 25:67–75. https://doi.org/10.1016/j.joca.2016. 08.003
- Jeon CY, Lokken RP, Hu FB, Van Dam RM (2007) Physical activity of moderate intensity and risk of type 2 diabetes: a systematic review. Diabetes Care 30:744–752
- Messier SP, Mihalko SL, Legault C et al (2013) Effects of intensive diet and exercise on knee joint loads, inflammation, and clinical outcomes among overweight and obese adults with knee osteoarthritis: The IDEA randomized clinical trial. JAMA - J Am Med Assoc 310:1263–1273. https://doi.org/10.1001/ jama.2013.277669
- Duncan BB, Schmidt MI, Pankow JS et al (2003) Low-grade systemic inflammation and the development of type 2 diabetes: the atherosclerosis risk in communities study. Diabetes 52:1799– 1805. https://doi.org/10.2337/diabetes.52.7.1799
- Rahman MM, Cibere J, Anis AH et al (2014) Risk of type 2 diabetes among osteoarthritis patients in a prospective longitudinal study. Int J Rheumatol 2014. https://doi.org/10.1155/ 2014/620920
- 32. Neogi T, Zhang Y (2013) Epidemiology of osteoarthritis. Rheum Dis Clin North Am 39:1–19
- 33. Agricola R, Heijboer MP, Roze RH et al (2013) Pincer deformity does not lead to osteoarthritis of the hip whereas acetabular dysplasia does: acetabular coverage and development of osteoarthritis in a nationwide prospective cohort study (CHECK). Osteoarthr Cartil 21:1514–1521. https://doi.org/10.1016/j.joca. 2013.07.004
- Valdes AM, Spector TD (2011) Genetic epidemiology of hip and knee osteoarthritis. Nat Rev Rheumatol 7:23–32. https:// doi.org/10.1038/NRRHEUM.2010.191
- Loughlin J (2005) The genetic epidemiology of human primary osteoarthritis: current status. Expert Rev Mol Med 7. https:// doi.org/10.1017/S1462399405009257
- 36. Zeggini E, Panoutsopoulou K, Southam L et al (2012) Identification of new susceptibility loci for osteoarthritis (arcOGEN): a genome-wide association study. Lancet 380:815–823. https:// doi.org/10.1016/S0140-6736(12)60681-3

- Warnera SC, Valdesa AM (2017) Genetic association studies in osteoarthritis: is it fairytale? Curr Opin Rheumatol 29:103–109
- Hochberg MC, Yerges-Armstrong L, Yau M, Mitchell BD (2013) Genetic epidemiology of osteoarthritis: recent developments and future directions. Curr Opin Rheumatol 25:192–197
- Rogers EL, Reynard LN, Loughlin J (2015) The role of inflammation-related genes in osteoarthritis. Osteoarthr Cartil 23:1933–1938. https://doi.org/10.1016/J.JOCA.2015.01.003
- Reynard LN, Loughlin J (2013) Insights from human genetic studies into the pathways involved in osteoarthritis. Nat Rev Rheumatol 9:573–583. https://doi.org/10.1038/NRRHEUM. 2013.121
- Goldring MB, Marcu KB (2012) Epigenomic and microRNAmediated regulation in cartilage development, homeostasis, and osteoarthritis. Trends Mol Med 18:109–118. https://doi. org/10.1016/J.MOLMED.2011.11.005
- Barter MJ, Bui C, Young DA (2012) Epigenetic mechanisms in cartilage and osteoarthritis: DNA methylation, histone modifications and microRNAs. Osteoarthr Cartil 20:339–349. https:// doi.org/10.1016/J.JOCA.2011.12.012
- Loughlin J, Reynard LN (2015) Osteoarthritis: epigenetics of articular cartilage in knee and hip OA. Nat Rev Rheumatol 11:6–7. https://doi.org/10.1038/NRRHEUM.2014.189
- 44. Shen J, Abu-Amer Y, O'Keefe RJ, McAlinden A (2017) Inflammation and epigenetic regulation in osteoarthritis. Connect Tissue Res 58:49–63. https://doi.org/10.1080/03008207. 2016.1208655
- Unnikrishnan A, Freeman WM, Jackson J et al (2019) The role of DNA methylation in epigenetics of aging. Pharmacol Ther 195:172–185. https://doi.org/10.1016/J.PHARMTHERA.2018. 11.001
- Allen KD, Golightly YM (2015) State of the evidence. Curr Opin Rheumatol 27:276–283
- Glyn-Jones S, Palmer AJR, Agricola R et al (2015) Osteoarthritis. In: The Lancet. Lancet Publishing Group, pp 376–387
- Agricola R, Waarsing JH, Arden NK et al (2013) Cam impingement of the hip-a risk factor for hip osteoarthritis. Nat Rev Rheumatol 9:630–634
- Neogi T, Bowes MA, Niu J et al (2013) Magnetic resonance imaging-based three-dimensional bone shape of the knee predicts onset of knee osteoarthritis: data from the osteoarthritis initiative. Arthritis Rheum 65:2048–2058. https://doi.org/10.1002/art. 37987
- Sharma L, Chmiel JS, Almagor O et al (2013) The role of varus and valgus alignment in the initial development of knee cartilage damage by MRI: the MOST study. Ann Rheum Dis 72:235–240. https://doi.org/10.1136/annrheumdis-2011-201070
- 51. Felson DT, Niu J, Gross KD et al (2013) Valgus malalignment is a risk factor for lateral knee osteoarthritis incidence and progression: findings from the multicenter osteoarthritis study and the osteoarthritis initiative. Arthritis Rheum 65:355–362. https://doi. org/10.1002/art.37726
- Harvey WF, Yang M, Cooke TDV et al (2010) Association of leg-length inequality with knee osteoarthritis a cohort study. Ann Intern Med 152:287–295. https://doi.org/10.7326/0003-4819-152-5-201003020-00006
- 53. Wang Y, Wluka AE, Berry PA et al (2012) Increase in vastus medialis cross-sectional area is associated with reduced pain, cartilage loss, and joint replacement risk in knee osteoarthritis. Arthritis Rheum 64:3917–3925. https://doi.org/10.1002/art. 34681
- Lievense AM, Bierma-Zeinstra SMA, Verhagen AP et al (2003) Influence of sporting activities on the development of osteoarthritis of the hip: a systematic review. Arthritis Care Res 49:228–236

- 55. Siebenrock KA, Kaschka I, Frauchiger L et al (2013) Prevalence of cam-type deformity and hip pain in elite ice hockey players before and after the end of growth. Am J Sports Med 41:2308– 2313. https://doi.org/10.1177/0363546513497564
- 56. Nevitt MC, Zhang Y, Javaid MK et al (2010) High systemic bone mineral density increases the risk of incident knee OA and joint space narrowing, but not radiographic progression of existing knee OA: The MOST study. Ann Rheum Dis 69:163–168. https:// doi.org/10.1136/ard.2008.099531
- Muthuri SG, McWilliams DF, Doherty M, Zhang W (2011) History of knee injuries and knee osteoarthritis: a meta-analysis of observational studies. Osteoarthr Cartil 19:1286–1293. https://doi.org/10.1016/j.joca.2011.07.015
- Blagojevic M, Jinks C, Jeffery A, Jordan KP (2010) Risk factors for onset of osteoarthritis of the knee in older adults: a systematic review and meta-analysis. Osteoarthr Cartil 18:24–33. https:// doi.org/10.1016/j.joca.2009.08.010
- Conde J, Scotece M, Gómez R et al (2011) Adipokines and osteoarthritis: novel molecules involved in the pathogenesis and progression of disease. Arthritis 2011:1–8. https://doi.org/10.1155/ 2011/203901
- Reyes C, Leyland KM, Peat G et al (2016) Association between overweight and obesity and risk of clinically diagnosed knee, hip, and hand osteoarthritis: a population-based cohort study. Arthritis Rheumatol 68:1869–1875. https://doi.org/10.1002/art. 39707
- 61. Gersing AS, Schwaiger BJ, Nevitt MC et al (2017) Is weight loss associated with less progression of changes in knee articular cartilage among obese and overweight patients as assessed with MR imaging over 48 months? Data from the osteoarthritis initiative. Radiology 284:508–520. https://doi.org/10.1148/radiol.20171 61005
- 62. Atukorala I, Makovey J, Lawler L et al (2016) Is there a dose-response relationship between weight loss and symptom improvement in persons with knee osteoarthritis? Arthritis Care Res 68:1106–1114. https://doi.org/10.1002/acr.22805
- Frey N, Hügle T, Jick SS et al (2017) Hyperlipidaemia and incident osteoarthritis of the hand: a population-based case-control study. Osteoarthr Cartil 25:1040–1045. https://doi.org/10.1016/j. joca.2017.01.014
- Garcia-Gil M, Reyes C, Ramos R et al (2017) Serum lipid levels and risk of hand osteoarthritis: the Chingford prospective cohort study. Sci Rep 7. https://doi.org/10.1038/s41598-017-03317-4
- 65. Driban JB, Lo GH, Eaton CB et al (2016) Exploratory analysis of osteoarthritis progression among medication users: data from the Osteoarthritis Initiative. Ther Adv Musculoskelet Dis 8:207–219. https://doi.org/10.1177/1759720X16664323
- 66. Lo GH, McAlindon TE, Katz JN et al (2017) Systolic and pulse pressure associate with incident knee osteoarthritis: data from the Osteoarthritis Initiative. Clin Rheumatol 36:2121–2128. https:// doi.org/10.1007/s10067-017-3656-z
- 67. Magnusson K, Bech Holte K, Juel NG et al (2017) Long term type 1 diabetes is associated with hand pain, disability and stiffness but not with structural hand osteoarthritis features - The Dialong hand study. PLoS ONE 12. https://doi.org/10.1371/journal.pone.0177118
- Frey N, Hügle T, Jick SS et al (2016) Type II diabetes mellitus and incident osteoarthritis of the hand: a population-based case– control analysis. Osteoarthr Cartil 24:1535–1540. https://doi.org/ 10.1016/j.joca.2016.04.005
- Garessus EDG, de Mutsert R, Visser AW et al (2016) No association between impaired glucose metabolism and osteoarthritis. Osteoarthr Cartil 24:1541–1547. https://doi.org/10.1016/j.joca. 2016.04.007
- 70. Wang X, Cicuttini F, Jin X et al (2017) Knee effusion-synovitis volume measurement and effects of vitamin D supplementation

in patients with knee osteoarthritis. Osteoarthr Cartil 25:1304–1312. https://doi.org/10.1016/j.joca.2017.02.804

- 71. Berenbaum F (2013) Osteoarthritis as an inflammatory disease (osteoarthritis is not osteoarthrosis!). Osteoarthr Cartil 21:16–21
- Hwang HS, Kim HA (2015) Chondrocyte apoptosis in the pathogenesis of osteoarthritis. Int J Mol Sci 16:26035–26054
- 73. Guilak F, Nims RJ, Dicks A et al (2018) Osteoarthritis as a disease of the cartilage pericellular matrix. Matrix Biol 71–72:40–50
- 74. Funck-Brentano T, Cohen-Solal M (2015) Subchondral bone and osteoarthritis. Curr Opin Rheumatol 27:420–426
- 75. Kovács B, Vajda E, Nagy EE (2019) Regulatory effects and interactions of the Wnt and OPG-RANKL-RANK signaling at the bone-cartilage interface in osteoarthritis. Int J Mol Sci 20
- Zhou X, Cao H, Yuan Y, Wu W (2020) Biochemical signals mediate the crosstalk between cartilage and bone in osteoarthritis. Biomed Res Int 2020
- Mathiessen A, Conaghan PG (2017) Synovitis in osteoarthritis: current understanding with therapeutic implications. Arthritis Res Ther 19
- Sarmanova A, Hall M, Moses J et al (2016) Synovial changes detected by ultrasound in people with knee osteoarthritis – a meta-analysis of observational studies. Osteoarthr Cartil 24:1376–1383. https://doi.org/10.1016/j.joca.2016.03.004
- Guermazi A, Hayashi D, Roemer FW et al (2014) Synovitis in knee osteoarthritis assessed by contrast-enhanced magnetic resonance imaging (MRI) is associated with radiographic tibiofemoral osteoarthritis and MRI-detected widespread cartilage damage: The MOST study. J Rheumatol 41:501–508. https://doi.org/10. 3899/jrheum.130541
- Felson DT, Niu J, Neogi T et al (2016) Synovitis and the risk of knee osteoarthritis: the MOST study. Osteoarthr Cartil 24:458– 464. https://doi.org/10.1016/j.joca.2015.09.013
- Prieto-Potin I, Largo R, Roman-Blas JA et al (2015) Characterization of multinucleated giant cells in synovium and subchondral bone in knee osteoarthritis and rheumatoid arthritis. BMC Musculoskelet Disord 16. https://doi.org/10.1186/s12891-015-0664-5
- Wojdasiewicz P, Poniatowski ŁA, Szukiewicz D (2014) The role of inflammatory and anti-inflammatory cytokines in the pathogenesis of osteoarthritis. Mediators Inflamm 2014
- 83. Klein-Wieringa IR, De Lange-Brokaar BJE, Yusuf E et al (2016) Inflammatory cells in patients with endstage knee osteoarthritis: a comparison between the synovium and the infrapatellar fat pad. J Rheumatol 43:771–778. https://doi.org/10.3899/jrheum.151068
- Kapoor M, Martel-Pelletier J, Lajeunesse D et al (2011) Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. Nat Rev Rheumatol 7:33–42
- Cai S, Ming B, Ye C et al (2021) Similar transition processes in synovial fibroblasts from rheumatoid arthritis and osteoarthritis: a single-cell study. Clin Dev Immunol 2019. https://doi.org/10. 1155/2019/4080735
- Jin X, Beguerie JR, Zhang W et al (2015) Circulating C reactive protein in osteoarthritis: a systematic review and meta-analysis. Ann Rheum Dis 74:703–710. https://doi.org/10.1136/annrheumdis-2013-204494
- 87. Stannus O, Jones G, Cicuttini F et al (2010) Circulating levels of IL-6 and TNF-α are associated with knee radiographic osteoarthritis and knee cartilage loss in older adults. Osteoarthr Cartil 18:1441–1447. https://doi.org/10.1016/j.joca.2010.08.016
- Livshits G, Zhai G, Hart DJ et al (2009) Interleukin-6 is a significant predictor of radiographic knee osteoarthritis: the Chingford study. Arthritis Rheum 60:2037–2045. https://doi.org/10.1002/ art.24598
- 89. Spector TD, Hart DJ, Nandra D et al (1997) Low-level increases in serum C-reactive protein are present in early osteoarthritis of the knee and predict progressive disease. Arthritis Rheum 40:723–727. https://doi.org/10.1002/art.1780400419

- Bulló M, Casas-Agustench P, Amigó-Correig P et al (2007) Inflammation, obesity and comorbidities: the role of diet. Public Health Nutr 10:1164–1172
- Presle N, Pottie P, Dumond H et al (2006) Differential distribution of adipokines between serum and synovial fluid in patients with osteoarthritis. Contribution of joint tissues to their articular production. Osteoarthr Cartil 14:690–695. https://doi.org/10. 1016/j.joca.2006.01.009
- 92. de Boer TN, van Spil WE, Huisman AM et al (2012) Serum adipokines in osteoarthritis; comparison with controls and relationship with local parameters of synovial inflammation and cartilage damage. Osteoarthr Cartil 20:846–853. https://doi.org/10.1016/j. joca.2012.05.002
- 93. Liu B, Gao YH, Dong N et al (2019) Differential expression of adipokines in the synovium and infrapatellar fat pad of osteoarthritis patients with and without metabolic syndrome. Connect Tissue Res 60:611–618. https://doi.org/10.1080/03008207.2019. 1620221
- 94. Tu C, He J, Wu B et al (2019) An extensive review regarding the adipokines in the pathogenesis and progression of osteoarthritis. Cytokine 113:1–12
- 95. Neumann E, Junker S, Schett G et al (2016) Adipokines in bone disease. Nat Rev Rheumatol 12:296–302
- 96. Zhao CW, Gao YH, Song WX et al (2019) An update on the emerging role of resistin on the pathogenesis of osteoarthritis. Mediators Inflamm 2019
- 97. Garten A, Schuster S, Penke M et al (2015) Physiological and pathophysiological roles of NAMPT and NAD metabolism. Nat Rev Endocrinol 11:535–546. https://doi.org/10.1038/NRENDO. 2015.117
- Travelli C, Consonni FM, Sangaletti S et al (2019) Nicotinamide phosphoribosyltransferase acts as a metabolic gate for mobilization of myeloid-derived suppressor cells. Cancer Res 79:1938– 1951. https://doi.org/10.1158/0008-5472.CAN-18-1544
- 99. Yu Q, Dong L, Li Y, Liu G (2018) SIRT1 and HIF1α signaling in metabolism and immune responses. Cancer Lett 418:20–26. https://doi.org/10.1016/J.CANLET.2017.12.035
- Dvir-Ginzberg M, Steinmeyer J (2013) Towards elucidating the role of SirT1 in osteoarthritis. Front Biosci (Landmark Ed) 18:343–355. https://doi.org/10.2741/4105
- 101. Chen C, Zhou M, Ge Y, Wang X (2020) SIRT1 and aging related signaling pathways. Mech Ageing Dev 187. https://doi.org/10. 1016/J.MAD.2020.111215
- 102. Tsai CH, Liu SC, Chung WH et al (2020) Visfatin increases VEGF-dependent angiogenesis of endothelial progenitor cells during osteoarthritis progression. Cells 9. https://doi.org/10. 3390/CELLS9051315
- Suzuki A, Yabu A, Nakamura H (2020) Advanced glycation end products in musculoskeletal system and disorders. Methods 203:179– 186. https://doi.org/10.1016/j.ymeth.2020.09.012
- 104. Xie J, Méndez JD, Méndez-Valenzuela V, Aguilar-Hernández MM (2013) Cellular signalling of the receptor for advanced glycation end products (RAGE). Cell Signal 25:2185–2197
- 105. Lambert C, Zappia J, Sanchez C et al (2021) The damage-associated molecular patterns (DAMPs) as potential targets to treat osteoarthritis: perspectives from a review of the literature. Front Med 7
- Motta F, Sica A, Selmi C (2020) Frailty in rheumatic diseases. Front Immunol 11
- 107. Fulop T, Larbi A, Pawelec G et al (2021) Immunology of aging: the birth of inflammaging. Clin Rev Allergy Immunol. https:// doi.org/10.1007/S12016-021-08899-6
- Franceschi C, Campisi J (2014) Chronic inflammation (Inflammaging) and its potential contribution to age-associated diseases. J Gerontol - Ser A Biol Sci Med Sci 69:S4–S9

- 109. Torre LA, Bray F, Siegel RL et al (2015) Global cancer statistics, 2012. CA Cancer J Clin 65:87–108. https://doi.org/10.3322/caac. 21262
- 110. Kennedy BK, Berger SL, Brunet A et al (2014) Geroscience: linking aging to chronic disease. Cell 159:709–713
- López-Otín C, Blasco MA, Partridge L et al (2013) The hallmarks of aging. Cell 153:1194
- Finkel T, Holbrook NJ (2000) Oxidants, oxidative stress and the biology of ageing. Nature 408:239–247
- 113. Reuter S, Gupta SC, Chaturvedi MM, Aggarwal BB (2010) Oxidative stress, inflammation, and cancer: how are they linked? Free Radic. Biol Med 49:1603–1616
- 114. Sosa V, Moliné T, Somoza R et al (2013) Oxidative stress and cancer: an overview. Ageing Res Rev 12:376–390
- 115. Karunakaran U, Park KG (2013) A systematic review of oxidative stress and safety of antioxidants in diabetes: focus on islets and their defense. Diabetes Metab J 37:106–112
- Pirillo A, Norata GD, Catapano AL (2013) LOX-1, OxLDL, and atherosclerosis. Mediators Inflamm. 2013
- 117. Yu W, Zhang H, Shin MR, Sesti F (2019) Oxidation of KCNB1 potassium channels in the murine brain during aging is associated with cognitive impairment. Biochem Biophys Res Commun 512:665–669. https://doi.org/10.1016/j.bbrc.2019.03.130
- Liu Z, Zhou T, Ziegler AC et al (2017) Oxidative stress in neurodegenerative diseases: from molecular mechanisms to clinical applications. Oxid Med Cell Longev 2017
- John-Schuster G, Günter S, Hager K et al (2016) Inflammaging increases susceptibility to cigarette smoke-induced COPD. Oncotarget 7:30068–30083. https://doi.org/10.18632/oncotarget. 4027
- Mateen S, Moin S, Khan AQ et al (2016) Increased reactive oxygen species formation and oxidative stress in rheumatoid arthritis. PLoS ONE 11. https://doi.org/10.1371/journal.pone. 0152925
- 121. Li Y, Goronzy JJ, Weyand CM (2018) DNA damage, metabolism and aging in pro-inflammatory T cells: Rheumatoid arthritis as a model system. Exp Gerontol 105:118–127
- 122. Franceschi C, Bonafè M, Valensin S et al (2000) Inflammaging. An evolutionary perspective on immunosenescence. In: Annals of the New York Academy of Sciences. New York Academy of Sciences, pp 244–254
- 123. Vitale G, Salvioli S, Franceschi C (2013) Oxidative stress and the ageing endocrine system. Nat Rev Endocrinol 9:228–240
- Fulop T, Witkowski JM, Olivieri F, Larbi A (2018) The integration of inflammaging in age-related diseases. Semin Immunol 40:17–35
- 125. Callender LA, Carroll EC, Beal RWJ et al (2018) Human CD8 + EMRA T cells display a senescence-associated secretory phenotype regulated by p38 MAPK. Aging Cell 17. https://doi.org/ 10.1111/acel.12675
- 126. Coppé JP, Patil CK, Rodier F et al (2008) Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. PLoS Biol 6. https://doi. org/10.1371/journal.pbio.0060301
- Coppé JP, Desprez PY, Krtolica A, Campisi J (2010) The senescenceassociated secretory phenotype: the dark side of tumor suppression. Annu Rev Pathol Mech Dis 5:99–118
- Bleve A, Motta F, Durante B et al (2022) Immunosenescence, inflammaging, and frailty: role of myeloid cells in age-related diseases. Clin Rev Allergy Immunol. https://doi.org/10.1007/ S12016-021-08909-7
- Coder BD, Wang H, Ruan L, Su D-M (2015) Thymic involution perturbs negative selection leading to autoreactive t cells that induce chronic inflammation. J Immunol 194:5825–5837. https:// doi.org/10.4049/jimmunol.1500082

- Coder B, Su DM (2015) Thymic involution beyond T-cell insufficiency. Oncotarget 6:21777–21778
- Brunner S, Herndler-Brandstetter D, Weinberger B, Grubeck-Loebenstein B (2011) Persistent viral infections and immune aging. Ageing Res Rev 10:362–369
- Ebersole JL, Graves CL, Gonzalez OA et al (2000) (2016) Aging, inflammation, immunity and periodontal disease. Periodontol 72:54–75
- 133. Franceschi C, Garagnani P, Vitale G et al (2017) Inflammaging and 'Garb-aging.' Trends Endocrinol Metab 28:199–212
- 134. Lee B-J, Min C-K, Hancock M et al (2021) Human cytomegalovirus host interactions: EGFR and host cell signaling is a point of convergence between viral infection and functional changes in infected cells. Front Microbiol 12:660901. https://doi.org/10. 3389/fmicb.2021.660901
- Lohr JM, Oldstone MBA (1990) Detection of cytomegalovirus nucleic acid sequences in pancreas in type 2 diabetes. Lancet 336:644–648. https://doi.org/10.1016/0140-6736(90)92145-8
- Donath MY, Shoelson SE (2011) Type 2 diabetes as an inflammatory disease. Nat Rev Immunol 11:98–107
- Low H, Mukhamedova N, Cui HL et al (2016) Cytomegalovirus restructures lipid rafts via a US28/CDC42-mediated pathway, enhancing cholesterol efflux from host cells. Cell Rep 16:186– 200. https://doi.org/10.1016/j.celrep.2016.05.070
- Yu Y, Clippinger AJ, Alwine JC (2011) Viral effects on metabolism: changes in glucose and glutamine utilization during human cytomegalovirus infection. Trends Microbiol 19:360–367
- Hotamisligil GS (2006) Inflammation and metabolic disorders. Nature 444:860–867
- Ye J, Keller JN (2010) Regulation of energy metabolism by inflammation: a feedback response in obesity and calorie restriction. Aging (Albany NY) 2:361–368. https://doi.org/10.18632/aging.100155
- 141. Collino S, Montoliu I, Martin F-PJ et al (2013) Correction: metabolic signatures of extreme longevity in Northern Italian centenarians reveal a complex remodeling of lipids, amino acids, and gut microbiota metabolism. PLoS ONE 8. https://doi.org/10. 1371/annotation/5fb9fa6f-4889-4407-8430-6dfc7ecdfbdd
- Biagi E, Nylund L, Candela M et al (2010) Through ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians. PLoS ONE 5. https://doi.org/10.1371/journal.pone. 0010667
- Biagi E, Candela M, Franceschi C, Brigidi P (2011) The aging gut microbiota: new perspectives. Ageing Res Rev 10:428–429
- Cevenini E, Monti D, Franceschi C (2013) Inflamm-ageing. Curr Opin Clin Nutr Metab Care 16:14–20
- 145. Biagi E, Franceschi C, Rampelli S et al (2016) Gut microbiota and extreme longevity. Curr Biol 26:1480–1485. https://doi.org/ 10.1016/j.cub.2016.04.016
- 146. Franceschi C, Salvioli S, Garagnani P et al (2017) Immunobiography and the heterogeneity of immune responses in the elderly: a focus on inflammaging and trained immunity. Front Immunol 8
- 147. Santoro A, Ostan R, Candela M et al (2018) Gut microbiota changes in the extreme decades of human life: a focus on centenarians. Cell Mol Life Sci 75:129–148
- Kundu P, Blacher E, Elinav E, Pettersson S (2017) Our gut microbiome: the evolving inner self. Cell 171:1481–1493
- 149. Lee C, Longo V (2016) Dietary restriction with and without caloric restriction for healthy aging. F1000Research 5
- Barzilai N, Huffman DM, Muzumdar RH, Bartke A (2012) The critical role of metabolic pathways in aging. Diabetes 61:1315–1322
- 151. Ristow M, Schmeisser K (2014) Mitohormesis: promoting health and lifespan by increased levels of reactive oxygen species (ROS). Dose-Response 12:288–341. https://doi.org/10.2203/ dose-response.13-035.Ristow
- 152. Das SK, Balasubramanian P, Weerasekara YK (2017) Nutrition modulation of human aging: the calorie restriction paradigm.

🖄 Springer

Mol Cell Endocrinol 455:148–157. https://doi.org/10.1016/j. mce.2017.04.011

- Mirzaei H, Suarez JA, Longo VD (2014) Protein and amino acid restriction, aging and disease: from yeast to humans. Trends Endocrinol Metab 25:558–566
- 154. Loeser RF, Olex AL, McNulty MA et al (2012) Microarray analysis reveals age-related differences in gene expression during the development of osteoarthritis in mice. Arthritis Rheum 64:705–717. https://doi.org/10.1002/ART.33388
- 155. Long D, Blake S, Song XY et al (2008) Human articular chondrocytes produce IL-7 and respond to IL-7 with increased production of matrix metalloproteinase-13. Arthritis Res Ther 10. https://doi. org/10.1186/AR2376
- 156. Rezuş E, Cardoneanu A, Burlui A et al (2019) The link between inflammaging and degenerative joint diseases. Int J Mol Sci 20. https://doi.org/10.3390/IJMS20030614
- Millerand M, Berenbaum F, Jacques C (2019) Danger signals and inflammaging in osteoarthritis. Clin Exp Rheumatol 37:48–56
- 158. Goekoop RJ, Kloppenburg M, Kroon HM et al (2010) Low innate production of interleukin-1β and interleukin-6 is associated with the absence of osteoarthritis in old age. Osteoarthr Cartil 18:942– 947. https://doi.org/10.1016/j.joca.2010.03.016
- 159. Ni Z, Kuang L, Chen H et al (2019) The exosome-like vesicles from osteoarthritic chondrocyte enhanced mature IL-1β production of macrophages and aggravated synovitis in osteoarthritis. Cell Death Dis 10. https://doi.org/10.1038/s41419-019-1739-2
- 160. Kato T, Miyaki S, Ishitobi H et al (2014) Exosomes from IL-1β stimulated synovial fibroblasts induce osteoarthritic changes in articular chondrocytes. Arthritis Res Ther 16. https://doi.org/10. 1186/ar4679
- 161. Chien SY, Tsai CH, Liu SC et al (2020) Noggin inhibits IL-1β and BMP-2 expression, and attenuates cartilage degeneration and subchondral bone destruction in experimental osteoarthritis. Cells 9. https://doi.org/10.3390/cells9040927
- 162. Nasi S, So A, Combes C et al (2016) Interleukin-6 and chondrocyte mineralisation act in tandem to promote experimental osteoarthritis. Ann Rheum Dis 75:1372–1379. https://doi.org/ 10.1136/annrheumdis-2015-207487
- Loeser RF, Collins JA, Diekman BO (2016) Ageing and the pathogenesis of osteoarthritis. Nat Rev Rheumatol 12:412–420
- Jeon H, Il IG (2017) Autophagy in osteoarthritis. Connect Tissue Res 58:497–508. https://doi.org/10.1080/03008207.2016.1240790
- Gao T, Guo W, Chen M et al (2016) Extracellular vesicles and autophagy in osteoarthritis. Biomed Res Int 2016. https://doi.org/ 10.1155/2016/2428915
- Ponchel F, Burska AN, Hensor EMA et al (2015) Changes in peripheral blood immune cell composition in osteoarthritis. Osteoarthr Cartil 23:1870–1878. https://doi.org/10.1016/j.joca.2015.06.018
- Zhu W, Zhang X, Jiang Y et al (2020) Alterations in peripheral T cell and B cell subsets in patients with osteoarthritis. Clin Rheumatol 39:523–532. https://doi.org/10.1007/s10067-019-04768-y
- 168. Shan Y, Qi C, Liu Y et al (2017) Increased frequency of peripheral blood follicular helper T cells and elevated serum IL-21 levels in patients with knee osteoarthritis. Mol Med Rep 15:1095–1102. https://doi.org/10.3892/mmr.2017.6132
- 169. de Lange-Brokaar BJE, Ioan-Facsinay A, van Osch GJVM et al (2012) Synovial inflammation, immune cells and their cytokines in osteoarthritis: a review. Osteoarthr Cartil 20:1484–1499. https://doi.org/10.1016/J.JOCA.2012.08.027
- 170. Pessler F, Chen LX, Dai L et al (2008) A histomorphometric analysis of synovial biopsies from individuals with Gulf War Veterans' illness and joint pain compared to normal and osteoarthritis synovium. Clin Rheumatol 27:1127–1134. https://doi. org/10.1007/S10067-008-0878-0
- 171. Mikolajczyk TP, Nosalski R, Szczepaniak P et al (2016) Role of chemokine RANTES in the regulation of perivascular

inflammation, T-cell accumulation, and vascular dysfunction in hypertension. FASEB J 30:1987–1999. https://doi.org/10.1096/fj. 201500088R

- 172. Lopes EBP, Filiberti A, Husain SA, Humphrey MB (2017) Immune contributions to osteoarthritis. Curr Osteoporos Rep 15:593–600. https://doi.org/10.1007/S11914-017-0411-Y
- 173. Siebuhr AS, Bay-Jensen AC, Jordan JM et al (2016) Inflammation (or synovitis)-driven osteoarthritis: an opportunity for personalizing prognosis and treatment? Scand J Rheumatol 45:87–98
- 174. Fried LP, Tangen CM, Walston J et al (2001) Frailty in older adults: evidence for a phenotype. J Gerontol - Ser A Biol Sci Med Sci 56. https://doi.org/10.1093/gerona/56.3.m146
- Kojima G, Liljas AEM, Iliffe S (2019) Frailty syndrome: implications and challenges for health care policy. Risk Manag Healthc Policy 12:23–30
- Ferrucci L, Fabbri E (2018) Inflammageing: chronic inflammation in ageing, cardiovascular disease, and frailty. Nat Rev Cardiol 15:505–522
- Motta F, Sica A, Selmi C (2020) Frailty in rheumatic diseases. Front Immunol 11. https://doi.org/10.3389/FIMMU.2020.576134
- Cacciatore F, Della-morte D, Basile C et al (2014) Long-term mortality in frail elderly subjects with osteoarthritis. Rheumatology (Oxford) 53:293–299. https://doi.org/10.1093/RHEUMATOLOGY/KET348
- 179. Chen P, Huang L, Ma Y et al (2019) Intra-articular plateletrich plasma injection for knee osteoarthritis: a summary of meta-analyses. J Orthop Surg Res 14. https://doi.org/10.1186/ s13018-019-1363-y
- Le ADK, Enweze L, DeBaun MR, Dragoo JL (2019) Platelet-rich plasma. Clin Sports Med 38:17–44. https://doi.org/10.1016/J. CSM.2018.08.001
- Spreafico A, Chellini F, Frediani B et al (2009) Biochemical investigation of the effects of human platelet releasates on human articular chondrocytes. J Cell Biochem 108:1153–1165. https:// doi.org/10.1002/JCB.22344
- Smyth NA, Murawski CD, Fortier LA et al (2013) Platelet-rich plasma in the pathologic processes of cartilage: review of basic science evidence. Arthroscopy 29:1399–1409. https://doi.org/10. 1016/J.ARTHRO.2013.03.004
- Battaglia M, Guaraldi F, Vannini F et al (2013) Efficacy of ultrasoundguided intra-articular injections of platelet-rich plasma versus hyaluronic acid for hip osteoarthritis. Orthopedics 36. https://doi.org/10. 3928/01477447-20131120-13
- 184. Dallari D, Stagni C, Rani N et al (2016) Ultrasound-guided injection of platelet-rich plasma and hyaluronic acid, separately and in combination, for hip osteoarthritis: a randomized controlled study. Am J Sports Med 44:664–671. https://doi.org/10.1177/ 0363546515620383
- 185. Doria C, Mosele GR, Caggiari G et al (2017) Treatment of early hip osteoarthritis: ultrasound-guided platelet rich plasma versus hyaluronic acid injections in a randomized clinical trial. Joints 5:152–155. https://doi.org/10.1055/S-0037-1605584
- Di Sante L, Villani C, Santilli V et al (2016) Intra-articular hyaluronic acid vs platelet-rich plasma in the treatment of hip osteoarthritis. Med Ultrason 18:463–468. https://doi.org/10.11152/ MU-874
- Hamilton JA, Cook AD, Tak PP (2016) Anti-colony-stimulating factor therapies for inflammatory and autoimmune diseases. Nat Rev Drug Discov 16:53–70. https://doi.org/10.1038/NRD.2016.231
- Lee KMC, Prasad V, Achuthan A et al (2020) Targeting GM-CSF for collagenase-induced osteoarthritis pain and disease in mice. Osteoarthr Cartil 28:486–491. https://doi.org/10.1016/j. joca.2020.01.012
- 189. Cook AD, Pobjoy J, Steidl S et al (2012) Granulocyte-macrophage colony-stimulating factor is a key mediator in experimental osteoarthritis pain and disease development. Arthritis Res Ther 14. https://doi.org/10.1186/AR4037

- 190. Steen-Louws C, Popov-Celeketic J, Mastbergen SC et al (2018) IL4-10 fusion protein has chondroprotective, anti-inflammatory and potentially analgesic effects in the treatment of osteoarthritis. Osteoarthr Cartil 26:1127–1135. https://doi.org/10.1016/j.joca. 2018.05.005
- 191. van Helvoort EM, de Visser HM, Lafeber FPJG et al (2021) IL4-10 fusion protein shows DMOAD activity in a rat osteoarthritis model. Cartilage 13:1155S-1164S. https://doi.org/10. 1177/19476035211026736
- 192. Hwang HS, Park IY, Choi SY, Kim HA (2017) PEP-1-GRX-1 modulates matrix metalloproteinase-13 and nitric oxide expression of human articular chondrocytes. Cell Physiol Biochem 41:252–264. https://doi.org/10.1159/000456090
- 193. Bin ZH, Zhang Y, Chen C et al (2016) Pioglitazone inhibits advanced glycation end product-induced matrix metalloproteinases and apoptosis by suppressing the activation of MAPK and NF-κB. Apoptosis 21:1082–1093. https://doi.org/10.1007/ s10495-016-1280-z
- 194. Campo GM, Avenoso A, D'Ascola A et al (2012) Hyaluronan differently modulates TLR-4 and the inflammatory response in mouse chondrocytes. BioFactors 38:69–76. https://doi.org/10. 1002/biof.202
- 195. Li Y, Zhang Y, Chen C et al (2016) Establishment of a rabbit model to study the influence of advanced glycation end products accumulation on osteoarthritis and the protective effect of pioglitazone. Osteoarthr Cartil 24:307–314. https://doi.org/10.1016/J.JOCA. 2015.08.001
- 196. Boileau C, Martel-Pelletier J, Fahmi H et al (2007) The peroxisome proliferator-activated receptor gamma agonist pioglitazone reduces the development of cartilage lesions in an experimental dog model of osteoarthritis: in vivo protective effects mediated through the inhibition of key signaling and catabolic pathways. Arthritis Rheum 56:2288–2298. https://doi.org/10.1002/ART.22726
- 197. Kobayashi T, Notoya K, Naito T et al (2005) Pioglitazone, a peroxisome proliferator-activated receptor gamma agonist, reduces the progression of experimental osteoarthritis in guinea pigs. Arthritis Rheum 52:479–487. https://doi.org/10.1002/ART.20792
- 198. Chayanupatkul M, Honsawek S (2010) Soluble receptor for advanced glycation end products (sRAGE) in plasma and synovial fluid is inversely associated with disease severity of knee osteoarthritis. Clin Biochem 43:1133–1137. https://doi.org/10.1016/j. clinbiochem.2010.07.007
- 199. Peng Y, Park HS, Tang LA et al (2019) Generation of sRAGE high transgenic mice to study inflammaging. Front Biosci - Landmark 24:555–563. https://doi.org/10.2741/4735
- Luo Y, Li J, Wang B et al (2021) Protective effect of glycyrrhizin on osteoarthritis cartilage degeneration and inflammation response in a rat model. J Bioenerg Biomembr 53:285–293. https://doi.org/ 10.1007/S10863-021-09889-1
- 201. Olcum M, Tufekci KU, Durur DY et al (2021) Ethyl Ethyl pyruvate attenuates microglial NLRP3 inflammasome activation via inhibition of HMGB1/NF-κB/miR-223 signaling. Antioxidants (Basel, Switzerland) 10. https://doi.org/10.3390/ANTIOX10050745
- Luo Y, Li J, Wang B et al (2021) Protective effect of glycyrrhizin on osteoarthritis cartilage degeneration and inflammation response in a rat model. J Bioenerg Biomembr 53. https://doi.org/10.1007/ s10863-021-09889-1
- Li S, Liang F, Kwan K et al (2018) Identification of ethyl pyruvate as a NLRP3 inflammasome inhibitor that preserves mitochondrial integrity. Mol Med 24. https://doi.org/10.1186/s10020-018-0006-9
- Xue J, Suarez JS, Minaai M et al (2021) HMGB1 as a therapeutic target in disease. J Cell Physiol 236:3406–3419
- Aulin C, Lassacher T, Palmblad K, Erlandsson Harris H (2020) Early stage blockade of the alarmin HMGB1 reduces cartilage destruction in experimental OA. Osteoarthr Cartil 28:698–707. https://doi.org/10.1016/j.joca.2020.01.003

- 206. Schelbergen RF, Geven EJ, Van Den Bosch MHJ et al (2015) Prophylactic treatment with S100A9 inhibitor paquinimod reduces pathology in experimental collagenase-induced osteoarthritis. Ann Rheum Dis 74:2254–2258. https://doi.org/10.1136/ annrheumdis-2014-206517
- 207. Van Den Bosch MH, Blom AB, Schelbergen RF et al (2016) Alarmin S100A9 induces proinflammatory and catabolic effects predominantly in the M1 macrophages of human osteoarthritic synovium. J Rheumatol 43:1874–1884. https://doi.org/10.3899/ jrheum.160270
- van den Bosch MHJ (2019) Inflammation in osteoarthritis: is it time to dampen the alarm(in) in this debilitating disease? Clin Exp Immunol 195:153–166
- 209. Cremers NAJ, van den Bosch MHJ, van Dalen S et al (2017) S100A8/ A9 increases the mobilization of pro-inflammatory Ly6Chigh monocytes to the synovium during experimental osteoarthritis. Arthritis Res Ther 19. https://doi.org/10.1186/s13075-017-1426-6
- 210. Jeon OH, Kim C, Laberge RM et al (2017) Local clearance of senescent cells attenuates the development of post-traumatic osteoarthritis and creates a pro-regenerative environment. Nat Med 23:775–781. https://doi.org/10.1038/nm.4324
- 211. Bay-Jensen AC, Mobasheri A, Thudium CS et al (2022) Blood and urine biomarkers in osteoarthritis - an update on cartilage associated type II collagen and aggrecan markers. Curr Opin Rheumatol 34:54–60. https://doi.org/10.1097/BOR.00000000000845
- 212. Kraus VB, Collins JE, Hargrove D et al (2017) Predictive validity of biochemical biomarkers in knee osteoarthritis: data from the FNIH OA Biomarkers Consortium. Ann Rheum Dis 76:186–195. https://doi.org/10.1136/ANNRHEUMDIS-2016-209252
- Luo Y, He Y, Reker D et al (2018) A novel high sensitivity type II collagen blood-based biomarker, PRO-C2, for assessment of cartilage formation. Int J Mol Sci 19. https://doi.org/10.3390/IJMS19113485
- 214. Siebuhr AS, Bay-Jensen AC, Leeming DJ et al (2013) Serological identification of fast progressors of structural damage with rheumatoid arthritis. Arthritis Res Ther 15. https://doi.org/10.1186/ AR4266
- Goldring MB, Goldring SR (2010) Articular cartilage and subchondral bone in the pathogenesis of osteoarthritis. Ann N Y Acad Sci 1192:230–237. https://doi.org/10.1111/J.1749-6632.2009.05240.X
- 216. Huebner JL, Bay-Jensen AC, Huffman KM et al (2014) Alpha C-telopeptide of type I collagen is associated with subchondral bone turnover and predicts progression of joint space narrowing

and osteophytes in osteoarthritis. Arthritis Rheumatol (Hoboken, NJ) 66:2440–2449. https://doi.org/10.1002/ART.38739

- 217. Engbersen M, Huang ZKV (2016) Bone biomarkers related to osteoarthritis. In: Preedy V (ed) Biomarkers in disease: methods, discoveries and applications. Dordrecht
- Haraden CA, Huebner JL, Hsueh MF et al (2019) Synovial fluid biomarkers associated with osteoarthritis severity reflect macrophage and neutrophil related inflammation. Arthritis Res Ther 21. https://doi.org/10.1186/S13075-019-1923-X
- Hsueh MF, Zhang X, Wellman SS et al (2021) Synergistic roles of macrophages and neutrophils in osteoarthritis progression. Arthritis Rheumatol (Hoboken, NJ) 73:89–99. https://doi.org/10.1002/ART. 41486
- 220. Sunahori K, Yamamura M, Yamana J et al (2006) The S100A8/ A9 heterodimer amplifies proinflammatory cytokine production by macrophages via activation of nuclear factor kappa B and p38 mitogen-activated protein kinase in rheumatoid arthritis. Arthritis Res Ther 8. https://doi.org/10.1186/AR1939
- 221. Van Lent PLEM, Blom AB, Schelbergen RFP et al (2012) Active involvement of alarmins S100A8 and S100A9 in the regulation of synovial activation and joint destruction during mouse and human osteoarthritis. Arthritis Rheum 64:1466–1476. https://doi.org/10. 1002/ART.34315
- 222. Swindell WR, Johnston A, Xing X et al (2013) Robust shifts in S100a9 expression with aging: a novel mechanism for chronic inflammation. Sci Rep 3. https://doi.org/10.1038/SREP01215
- 223. Gerss J, Roth J, Holzinger D et al (2012) Phagocyte-specific S100 proteins and high-sensitivity C reactive protein as biomarkers for a risk-adapted treatment to maintain remission in juvenile idiopathic arthritis: a comparative study. Ann Rheum Dis 71:1991–1997. https://doi.org/10.1136/ANNRHEUMDIS-2012-201329
- 224. Choi IY, Gerlag DM, Herenius MJ et al (2015) MRP8/14 serum levels as a strong predictor of response to biological treatments in patients with rheumatoid arthritis. Ann Rheum Dis 74:499–505. https://doi.org/10.1136/ANNRHEUMDIS-2013-203923
- 225. Holzinger D, Nippe N, Vogl T et al (2014) Myeloid-related proteins 8 and 14 contribute to monosodium urate monohydrate crystalinduced inflammation in gout. Arthritis Rheumatol (Hoboken, NJ) 66:1327–1339. https://doi.org/10.1002/ART.38369

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.